



**University of Babylon
College of Medicine**

A Bacteriological Study of Maxillary Sinusitis in Babylon Province

A Thesis

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of Babylon In Partial Fulfillment of the Requirements
For the Degree of Master of Science
In Medical Microbiology**

By

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**Dhul -
Hijja/١٤٢٧**

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

((قَالُوا سُبْحَانَكَ لَا عِلْمَ لَنَا إِلَّا مَا عَلَّمْتَنَا

إِنَّكَ أَنْتَ الْعَلِيمُ الْحَكِيمُ))

حَسْبُكَ اللَّهُ الْعَلِيُّ الْعَظِيمُ

سورة البقرة
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DEDICATION

To

Holy Imam Al- Hujja...

Mohammed Al-Mahdi....

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Abbreviations

Symbol	Description
DW	Distilled water
ENT	Ear nose throat
MRSA	Methecilin Resistant <i>Staphylococcus aureus</i>
MSSA	Methecilin Sensetive <i>Staphylococcus aureus</i>
NCCLs	National Committee for Clinical Laboratory Standards
URTI	Upper respiratory tract infection

Abstract

A total of 100 samples were collected from 100 patients (60 of them were females and 40 were males) who were referred to Hilla Teaching Hospital and special clinics during a period of ten months (from October 2008 to July 2009). The collected samples were investigated for bacterial analysis. These patients were suffering from sinusitis (68 cases were acute sinusitis and other 32 cases were chronic) as being diagnosed by ENT experts (through clinical and radiological examination X-ray and / or CT scan).

The ages of those patients ranged from 10 to 60 years. The results indicated that, the most age group being susceptible for both acute and chronic sinusitis was between 20 to 29 years old.

The results of bacterial culture for acute sinusitis were Positive in 68 patients (68.0%) versus 32 (32.0%) revealed negative bacterial culture, while for chronic sinusitis 40 patients (40.0%) had positive culture and 60 patients (60.0%) versus negative culture. The most common type of bacteria which was isolated from acute sinusitis was *Streptococcus pneumoniae* (20%) followed by *Moraxella catarrhalis* (18%), *Staphylococcus aureus* (16.7%), Coagulase negative staphylococci (14%), Viridans group (12%), *Klebsiella pneumoniae* (8%) and *Streptococcus pyogenes* (6%).

In chronic sinusitis the dominant bacterial type was *Staphylococcus aureus* (22.5%) followed by *Streptococcus pneumoniae* (23.5%), *Moraxella catarrhalis* (10.5%), *Klebsiella pneumoniae* (9%), *E.coli* (6.5%), Viridans group (6.5%) and *Proteus mirabilis* (3.5%).

The Methicillin Resistance *Staphylococcus aureus* (MRSA) was detected in both acute and chronic sinusitis which are accounted for 14% and 37% respectively.

The potency (Ampicillin, Amoxicillin, Gentamycin, Cefotaxim, Cefalothin, Ciprofloxacin, Erythromycin, Tetracyclin, Vancomycin, Trimethoprim) of antibiotics against bacterial isolates in this study was investigated. The results of this study showed that ciprofloxacin and cefotaxim were the most effective antibiotics towards the tested bacterial isolates.

Chapter One

Introduction and Literature Review

1-1 Introduction:

Sinusitis is one of the most common infections of humans respiratory tract (Gwaltney, 1999). The lining of the nose and paranasal sinuses is continuous and inflammation which affects the lining of the nose will spread to variable extent, into the sinuses (Buchem *et.al.*, 1990).

Sinusitis generally develops as a complication of viral or allergic inflammation of the upper respiratory tract, because the inflamed nasal mucosa block ostial draining from the sinus and the result in stasis in the sinuses encourage development of infection (Calhoun, 1992).

Acute sinusitis is a short-term condition that responds well to antibiotic and decongestant. Patients with acute sinusitis may be difficult to diagnose on clinical ground only, since symptoms includes; malaise, fever, purulent discharge, tenderness and facial pain (Ferguson, 1990).

Chronic sinusitis is more prolonged than acute sinusitis and it is characterized by at least four recurrences of acute sinusitis, either medication or surgery is a possible treatment for that. Symptoms of chronic sinusitis include facial pain, purulent discharge, rhinorrhea and malaise (Finegold *et.al.*, 2002). It is important to differentiate between acute and chronic sinusitis on the bases of the duration of symptoms because their etiologies and treatment may differ (Snow *et.al.*, 2001).

Early identification of sinusitis is medically important because the infection in such closed space carries a risk of complication and may warrant on active therapeutic approach including drainage of infected secretion and administration of antimicrobial agent (Lundberg *et.al.*, 1979 and Stankiewicz *et.al.*, 1993). Radiological examination by CT-

scan is the most reliable method of diagnosing of maxillary sinusitis (Lund and Lloyd, 1983).

Aspiration is important for the demonstration of secretion in the sinus and for the identification of the specific etiology by culture, which may be useful as a guide for antimicrobial therapy (Bridger, 1980; Axelsson and Brorson, 1973 and Little *et.al.*, 1998).

The etiology of acute sinusitis is primarily bacterial *S. pneumoniae* , *H. influenzae* and *S. aureus* are the main cause and present over half of the cases (Kortekangas, 1964; Lystad *et.al.*, 1964; Mann *et.al.*, 1982; Berg *et.al.*, 1988 and Diaz and Bamberger, 1990). Lately *M. catarrhalis* has also been implicated as a pathogen in sinusitis (Wald *et.al.*, 1984; Chapman *et.al.*, 1980), *S. pyogenes* is relatively uncommon pathogen in acute maxillary sinusitis, with frequencies 1 and /or 3 % (Wald *et.al.*, 1981; Gwaltney *et.al.*, 1981). In chronic sinusitis mostly the same type of bacteria with anaerobic bacteria like (*Bacteriodes*, *Peptostreptococcus*, *Fusobacterium* and *Viellonella*) is well established, but the occurrence of anaerobe bacteria in acute sinusitis is less clear (Radosz *et.al.*, 1997; Liu *et.al.*, 1998).

This study was suggested in an attempt to fulfill the following goals :-

- 1-Detection of the bacterial causes of sinusitis.
- 2-Determination of the environmental factors related with sinusitis.
- 3-Determination of some risk factors associated with the infection.
- 4-Determination of the susceptibility of isolated bacteria to traditional antibiotics.

1-2 Literature review:

In this chapter light will be thrown upon the sinuses through the summarized themes below:-

1-2-1 Embryology:-

The maxillary sinus is present at birth, undergoing further expansion with age. Two main points of rapid growth occur from birth until the age of 3 Years, and then from the age of 5 years through early adolescence. The sinus then slowly grows until it reaches adult size by the age of 18 years (Maresh, 1989).

Later in adult life, the sinus continues to pneumatize inferiorly and may expand to contain tooth root. The floor of the maxillary sinus is below the level of the nasal floor (Hengerer, 1984).

1-2-2 Anatomy of the Paranasal sinuses :

There are five Paranasal sinuses (air – filled spaces lined by mucous membrane) on each side of the head. They are ethmoid, maxillary, frontal and sphenoid sinuses (Lazar, 1993). Sinuses are divided into two groups, the anterior group and the posterior group. The anterior group comprises the frontal, maxillary and anterior ethmoidal sinuses while the posterior group is comprised of sphenoid and the posterior ethmoid sinuses. They are lined with a pseudostratified columnar (respiratory epithelium) which is continuous with the nasal mucosa (Baily *et.al.*, 1993).

The lining provides a mucous secretion which traps bacteria and subsequently extrudes the mucous and bacteria through the sinus ostia to be swallowed or expectorated. Under normal conditions, the sinuses are air filled and communicate directly with the nasal passages through patent ostia (Chow, 1998).

1-2-3 The Maxillary Sinus:

The maxillary sinus occupies the body of the maxilla and is the largest paranasal sinuses, which has pyramidal shape. The maxillary sinus cavity has a volume of 10 to 30 ml; the two sinuses are usually equal in size so rarely one sinus is completely absent (Geurkink, 1983).

There is a relationship between the teeth and floor of sinus where the root of first, second and the 3rd premolar and sometimes the root of canine project up into sinus, hence root infection of these teeth is the frequent cause of maxillary sinusitis. The maxillary sinus is lined by ciliated columnar epithelium and covered with a mucus blanket; this epithelium is well supplied with goblet cell (Jack and Gwaltney, 2000).

1-2-4 Physiology of Paranasal sinuses : -

There are many theories about the function of these sinuses, but there is general agreement to their specific functional role: - (Courtiss *et.al.*, 1984)

- 1- Air conditioning.
- 2- Aid to balance the head.
- 3- Vocal resonance.
- 4- Thermal insulators.

Moreover sinuses have defensive functions for the respiratory tract represented by:

1-Local defensive functions include specific immunoglobulin mechanisms with secretory IgA, IgG and lymphocyte. Non specific defensive functions reflexes coughing, sneezing mucocilliary system, nasal hair epithelial integrity, lining fluid and antimicrobial substances in lining fluid.

2-Systemic mechanisms which include specific serum IgA, IgE, and lymphocytes. And non specific serum factors opsonin, complement and mononuclear phagocytes (Dark-Lee, 1997).

1-2-5 Infection of Paranasal Sinuses:

Since the lining of the nose and paranasal sinuses continuous, the inflammation which affects the lining of the nose will spread to variable extent into the sinuses (Norrby, 1983).

The inflammation process that is primarily present in the sinuses will in turn extend to the nasal cavity and result in a variable amount of rhinitis; most conditions of the nose therefore affect both the nasal cavity and the sinuses (rhino sinusitis) (Peter, 2000).

rhinosinusitis could be classified into allergic and non-allergic, and the later being infective and non infective has advantage of simplicity, but masking the fact that rhinosinusitis are multifactor etiology with considerable overlap of clinical manifestation (Macky and Bull, 1997).

There is a key element which is used for the diagnosis of sinusitis which consist of two groups of symptoms:-

1-The first group is called major symptoms (Facial pain –pressure, facial congestion – fullness, nasal obstruction, nasal discharge –purulent or discolored postnasal drainage, hyposominal-anosmia, fever in acute sinusitis).

2-The second group is called minor symptoms (headaches, Halitosis, fatigue, dental pain, cough). In general terms the diagnosis of sinusitis = 2 or 1 major symptom + 2 minor symptoms (Evans, 1998).

Sinusitis can be classified into two groups; acute and chronic sinusitis; this classification depends on the period of the infection and symptom (Pool, 1999).

Acute sinusitis: - Bacterial infection of Paranasal sinuses lasting less than 30 days in which symptoms completely are resolved.

Chronic sinusitis: - Episodes of inflammation of the Paranasal sinuses are lasting more than 90 days. Patients have a persistent residual obstruction (Spector *et.al.*, 1998)

1-2-5-1 Acute sinusitis:-

Acute bacterial sinusitis usually occurs following an upper respiratory tract infection that result in obstruction of osteomeatal complex, impaired mucociliary clearance and over production of fluid secretions (Gooch, 1998). Acute infection may be suppurative or non - suppurative. It is one of the most common infections of humans (Osguthorpe and Hadley, 1999).

In general it develops as a complication of viral or allergic inflammation of the upper respiratory tract (Brook, 1996). Low *et.al.*, (1997) classified acute sinusitis according to its microbial etiology (viral, bacterial, parasitic). The distinguishing of acute viral rhinosinusitis of cold

influenza like illness from cases with secondary bacterial infection is a frequent challenge to the physicians in primary care.

The distinguishing is important because antibiotics should be administered to patients with acute bacterial sinusitis but not to those with viral rhino sinusitis (Dykewicz, २००३).

A- Pathology:

The changes in the mucous membrane in acute sinusitis are those of acute inflammation in nasal tissue increased blood supply with outpouring of serum and polymorphonuclear leukocytes associated with local swelling, redness and edema which is caused by obstruction of veins and lymphatic duct (Davidson *et.al.*, १९८९). If the obstruction and edema persist, cell degeneration occurs with cloudy swelling and necrotic changes will take place if interference with circulation becomes prolonged (Stamberger, १९८६).

Clinically, inflammation of the sinus may be either catarrhal or suppurative; catarrhal inflammation is characterized by hyperemia and excess secretion of mucous, but with few leukocytes and little or no destruction of the mucous membrane (Krishna, २००६).

If the inflammatory reaction is more severe there will be more extensive exudation of leukocyte together with the mucous membrane or with severe infection; the discharge becomes increasingly purulent and less mucoid until, with necrotic mucous membrane, pus alone will be found although all combinations of this catarrhal condition and suppurative sinusitis are found (Wald, १९९८).

It is an undoubted and important fact that in some cases the acute inflammation quickly produces a suppurative sinusitis and this is particularly true in the maxillary sinus with dental infection. The opposite

case is one in which the acute inflammation is accompanied by a marked swelling and edema of the sinus mucous membrane and in which hyperplasia gives rise to the most prominent sign, moreover the mucous gland becomes hypertrophied and secretes unusual thick and tenacious mucous and the membrane may become so thick as to fill the sinus cavity completely. Polypoid mucous membrane or polypi is often present (Mackay and Bull, 1997).

B- Pathogenesis:-

The most common sequence of events leading to infection community acquired is first a viral rhino sinusitis to be complicated by a second bacterial infection in sinus cavity (Gwaltney, 1996).

Sinusitis can be the result of an alteration in ostial size, mucociliary transport, oxygen exchange or mucosal blood flow; so the most important factor in the pathogenesis of sinusitis is narrowing of the ostia, either by mucosal inflammation or an anatomic abnormality (Wagenmann and Naclerio, 1992).

Development of purulence may decrease the oxygen tension to near zero, thus favoring growth of anaerobic bacteria and inhibiting the function of the phagocytic cell (Robert, 1991).

1-2-5-2 Chronic sinusitis :

Chronic sinusitis is a common illness. In many cases it follows an incompletely resolved acute sinusitis, but it may appear insidiously following a cold or a tooth infection (Lanza and Kennedy, 1992). There are some factors contributing to sinusitis pathogenesis including anatomic factors, disturbances in mucociliary clearance, microbial pathogen and inflammatory factors (Hamilos, 2000).

A- Pathophysiology:-

Wald (1998) showed that patients typically experience several episodes of acute sinusitis before developing chronic sinusitis; the resistance of the maxillary ostium is more pronounced in cases of chronic sinusitis than in the normal or acutely inflamed antrum.

The common cold and other respiratory viral syndromes may in turn lead to sinusitis. More commonly they lead to swelling of the lining of the nose and sinuses, blockage of ostia draining the sinus (particularly the ostia of the maxillary sinus), interference with the clearance mechanism and stasis (Turner *et.al.*, 1992). These factors favor secondary infection by the multitude of bacteria already present in the nasal cavity. With chronic infection the sinus ostia may well become totally blocked leading to a negative pressure and low oxygen concentration combined with an impaired blood supply to the nasal mucosa may explain the high frequency of anaerobic organisms found (Fredrich and Braude, 1974).

There are some conditions that lead to chronic or recurrent sinusitis as follows :- (Zinreich, 1992).

- 1-Untreated acute sinusitis that results in damage of the mucous membranes.
- 2-Chronic medical disorders that cause inflammation in the air ways or persistent thickened stagnant mucus.
- 3-Structural abnormalities.

1-2-6 Normal Flora of The Nose :-

A wide spectrum of organisms colonizes the nose, such as *Streptococcal* and *Staphylococcal* species. Occasional outbreaks of

disease due to this organism, can be traced to nasal, skin or perianal carriage by personnel (Baron and Finegold, 1991).

The flora of paranasal sinuses include *streptococcus pneumonia*, *Haemophilus influenza*, *Staphylococcus aureus* and *Staphylococcus epidermidis* and anaerobes such as *Bacteroides spp.*, anaerobic gram positive cocci and *fusobacterium spp.*, Corynebacteria, Viridans Streptococci, β -haemolytic Streptococci and *M. Catarrhalis* (Almadori *et.al.*, 1986). Enterobacteriaceae and non-fermentative Gram-negative group (*Acinetobacter spp. and Pseudomonas spp.*) may colonize the nasopharynx of elderly, immunodeficient or malnourished patients particularly when they have received antimicrobial agents. Normal flora are generally non pathogenic and the use of corticosteroid and immune suppressive agents leads to the emergence of opportunistic infection by organisms which are not previously regarded as pathogens (Vandepitte *et.al.*, 1991). The most common normal flora which causes sinusitis will be discussed in detail as follows:

1-2-6-1 Gram-positive bacteria

A- *Streptococcal spp.*:

Streptococcal spp are Gram positive, catalase- negative, facultative anaerobic bacteria, form spherical or ovoid cells that characteristically form pairs or chains during growth (Facklam and Eliot, 1990).

- *Streptococcus pyogenes*:

β - Hemolytic, bacitracin susceptible contain group A antigen. It's the main human pathogen which causes a wide range of suppurative infections in the respiratory tract and skin life-threatening soft tissue infections (Kirby and Ruoff, 1990).

Strains of *Streptococcus pyogenes* express a large arsenal of virulence factors and hence their pathogenicity and the clinical signs that they induce are very diverse (Boyle, 1990).

The adhesion by interaction with host fibronectin, a matrix protein on prokaryotic cells, is considered the principal mechanism by which *Streptococcus pyogenes* bind to epithelial cells of respiratory tract and skin (Kilian, 2002).

- **Viridans Streptococci:-**

Viridans group consists of several species which may be either α - hemolytic with production of green color around their colonies on blood agar or non hemolytic, these bacteria are dominant members of the resident flora of oral cavity and pharynx in all age groups (Douglas *et.al.*, 1993).

Viridins groups are part of normal flora and comprise a large proportion of the commensal bacteria in occasionally cause Bacteremia or endocardities; however the mechanism of pathogenesis is poorly understood (Jacops *et.al.*, 1990).

- ***Streptococcus pneumoniae*:-**

It is one of the major causes of sinusitis both in adult and children; *Streptococcus pneumonia* is a primary cause of bacterial pneumonia, meningitis and otitis media (Tinkel man and silk, 1989). The most important virulence factor is the capsular polysaccharide and disease results from the ability of this microbe to invade and multiply in tissue (Monora *et.al.*, 1997).

The production of pneumolysin and H_2O_2 which acts as toxins by *S. pneumoniae* in certain concentration leads to inhibition of cilia beating frequency of the epithelium of the respiratory tract and inhibiting the antimicrobial properties of the neutrophils and opsonic activity of serum (Chilvers *et.al.*, ۲۰۰۰) .

Streptococcus pneumoniae infection frequently follows the viral infection that may produce mucosal damage, diminish the epithelial ciliary's activity and depress the function of macrophage (Kim *et.al.*, ۱۹۹۵). In the tissue, *S. pneumoniae* multiplies and spreads throughout the lymphatic or direct extension from the local site of infection or through blood stream causing bacterimia (Torzillo *et.al.*, ۱۹۹۵).

B- Staphylococcal spp.:-

They are gram-positive spherical bacteria, non spore forming, aerobic, non motile usually arranged in grape like irregular clusters. The main species are *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Staphylococcus saprophyticus* (MacFaddin, ۲۰۰۰).

The pathogenicity of *Staphylococcus aureus* contributes to hemolysis of the blood, coagulation of the plasma and production of intracellular enzymes and toxins which act on host cell membrane and mediate the cell destruction (Brooks *et.al.*, ۲۰۰۴).

Infection of maxillary sinusitis with this organism has been described as the staphylococcal beta-toxin and may reduce ciliary's activity and induce sinusitis without occlusion of the natural ostium of maxillary sinus (Kim *et.al.*, ۲۰۰۰). Treatment of *staphylococcus aureus* infection has become increasingly problematic due to the emergence of multidrug resistant strains (Lowy, ۱۹۹۸). More than ۴۰% of *Staphylococcus aureus* contain plasmids that encode B-lactamase,

the enzyme that degrades many penicillin drug by virtue of change in the penicillin-binding protein in their cell membrane; these strains are commonly known as Methicillin Resistant *Staphylococcus aureus* (MRSA) (Herwald, 1999).

Staphylococcus epidermidis (coagulase-negative Staphylococci): this type of Staphylococci constitutes a major component of the normal flora of humans; causing nosocomial infections (Kloss and Bannerman, 1994).

The virulence factors of coagulase-negative Staphylococci follow initial colonization, a copious amount of extra cellular polysaccharide or slim which may correlate with pathogenicity and bacterial adherence (Heikens *et.al.*, 2000).

1-2-6-2 Gram-negative bacilli

A- *Enterobacteriaceae*:-

Enterbacteriaceae are large heterogeneous group of gram negative rods whose natural habitat in intestinal of human (Eisenstin and zaleznik, 2000). The gut is considered a primary source for dissemination and transmission of these pathogens to susceptible sites (Zogaj *et.al.*, 2003).

- *Escherichia coli*:-

Escherichia coli is one of most important Enterbacteriaceae species, usually motile, produce polysaccharide capsule; they grow on non selective media, most strain ferment lactose producing large red colony on MacConkey agar and this bacteria is predominant among aerobic commensal bacteria in healthy human intestine (Smith and Scotland, 1993).

Escherichia coli has many virulence factors that enable it to cause disease and the most important virulence factor is polysaccharide of somatic antigen O and capsular antigen K which protect the organisms from bactericidal effect of complement phagocytes in the absence of antibody (Chart, 1998).

Many strain express haemolysin S and in general strain of *Escherichia coli* isolated from extra intestinal infection are more haemolytic than strain isolated from the fascies of healthy human (Chart and Tenkin, 1999).

- ***Klepsiella pneumoniae***:-

Non motile exhibit mucoid growth, they have large polysaccharide capsule; *Klepsiella pneumoniae* is opportunistic bacteria among gram-negative bacilli responsible for nosocomial infections (Henry and Richard, 1998).

The polysaccharide capsule protects the bacteria by preventing phagocytosis and plays an important role in pathogenesis by interacting to the mucus producing cell (Podschum and Ullman, 1998).

- ***Proteus mirabilis***:-

Proteus mirabilis is a small gram negative, oxidase negative, swarming motility appearing flat with tapered edge on MacConkey agar and not ferments lactose (Farmer, 2000).

This bacteria have many virulence factors like fimbria, flagella, out membrane protein, lipopolysaccharide, capsule antigen and hemolysin which attribute to the pathogenesis of this bacteria (O'Hara *et.al.*, 2000).

B- *Haemophilus influenzae*

Haemophilus influenzae is a small, non motile, non spore forming bacterium and a parasite of humans which is found principally in the upper respiratory tract (Monox and Murphy, ۲۰۰۰).

Among *H. influenzae* strains, there are ۲ broad categories typeable and non-typeable strains are typed based on capsular characteristic. The capsule is composed of sugar-alcohol phosphate (i.e. polyribitol phosphate).complex. Differences in this complex are the basis for separating encapsulated strains into one of six groups: type a, b, c, d, e, r, and f, type b-*inflauzae* is most commonly encountered in serious infections in human (Moller *et.al.*, ۱۹۹۵).

Harabuchi *et.al.*(۱۹۹۴) stated that non-typeable strains do not produce a capsule and are most commonly encountered as normal inhabitants of the upper respiratory tract. Non capsulated strain usually cause localized infections such as otitis media, sinusitis, conjunctivitis and exacerbation of chronic bronchitis (Nizet *et.al.*, ۱۹۹۶). For encapsulated strains of *H. influenzae* (is most common) the capsule is antiphagocytic in the absence of specific anticapsular antibodies and highly associated with virulence of other cell envelope factors like pilli, lipopolysaccharied and outer membrane protein also which may facilitate attachment to host cell (Harper and tilse, ۱۹۹۱) .

For non-capsulated strains,pili and other cell surface factors, not fully understood, play a role in attachment to host cell; *H. influenzae* produces no demonstrable exotoxine and the role of its somatic antigens in natural disease is not clearly understood. The events that result in entry into the intravascular compartment by *H. inflaenzae* serotype b are unclear (Stephenson *et.al.*, ۱۹۸۵).

1-2-6-3 Gram-negative cocci

- *Moraxella catarrhalis*

M. catarrhalis is gram-negative, aerobic, oxidase positive diplococcic. It was described for the first time in 1896. It is considered to be a member of the normal flora of upper respiratory tract; it can cause sinusitis, otitis media, bronchitis and pneumonia (Abuhammour *et.al.*, 1999).

The way of transmission is by the spread of patients endogenous strain to normally sterile site, person to person; nosocomial spread by contaminated respiratory droplets also can occur (Janda and Knapp, 2003). Liberation of endotoxin, histamine and chemotactically active factors can be considered the major pathogenicity factors of *M. catarrhalis*. The surface of these bacteria contains an outer membrane protein (Karalus and Campagnari, 2000).

The pathogen can protect itself by binding of the C₃ a subcomplement of the complement system followed by subsequent formation of a functionally inactive complex with C₃ and, on the other hand, by inactivation of the terminal lytic complement complex by means of specific protein on the surface of the outer cell wall (Murphy, 1996 and Cullman, 1997).

1-2-7 Bacterial Causes of Sinusitis :-

Various studies were done concerning the microbiology of acute maxillary sinusitis: Bjorkwall (1900) pointed out that *S. pneumonia* was the most common microorganism being isolated from sinusitis which was followed by other *Streptococcal spp.* Mounier-kuhn (1903) observed similar proportions. Dishock and Franssen (1907) demonstrated three times as many cases infected by *Streptococcus pyogenes* as by *S.*

pneumonia and *H. influenzae*. These findings were confirmed by Gronroos and Palva (1962).

Wald *et.al.*(1981) mentioned that the most common organisms found in acute maxillary sinusitis were *S. pneumoniae*, *S. pyogenes*, *staphylococcus aureus*, *H. influenzae*, *E. coli* and *M. catarrhalis*. Hamory *et.al.*(1979) reported that the bacterial causes of acute sinusitis were: *S. pneumoniae*, *H. influenzae*, *M. catarrhalis*, *S. pyogenes*, *staphylococcus aureus*, *pseudomonas aeruginosa* and *E. coli*.

Moreover, studies showed that *H. influenzae* constitute the highest percentage (20%), followed by *S. pneumoniae* (19%), coagulase-negative staphylococcus (8%), *S. pyogenes* (2%), *M. catarrhalis* (2%), *Staphylococcus aureus* (1%), Viridanse Streptococcal group (1%) and negative culture was (24%) (Hannell *et.al.*, 1988). Farhadi *et.al.*(1989) showed that the most common bacteria were *Staphylococcus aureus* followed in order by *S. pneumoniae* and some members of enteric group were represented by *E. coli*, *klebsiella pneumoniae*, *proteus merabilis*, *Enterobacter*, and *pseudomonas*.

Berg *et.al.*(1988) showed that the pathogens being isolated from patient with untreated acute sinus empyema were *S. pneumoniae* being the most prevalent followed by *H. influenzae*, *staphylococcus aureus* and anaerobic *Bacteriodes spp.*, while Brook *et.al.*(1990) discussed the frequency of potential bacterial pathogens including *S. pneumoniae*, *H. influenzae* and *M. catarrhalis* in patients clinically diagnosed with sinusitis.

In chronic sinusitis bacteriological culture showed that beside *S. pneumoniae*, *H influenzae* and *M. Catarrhalis* additional bacteria are present including anaerobes (*Bacteriodes spp.*, *peptostreptococcus spp.*) and gram negative bacteria (*pseudomonas aeruginosa*, *klbsiella pneumoniae*, *E. coli*). Evan *etal.*,(1970); Laine and Smoker (1994) and

Berg *et.al.* (1988) found that *S. aureus* and anaerobic bacteria were frequently finding in chronic sinusitis. Hartog *et.al.* (1990) found that the predominant aerobic bacteria were *S. aureus* (42%) and *S. pneumoniae* (32%) while the predominant anaerobic bacteria were *propionibacterium spp.* (8%) and gram negative rods (9%). Brook *et.al.* (1996) concluded that the predominant bacteria in chronic sinusitis were *S. aureus* followed by *Prevotella spp.*, *Fusobacterium* and *Bacteriodes fragilis*.

Another study done by Bile *et.al.* (1998) found that Coagulase negative staphylococci were the most common isolates (36%) followed by *S. aureus* (20%), *S. viridans* (8.3%), *Corynebacterium* (4.6%) and anaerobe bacteria (6.4%). Brook and Fraizer (2001) found that the predominant aerobic isolates were *S. aureus* (14) isolates, *S. pneumoniae* (14) isolates, *pseudomonas aeruginosa* (12) isolates and *M. Catarrhalis* (10) isolates; the predominant anaerobes were *Peptostreptococcus spp.* (6) isolates, *Prevotella spp.* (4) isolates, *Fusobacterium spp.* (1) isolates and *propionibacterium acnes* 14 isolates. Ologe and Nwabisi (2003) recorded that the main cause of chronic sinusitis were *S. aureus* (48.1%), *E. coli* (20.4%), *Klebsiella spp.* (20.4%) and *S. pneumoniae* (3.7%). In (2000) Brook and Frazier found that the predominant aerobic bacteria were *S. aureus* followed by *S. pneumoniae* and *M. Catarrhalis* while anaerobic bacteria were *Peptostreptococcus spp.* followed by *Prevotella spp.*, *Porphyromonas asaccharolytica* and *Fusobacterium spp.* Recently, Mounghong *et.al.* (2000) reported that the predominant organisms among sinusitis were *pseudomonas aeruginosa*, *Staphylococcus spp.*, and *Streptococcus spp.* also *H. influenzae* consequently.

The causative agents of sinusitis are widely variable. This may be related to the environmental, climatic, familial or other factors. Hence the suggested recommendation is to study sinusitis locally.

1-2-8 Treatment:-

The goal of treatment for both acute and chronic sinusitis is drainage of the congested sinus and elimination of the pathogenic bacteria (Benninger *et.al.*, 1997). Studies using pre and post treatment sinus aspirate culture have shown that the antimicrobials with appropriate spectra and given in adequate doses and with an adequate duration are effective in eradicating or substantially reducing bacterial titer in the sinus (Gwaltney *et.al.*, 1992); but pre treatment sinus aspiration may not yield bacterial pathogen in approximately 30% to 40% of patients with acute sinusitis; for this the initial choice of antibiotic should be selected on an empirical basis (Chow *et.al.*, 1992).

A broad spectrum antibiotic is typically chosen to cover the usual sinus pathogen (*H. influenzae*, *S. pneumoniae*, *M. catarrhalis*, *S. aureus*) like amoxicillin or tetracycline, erythromycin as an initial treatment for management of sinusitis (Temple and Nahata, 2000). There is an increasing incidence of resistance to β -lactam drug, cephalosporin providing more powerful alternative for non responders (Shikani, 1997). If drug therapy fails to unblock the sinuses or other complications such as structural abnormalities so in this cases surgery may be required (Lanza and Kennedy, 1992).

Chapter Two

Materials & Methods

۲-۱) Materials:-

Many types of instruments and chemical materials in addition to biological materials were used in this study for completing the research work. These materials were taken from different sources and companies which are listed in the following tables.

۲-۱-۱) Cultures media:-

Table ۲-۱ shows the list of culture media which were used for isolating and identifying the isolated bacteria.

Table ۲-۱: The Culture Media

S.No	Culture media	company
۱.	Nutrient agar	Mast Lab. (Uk)
۲.	nutrient broth	Mast Lab. (Uk)
۳.	brain heart infusion broth	Mast Lab. (Uk)
۴.	brain heart infusion agar	Mast Lab. (Uk)
۵.	MacConcky agar	Mast Lab. (Uk)
۶.	blood agar	Mast Lab. (Uk)
۷.	Muller-Hinton agar	Mast Lab. (Uk)
۸.	Agar-agar media	Mast Lab. (Uk)
۹.	Peptone water medium	Mast Lab. (Uk)
۱۰.	Simmon Citrate agar	Difco-Michigan
۱۱.	Kliglar iron agar	Difco-Michigan
۱۲.	MR-VP broth	Difco-Michigan
۱۳.	Meet extract	Difco-Michigan
۱۴.	Urea base agar	Difco-Michigan
۱۵.	EMB agar	Difco-Michigan

५-१-५ Chemical materials:-

Table ५-५ shows chemical materials used in this study for identifying different types of bacteria.

Table ५-५ Chemical Materials Used in This Study.

S.No	Company	Chemical materials
१	Crystal violet	Fluka (UK)
२	Safranin	Fluka (UK)
३	Tetramethyl-p Phenylene-diamine dihydrochloride (oxidase test)	Fluka (UK)
४	Nacl	Fluka (UK)
०	H ₂ SO ₄	Fluka (UK)
६	Urea	Fluka (UK)
७	Kovac's reagent	Fluka (UK)
८	Phenol red	B.D.H (UK)
९	Bromo phenol blue	B.D.H (UK)
१०	Iodine	B.D.H (UK)
११	KOH	B.D.H (UK)
१२	Barium chloride	B.D.H (UK)
१३	α-naphthol amine	B.D.H (UK)
१४	Glycerol	B.D.H (UK)
१०	Absolute Ethanol	B.D.H (UK)
१६	Hcl	B.D.H (UK)
१७	Toluidine blue	B.D.H (UK)
१८	Acetic acid (०m)	B.D.H (UK)
१९	Amyle-alchole	B.D.H (UK)
२०	kNo ₃	B.D.H (UK)
२१	H ₂ O ₂ (hydrogen peroxide)	Oxide

ॡ-ॡ-ॢ Carbohydrate Materials:-

Table ॡ-ॢ shows carbohydrate materials used in the sugar fermentation test.

Table ॡ-ॢ Carbohydrate Materials Used in This Study

S.No	Materials	Company
ॡ	Sucrose	Difco (USA)
ॢ	manitol	Difco (USA)
ॣ	Lactose	Difco(USA)
।	Maltose	Difco(USA)
॥	Rhamnose	Fluka (UK)
०	Trehlose	Fluka (UK)
ॡ	Starch	Sigma
ॢ	Arabinose	B. D.H (UK)

ॡ-ॡ-। Diagnostic Disks:-

- ॡ. Bacitracin (ॡ० IU) (Himedia, India)
- ॢ. Optochin (ethyl hydrocuprein hydrochloride ०µg) (Himedia, India)

ॡ-ॡ-॥ Antibiotic Sensitivity Disks:-

The antimicrobial sensitivity disks used in this study were made by Bioanalyse, Oxoid Company and listed in Table ॡ.॥. They were used for detecting the sensitivity of isolated bacteria for these antibiotics. The results of this study were recorded according to standard guidelines recommended by National Committee for Clinical Laboratory Standards (NCCLS, ॡ०००).

Table 3-3 Antimicrobial Discs Used in This Study

Antimicrobial agent	Symbol	Concentration (μg)/disk	Diameters of inhibition zones (mm)		
			resistance	intermediate	sensitive
Ampicillin	Am	10	≤ 11	12-21	≥ 22
Amoxicillin	Amx	20	≤ 19	-----	≥ 20
Trimethoprim	TMP	30	≤ 10	11-15	≥ 16
Erythromycin	E	10	≤ 13	14-22	≥ 23
Tetracycline	TE	30	≤ 14	15-18	≥ 19
Cefotaxime	CTX	30	≤ 14	15-22	≥ 23
Ciprofloxacin	Cip	5	≤ 10	16-20	≥ 21
Cephalothin	KF	30	≤ 14	15-17	≥ 18
Gentamicin	GN	10	≤ 13	14-16	≥ 17
Vancomycin	VA	30	≤ 9	10-11	≥ 12
Oxacillin	OX	1	≤ 13	-----	≥ 10

2-1-6 Preparation of The Culture Media:-

The general culture media described below were prepared by the routine methods and used in appropriate experiments:-

2-1-6-1 *Blood Agar Medium:*

Blood agar medium has been prepared according to Macfaddin (2000) by dissolving 4 gm blood agar base in 100 ml D.W. and autoclaved at 121°C for 10 min, then cooled to 50°C and 8% of human blood was added. This medium was used to cultivate bacterial strain and to determine their ability to blood haemolysis.

2-1-6-2 *Chocolate Agar Medium:*

Chocolate agar medium has been prepared by dissolving 4 gm of blood agar base in 100 ml D.W. and sterilized by autoclaving. Then 8% of human

blood was added to the medium after cooling to 40 C°. This medium was especially used for isolation and cultivation of bacteria that need 5-10% CO₂ tension (Baron and Finegold, 1996).

2-1-6-3 *MacConkey Agar Medium:*

MacConkey agar medium has been prepared according to the method recommended by the manufacturing company and it is used for the primary isolation of most Gram- negative bacteria and differentiation of lactose fermentative from the non lactose fermentative (Collee *et.al.*, 1996).

2-1-6-4 *Nutrient Agar Medium:*

Nutrient agar medium has been prepared according to the manufacturing company. It has been used for general experiment isolate culture, cultivation and activation of bacterial isolates when it is necessary (Macfaddin, 2000).

2-1-6-5 *Mannitol Salt Agar Media*

This media has been used as a selective media for the isolation of Staphylococci and differentiation of *Staphylococcus aureus* (Macfaddin, 2000).

2-1-6-6 *Muller- Hinton Agar*

Muller- Hinton agar has been prepared according to the method recommended by (Cruikshank *et.al.*, 1970) and it is used in anti-microbial susceptibility testing.

2-1-6-7 *Brain Heart Infusion Broth*

This medium has been used for the enhancement of bacterial growth and reactivation of bacterial isolates when it is necessary (Macfaddin, 2000).

۲-۱-۶-۸ *Brain Heart Infusion Agar*

This medium has been used as a storage medium after the addition of ۰% glycerol it is prepared according to (Cruikshank *et.al.*, ۱۹۷۰).

۲-۱-۶-۹ *Gelatin agar media*

Gelatin agar media used for the detection of bacterial ability to proteolytic or liquefy gelatin. It was prepared by adding ۴.۴ % of gelatin to the nutrient agar medium (Macfaddin, ۲۰۰۰).

۲-۱-۶-۱۰ *Kligler Iron Agar*

Kligler Iron agar has been used for determining glucose and lactose fermentation and possible hydrogen sulfide H₂S production as a first step in the identification of Gram- negative bacilli (Macfaddin, ۲۰۰۰)

۲-۱-۶-۱۱ *Pepton Water Media*

Pepton Water Media was prepared and used to detect the ability of bacteria to produce Indole according to the method described by (Baron and Finegold, ۱۹۹۶).

۲-۱-۶-۱۲ *MR-VP Medium*

MR-VP medium has been prepared and used to detect the partial and complete hydrolysis of glucose according to Macfaddin (۲۰۰۰).

۲-۱-۶-۱۳ *Simmons' Citrate Medium*

Simman's Citrate Medium has been used for determining the ability of bacteria to utilize citrate as the sole carbon source (Macfaddin, ۲۰۰۰).

2-1-6-14 *Nitrate Reduction Medium*

Nitrate reduction medium has been prepared by dissolving a mixture of 0.5 gm of potassium nitrate (KNO₃) and 0 gm peptan in 100 ml of D.W. then the medium was dispensed in test tubes, 5 ml in each one and autoclaved. It was used for detection the ability of bacteria to reduce nitrate into nitrite as recommended by Baron and Finegold (1996).

2-1-6-15 *Sugar Fermentation Medium*

Sugar fermentation medium has been used for determining the ability of an organism for sugar fermentation and gas production. The medium was prepared according to Macfaddin (2000) as follows:-

a- Medium Base:-

Medium Base was prepared by dissolving 1 gm peptone, 1 gm meat extract, 0 gm sodium chloride and 0.01 gm phenol red in one liter of D.W. the ph was adjusted to 7.4; the medium was dispensed in tubes in 5 ml amounts. Durham tube was inverted in each tube and then sterilized by autoclave.

b- Sugar Solution:-

Sugar solution was prepared by dissolving 1 gm of required sugar in 100 ml D.W and sterilized by chloroform (Macfaddin, 2000). Then 0.1 ml of this solution was added to each test tube mentioned in (a) above.

2-1-6-16 *Eosin Methylene Blue (EMB) Agar:-*

Eosin Methylene Blue has been prepared according to Collee *et.al.* (1996) and used to differentiate *E. coli* which exhibits a characterized greenish metallic sheen colonies medium.

2-1-6-17 *Urea agar medium (Biolife)*

The base medium was prepared according to manufacturing company, autoclaved and cooled to 50°C then added 5ml of 20% filtration sterilized urea solution to 90ml of media in which this media insured that their pH was (6.8-6.9) and tubed the medium as deep slopes. This medium was used for detecting bacterial ability to produce urease enzyme (MacFaddin, 2000).

2-1-6-18 *Motility medium*

This medium was prepared according to McFadden(2000) by adding 2gm of agar-agar to 100 ml of nutrient broth in 1000 ml of distilled water then sterilized by autoclave at 121°C for 10 minutes then it was distributed in tubes; it was used to detect bacterial motility (MacFaddin, 2000).

2-1-7 *Reagent and Solutions:-*

2-1-7-1 *Oxidase Preparation:-*

This reagent was prepared by dissolving 1 gm of (tetramethyl-paraphenylene-diamine-dihydrochloride) in 100 ml of D.W and immediately used (Baron and Finegold, 1991).

2-1-7-2 *Catalase Preparation :-*

This reagent was prepared in (30%) using H₂O₂ dilute by D.W and stored in a dark container (Baron and Finegold, 1991).

2-1-7-3 *Methyl Red Reagent:-*

Methyl Red Reagent was prepared by dissolving 0.1 gm of methyl red in 300 ml of 90% ethanol and then the volume was completed to 500 ml by D.W(Macfaddin, 2000).It was used to detect the complete glucose hydrolysis .

2-1-7-4 Voges-Proskaur Reagent:-

It is composed of two solutions as below:-

- a- α -Naphthol-reagent was dissolved in 100 ml of 99% ethanol.
- b- Potassium Hydroxide (KOH) solution: - 4 gm of KOH was dissolved in 100 ml of D.W. and it was used to detect the partial glucose hydrolysis (Collee *et.al.*, 1996).

2-1-7-5 Indication of Nitrate Reduction:-

It is composed of two solutions:-

- a- Sulfanic acid solution: - prepared by dissolving 1 gm of Sulfanic acid in 100 ml of acetic acid(0m).
- b- α - Naphthylamine Solution: - prepared by dissolving 1 ml of α - naphthylamine in 100 ml of acetic acid (Collee *et.al.*, 1996).

2-1-7-6 Gram Stain Solutions:-

The reagents of this staining method were routinely prepared according to Talib (1996).

2-1-7-7 Covac's Reagent:-

It was prepared by dissolving 0 gm of P-dimethylamine benzylaldehyde in 100 of amyle-alcohol and was added 30 ml of concentrate HCl acid. It was used to detect the Indol production (Baron and Finegold, 1991).

2-1-7-8 McFarland standard solution:

McFarland standard solution was used in antimicrobial susceptibility test, tube No. (1.0) was used which prepared by adding 1.0 ml of $\text{BaCl}_2 \cdot 2\text{H}_2\text{O}$ con. (1.170%) to 9.9 ml of H_2SO_4 con. (1%). The

tube No.(10) of McFarland standard tubes was used to compare the bacterial cell in suspension which given a cell density 1.0×10^8 cell/ml. (Baron *et.al.*, 1996).

2-1-7-9 Frazier's reagent

This reagent was prepared according to the method being recommended by (Collee *et.al.*, 1996). It was prepared by dissolving 0 gm HgCl₂ in 20 ml of concentrated HCL (38%), with the addition of 100 ml distilled water. It was used to detect the ability of bacteria to analyze gelatin (Collee *et.al.*, 1996).

2-2 Methods:-

2-2-1 Subjects:-

This study was including one hundred patients 60 were females and 40 were males. Fifty eight cases were clinically diagnosed as acute sinusitis and 42 were diagnosed as chronic sinusitis patients who were attending the ENT department in Hilla Teaching Hospital during the period between October 2004 and July 2006. The age of the patient ranged from 10-79 years. The primary clinical diagnosis of those patients was done by consultant ENT sergeant through clinical examination, sinus occipitontal view (CT-Scan) of sinuses or Nasoendoscopy.

Patients who were under antibiotic treatment or had stopped antibiotics therapy for less than one week were excluded. All patients were subjected to especially made case sheet which has been highlighted on patient's age, type of sample, antibiotic history, clinical feature, concomitant disease, clinical diagnosis as shown below:-

2-2-2 Collection of Specimens:-

The proper specimens collected for bacteriological analysis are described below. Those specimens were collected under the help of advisory to avoid any possible contamination.

For acute sinusitis the specimens were collected by nasal swabs (middle meatus) especially for maxillary sinusitis excluded other sinusitis while antral wash (lavage) for chronic sinusitis. The nasal swabs were obtained as follows:

- 1- Sterile cotton swabs were moistened with two drops of sterile saline and carefully inserted into the patient's nostril (the swab tip must be inserted up to 2.0 cm from the edge of the nares).
- 2- The swabs were rotated 6 times.
- 3- Other swabs were inserted into the second nostril following the same way above (Fagnan, 1998).

These swabs were immediately cultured and incubated for a further laboratory diagnosis.

In chronic sinusitis antral wash samples were obtained according to Sutter *et.al.* (1980) by using lavage instruments (Trocar and Canula) under local or general anesthesia. Sinus secretion was aspirated into sterile 20 ml syringe. If aspiration yields nothing, then the canula was held in the sinus and one to two milliliter of sterile physiological saline was injected into the sinus through the canula and was aspirated again (this is called injection aspiration). Each specimen was immediately inoculated on the blood agar plates, chocolate agar plates and MacConkey's plates. All plates were incubated aerobically at 37°C for 24-48 hrs.

2-2-3 Laboratory Diagnosis

According to the diagnostic procedures recommended by Maccfaddin (٢٠٠٠); Baron and Finegold (١٩٩١); Prescott *et.al.*(١٩٩٠), the isolation and identification of gram positive and gram negative bacteria in the nasal sinus of patients were performed as follows:-

٢-٢-٤ Microscopy Examination and Colonial Morphology

Microscopical examination included the examination of shape, gram stain reaction, arrangement of cell, capsule, flagella and spore formation which has been done for the different colonies, then the colonial morphology which included color and natural of pigments, translucency, edge, and elevation have been studied with respect to the ٢٤ hours bacterial growth. Plates which showed no growth were further incubated up to ٧٢ hours before discarding them as negative.

٢-٢-٥ Physiological and Biochemical Tests

٢-٢-٥-١ Oxidase Test

A filter paper circle was placed into a sterile plastic disposable petridish and moistured with several drops of the freshly prepared oxidase reagent, then a small portion of the colony to be tested was removed and rubbed on the filter paper changing in the color to blue or purple within ١٠ seconds indicated for a positive result (Baron and Fingold, ١٩٩١).

٢-٢-٥-٢ Catalase Test:-

By streaking the nutrient agar medium with the selected bacterial colonies and incubated at ٣٧ C° for ٢٤ hrs then transfer the growth by the loop and put it on the surface of a clean slide and add a drop of (٣٠٪ H₂O₂), positive result when the gas bubbles appear (Barond Finegold, ١٩٩٦).

ॡ-ॡ-ॡ-ॡ Blood Hemolysis Test

Blood agar medium was streaked with a pure culture of bacterial isolate to be tested and incubated at ॡॡ C° for ॡॡ-ॡॡ hrs. The appearance of a clear zone surrounding the colony is an indicator of β - hemolysis while the greenish zone is an indicator of α - hemolysis (Cowan, ॡॡॡॡ).

ॡ-ॡ-ॡ-ॡ Coagulase Test:-

This enzyme was tested by two methods:-

a. Slide test for bund coagulase (clumping factor):-

A drop of human plasma was placed on a clean, dry glass slide, a drop of D.W. was placed next to the drop of plasma as a control. By a sterile loop an amount of the isolated colony was emulsified with each drop. When clumping the plasma, bacteria was observed and a smooth homogenous in the control; the result was recorded positively (Baron and Finegold, ॡॡॡॡ).

b. Tube test for free coagulase:-

ॡ.ॡ ml of Human plasma was placed in a glass tube and a visible portion of growth from isolated colonies was emulsified in the plasma by rubbing the material on the slide of the tube while holding the tube at an angle, then the suspension was incubated for ॡ-ॡ hrs at ॡॡ C°; the presence of clot that cannot be resuspended by gentle shaking was recorded as a positive result. The organism that fails to clot the plasma within ॡॡ hrs is considered as coagulase negative (Baron and Finegold, ॡॡॡॡ).

ॡ-ॡ-ॡ-ॡ Urease Production Test

A slant of medium was inoculated with the colony of tested organism and incubated at 37 C° and examined after 24 hrs and 48 hrs. Urea splitting organisms were identified by the change of the color from yellow to purple-pink (Cruikshank *et.al.*, 1970).

2.2.5.6 *Gelatin Hydrolysis:-*

Gelatin agar medium was inoculated with the colony of tested organism and incubated for 3-5 days at 37 C°, and then the plates were flooded in solution of Frazier reagent for 10 min. Gelatin-liquefying organism was identified by the presence of clear zone around the colony of the organisms under test (Maccfadin, 2000).

2.2.5.7 *Citrate Utilization Test :-*

The surface of simmon's citrate slant medium was inoculated with colony of the tested bacteria and incubated at 37 C° for 1-3 days. Conversion of the indicator's color from green to blue indicates that the organism was able to utilize citrate as a sole carbon source (Cruikshank *et.al.*, 1970).

2.2.5.8 *Nitrate Reduction Test:-*

Nitrate broth was inoculated with the tested bacterial isolate and incubated at 37 C° for 24 hrs. Three drops of reagent A and 3 drops of reagent B were added to the suspension of bacteria in broth positive test for reduction of nitrate to nitrite was indicated by the development of a red color within 10 mins. (Baron and Finegold, 1991).

2.2.5.9 *Motility Test:-*

Tubes containing motility test medium were stabbed once at the center with an inoculating needle and incubated at 30 C° for 24-48 hrs

motile bacteria speed out from the line of inoculation; but none motile bacteria grows only along the stab line, while the surrounding medium remains clear (Baron and Finegold, 1991).

2.2.5.10 Sugar Fermentation Test:-

Tubes containing sugar broth were inoculated from 24 hrs old bacterial culture and incubated at 37°C for 1-3 days. The positive result was detected by a change in color of the broth from red to yellow, with or without the appearance of air bubbles in Durham tube (MacFaddin, 2000).

2.2.5.11 Kligler's Iron Agar Test for H₂S Production:-

Only the colonies growing on MacConkey agar were touched by a straight wire and inoculated on the media by stabbing the butt of the tube and streaking the slant. Fermentation was detected by a change in the indicator phenol red to yellow. The PH changes in the butt and the slant of medium were recorded after 18- 24 hrs of incubation of gas formation which is usually visualized as bubbles in the medium caused by the gas formed in the agar. Organisms can produce H₂S form black precipitate in the butt (Baron and Finegold, 1996).

2.2.5.12 Indol Test

Tubes containing a peptone water medium were inoculated with the colony of the tested bacteria and incubated at 37°C for 18 hrs, then several drops of Kovac's reagent were added to the broth medium. After shaking, the appearance of the red ring on the surface was regarded as a positive result (Cruikshank *et. al.*, 1970).

2.2.5.13 Methyl Red Test:-

The tubes of the (MR-VP broth) were inoculated with the selected bacterial colonies and incubated at 37°C for 24 hrs, then (2 drops) of methyl red reagent were added to it. The appearance and observation of red colour means a positive result and a complete analysis of glucose (MacFaddin, 2000).

2-2-5-14 *Voges Proskaur Test:-*

The tubes of (MR-VP broth) were seeded with the specific bacterial culture and were incubated at 37°C for 48 hrs., then we read the result by adding (0.1 ml of α - naphthol reagent) and (0.2 ml of 1% NaOH solution); appearance of red colour after 10 min. means positive result due to partial analysis of glucose, which produce acetone or (Acetyl methyl-carbinol) (MacFaddin, 2000).

2-2-5-15 *Mannitol Salt Agar Test:-*

The differentiation between *Staphylococcus aureus* and other Staphylococci e.g. *Staphylococcus epidermidis* was done by sub culturing selected colonies on mannitol salt agar for 24 hrs at 37 C°. Colonies surrounded by a yellow halo indicating mannitol fermentation and isolates colony, is *Staphylococcus aureus* (MacFaddin, 2000).

2-2-5-16 *Optochin Susceptibility Test:-*

A half of 1% human blood agar plate was streaked with an inoculum from a pure isolates of the organism to be tested , then an optochin disc was placed in the center of the inoculum and incubated for 24 hrs at 37 C° in a candle jar , then observation of zones of growth inhibition greater than 14 mm surrounding the disc was considered positive and it was presumptive indication of *Streptococcus pneumoniae* (Baron and Finegold, 1991).

2-2-5-17 *Antimicrobial Sensitivity Test:-*

Two ml of brain heart infusion broth have been inoculated with an isolated colony of tested bacteria and incubated for 24 hrs at 37 C°. After that, the turbidity of bacterial suspension has been adjusted according to McFarland standard tube (1.0). 0.1 ml of bacterial suspension has been spread on the surface of Muller Hinton medium plate and left to dry. Antimicrobial disks have been placed properly and incubated for 24 hrs at 37 C° (Bauer *et.al.*, 1976).

The inhibition zone have been measured by using a ruler in compared with inhibition zone determined by (NCCLS, 2000) and to decide the susceptible of bacteria to anti microbial agent whether being resist or sensitive.

2-2-6 *Statistical Analysis*

Differences between means were compared by X² tests under confidence level of 0.90; the **P** value \leq 0.05; the **P** value \leq 0.01 and 0.99 was considered a significant difference and **P** value $>$ 0.05, 0.01 was considered a non-significant difference.

Chapter Three

Results & Discussion

٣-١ Age Related Disease:-

Figure (٣-١) shows that the most affected age group in this study in both acute and chronic sinusitis was between ٢٠ to ٢٩ years old. These results agree with the results obtained by Sarmad (٢٠٠٥) and Baily (١٩٨١) who found that the affected age group ranged from ٢٠ to ٣٠ years old who were the most susceptible for sinusitis.

These results can be explained as this age group is more exposed to environmental factors such as allergens, climatic factors, air pollution and the extremely weather changes. Furthermore, this age group represents the active age group having the chance of person to person transmission. Statistical analysis by using Chi Square showed that there is a significant relationship between age and sinusitis ($p < ٠.٠٥$).

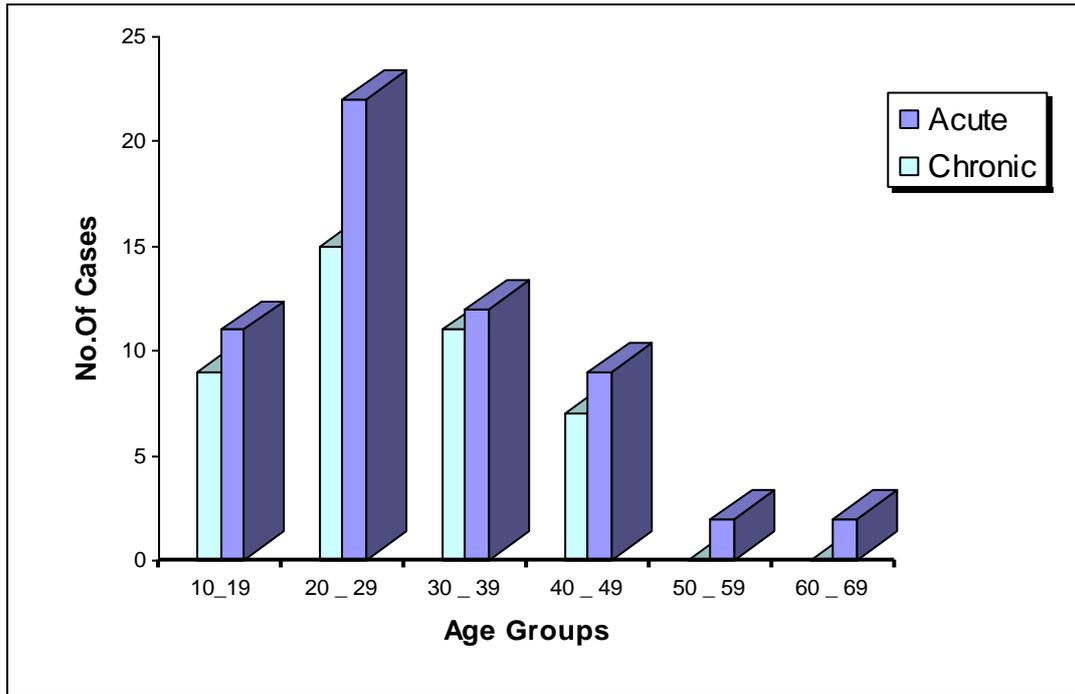


Figure 3-1: Age Distribution of Acute and Chronic Sinusitis Disease.

3-2 Clinical Features and Related Disease:-

Like other diseases, the clinical presentation depends on the severity of the disease. The purulent discharge, nasal obstruction and periorbital pain are the major complaint in sinusitis as shown in the Table (3-1).

In acute sinusitis the purulent discharge was observed in (80%), periorbital pain (90%) and nasal obstruction (97%). The results were in accordance with the results of a similar study done by Stalman *et.al.*(2001). Fever in acute sinusitis was the most frequent symptom. This result came to confirm the results being reported by (Brook, 2004) who states that fever is one of the most frequent features of patients suffering from acute sinusitis. In chronic sinusitis the purulent nasal discharge was observed in (47%), nasal obstruction (90%) and periorbital

pain observed in (90%). These results agree with the results obtained by Williams *et.al.* (1993). The percentage of fever in acute sinusitis was found to be higher than that at chronic sinusitis. This may be due to induction of the immune system with chronic sinusitis and to the administration of antibiotics.

Table 3-1 Clinical Features of Sinusitis Disease*:

Clinical features	Acute sinusitis	Chronic sinusitis
Purulent discharge	80%	47%
Periorbital pain	90%	90%
Nasal obstruction	97%	90%
Fever	90%	30%

* The clinical features were diagnosed by ENT expert.

3-2 Clinical Conditions and Related to Disease:-

According to the clinical conditions of sinusitis disease shown in Table (3-2), the most important predisposing conditions in chronic sinusitis were allergy (90%) and nasal polyp (80%) and Nasal septal division (80%). The results came to ensure the results of Wu *et.al.*(2000), who stated that the anatomical abnormalities of nasal cavity can be the basic reason of sinusitis. In addition to that the edema of nasal mucosa and the change of nasal discharge caused by allergy triggering factor of chronic sinusitis. These abnormalities will interfere with ventilation and the free passage of air through the nasal chamber, and with secretion and movement of mucous blanket thus predispose to infections. The

frequency of diabetes mellitus and dental problem in chronic sinusitis was (4%) and (2%) respectively. These results were close to those results obtained by Sarmad (2005), who found that (2.3%) of patients with sinusitis were suffering from diabetes mellitus versus (1%) who have dental problems.

The percentage beginning with common cold in both acute and chronic sinusitis was high enough to be in agreement to those results obtained by Desrosiers *et.al.* (2002), who found that (91%) of individuals with common cold will develop sinus disease by secondary infection of bacteria, and this was also ensured by Buchman *et.al.* (1994).

In acute sinusitis the percentages of nasal polyp, allergy and nasal septal division were 20%, 23% and 12% respectively as shown in Table (3-2). Sarmad (2005) who found that allergy and nasal polyp were accounted for (18.7%) and (7.3%) respectively in sinusitis cases while diabetes mellitus and dental problem was (3%), (1%) respectively.

Table 3-2 Clinical Conditions associated with Sinusitis Disease:

Clinical conditions	Acute sinusitis	Chronic sinusitis
beginning with common cold	97%	80%
allergy	23%	90%
nasal polyp	20%	85%
Nasal septal division	12%	80%
Diabetes mellitus	3%	4%
Dental problem	1%	2%

3-4 Sex Related Disease:-

In this study, 100 specimens were investigated, 68 specimens of them were from acute cases of sinusitis and 32 specimens were chronic cases. Among the acute cases, 60 specimens revealed positive results of bacterial culture versus 40 specimens which revealed no growth for bacterial culture. Among the 60 positive cases 36 specimens (60%) were females versus 24 specimens (40%) were males as shown in Figure (3-2). The result was in agreement with those results obtained by Stalman *et.al.* (2001) who found that (66%) of patients with acute sinusitis were females and (34%) were males. Moreover, a study done by Varonen *et.al.* (2003) reported that (50%) of the patients with acute rhinosinusitis were females and (50%) were males.

Among the cases of chronic sinusitis 30 bacterial cultures samples were positive and 20 bacterial cultures were negative; 20 (66%) of the positive cultures were females while 10 (33%) were males as shown in Figure (3-2). The results were similar to the results obtained by Ron *et.al.* (2004) who found that (66%) of patients with chronic sinusitis were females and (34%) were males. From these results one can conclude that the incidence of sinusitis is common in females in comparison with that of males. The reasons for this are not quite clear. The available literature regarding sinusitis is quite rare and no explanation was found in that literature explains the variability of sinusitis between males and females. However the reasons for that are attributed to the common exposure of females (as house worker) to various allergens such as (house dust, flour, dust, foods and food flavors, heat, humidity and other allergens), or to a lesser degree pregnancy and lactating women can effect their immunity leading to an increase in infection. However physiology of pregnant

women like increase in Progesterone and Estrogen will lead to increase incidence of allergy. Statistical analysis by using Chi Square showed that there is a highly significant relationship between sex and sinusitis ($p < 0.01$).

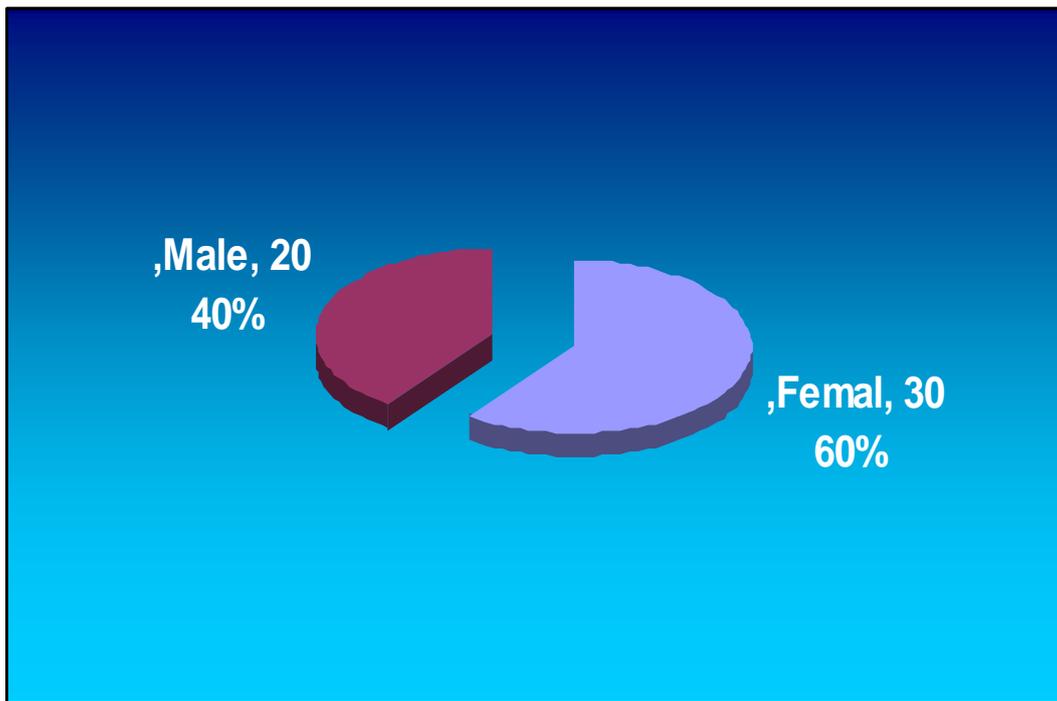


Figure 3.2: The Ratio of Male and Female in Positive Acute Sinusitis.

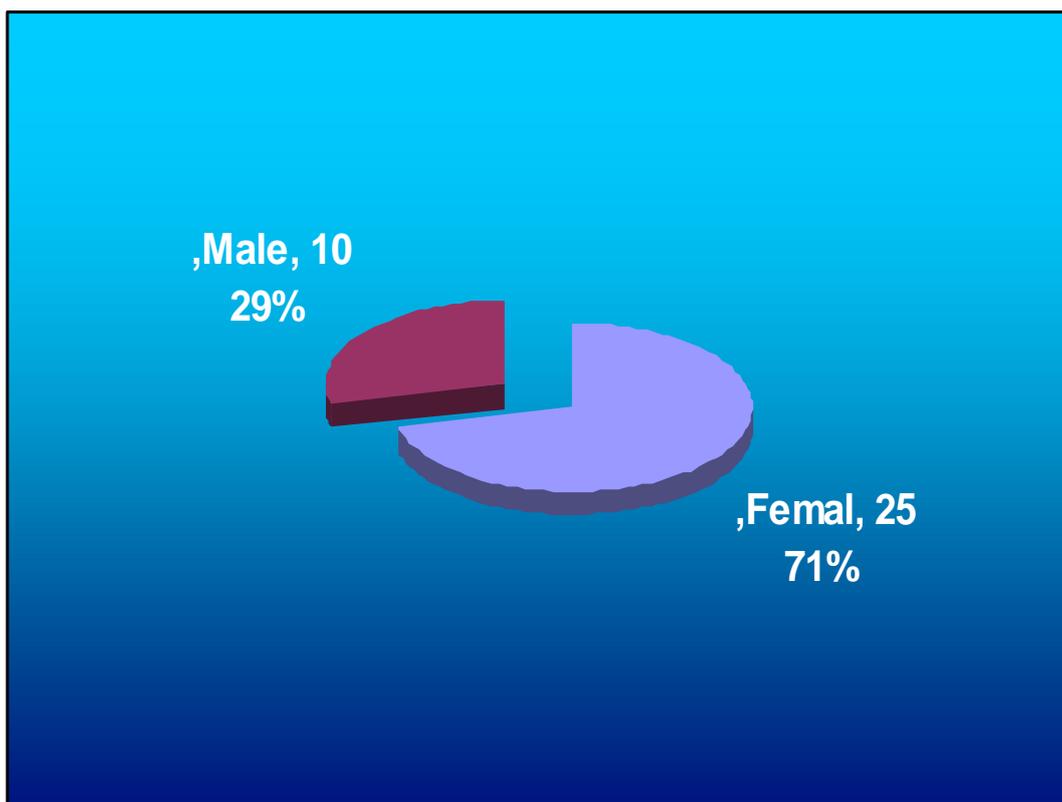


Figure 3.3: The Ratio of Male and Female in Positive Chronic Sinusitis.

3-5 Isolation of Bacteria Associated with Sinusitis:-

The study included 100 patients suffering from sinusitis who attend to Hilla Teaching Hospital-Department of Ear Nose and Throat (ENT). The results showed that (80%) of patients from both chronic and acute sinusitis revealed positive bacterial cultures as mentioned in Table (3-3).

As mentioned above, in chronic sinusitis the positive culture was in 30 cases (83.3%), while the negative culture was only in 6 cases (16.7%). These results were in accordance with the results obtained by Hartog *et.al.* (1990) who found that the positive cultures were as high as (81%) versus the negative cultures which were (19%). However the results of the study were not agreed with the results obtained by Aneke and Ezeanoluc

(200%) since they found that (9%) of the cases revealed negative cultures and (91%) were positive for bacterial culture.

Table (3-3) shows that (86.2%) of the patients in acute sinusitis have positive culture and (13.8%) of them had negative culture. These results were similar to the results obtained by Penttila *et.al.* (1997). Some specimens of both chronic and acute sinusitis cases, being investigated, revealed negative bacterial culture. In chronic sinusitis, this could possibly be due to anaerobic microorganisms which colonizes the blocked sinus providing a suitable environment for them mainly *Bacteroides spp.*, since they are described as the main causative agent of microorganisms in chronic sinusitis patients (Brook, 1996). Unfortunately, the study focused mainly on aerobic and / or facultative anaerobes since anaerobes fastidious organisms are difficult to be isolated from infections sites and are often overlooked (Brook, 2002). The negative results of bacterial cultures can also be attributed to other microbial causes of sinusitis are not included in this study, such as viral or fungal infections.

Table (3-3) Number and Percentage of Sinusitis Culture Result

(Acute and Chronic):

Culture	Acute No. (%)	Chronic No.(%)
Positive	50 (86.2)	30(83.3)
Negative	8 (13.8)	7(16.7)
Total	58 (100)	37(100)

The study detected 38 isolates of *Staph. aureus*, 15 isolates from acute sinusitis and 23 isolates from chronic sinusitis. These isolates were

subjected to sensitivity test against Oxacillin for detecting the Methicillin Resistant *Staphylococcus aureus* (MRSA) as these isolates are more resistant to antibiotics and more aggressive compared with Methicillin Sensitive *Staphylococcus aureus* (MSSA). Consequently the MRSA isolates 11 isolates (29 %) from both acute and chronic sinusitis versus 27 (71%) isolates are accounted for Methicillin sensitive *Staphylococcus aureus* (MSSA). There were 9 (37.0%) isolates from chronic sinusitis of MRSA and only 2 isolates (14.2%) of MRSA from acute sinusitis.

These results can be compared with the results obtained by Manarey *et.al.* (2004) who found that only (9.22%) of MRSA cause chronic rhinosinusitis where as Liu *et.al.* (2001) found that (96.2%) of MRSA were detected in chronic maxillary sinusitis.

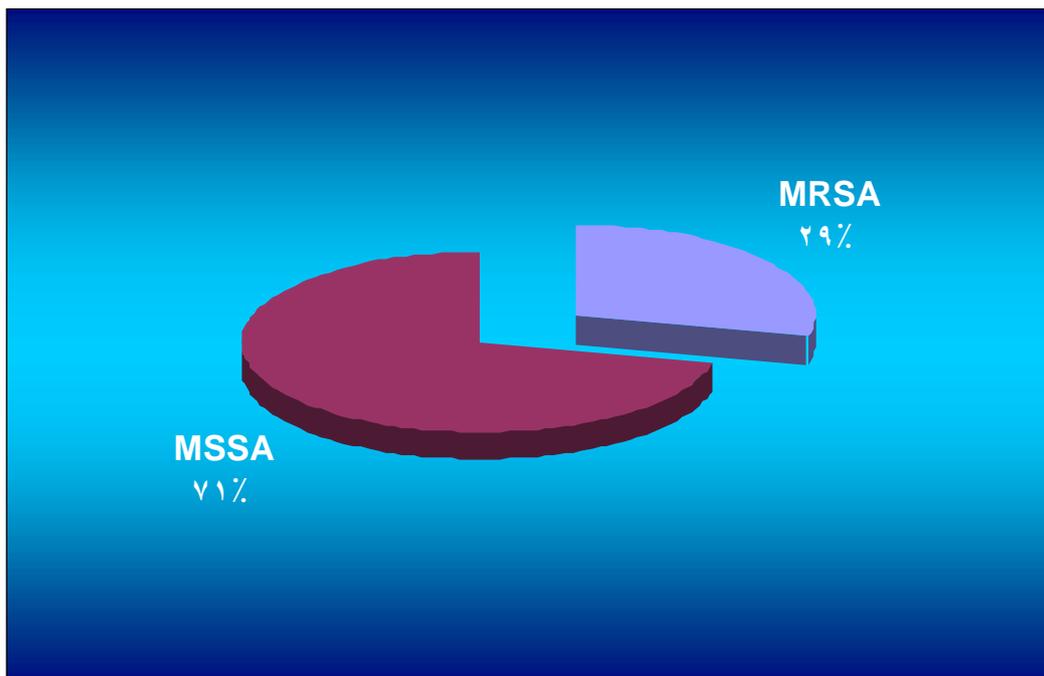


Figure 3.4: The Ratio of MRSA and MSSA in Sinusitis Disease.

3-6 Types of Bacterial Isolates

Table (3-4) shows that the most common organism being cultured from acute sinusitis patients was *S. pneumonia* (20%) followed by *M. catarrhalis* (18%), *S. aureus* (16.7%), Coagulase negative Staphylococci

(14%), Viridans streptococci (12%), *K. pneumonia* (8%) and lastly (6%) isolates of *S. pyogenes* were also obtained.

The results of the present study agreed with the results obtained by Diaz and Bamberger (1990); Brook and Frazier (2004) who showed that *S. pneumoniae* was the most common organism being isolated for sinusitis 20% followed by *M. catarrhalis* (18%) and *S. aureus* (16.7%), Coagulase negative Staphylococci (12%), Viridans streptococci (12%), *K. pneumoniae* (8%) with (0%) isolates of *S. pyogenes* furthermore Brook and Frazier (2005) stated that the most common organism causing acute sinusitis was *S. pneumoniae*. While other studies reported that *Haemophilus spp.* was the most predominant pathogen in acute sinusitis followed by *S. pneumoniae* (Kinman *et.al.*, 1967; VanCauwenberg *et.al.*, 1976; Hannel *et.al.*, 1988; Hannel *et.al.*, 1989 and Sarmad, 2005). Pfaller *et.al.* (2001) stated that the predominant type of bacteria being isolated from acute sinusitis was *M. catarrhalis* followed by *Haemophilus spp.* and *S. pneumoniae*.

Table (3-4) Number and Percentage of Acute Bacterial Isolates:

Bacteria	Number	Percentage %
<i>S. pneumonia</i>	21	20
<i>M. catarrhalis</i>	18	18
<i>S. aureus</i>	14	16.7
<i>S. epidermidis</i>	12	12
<i>S. viridans</i>	12	12
<i>K. pneumonia</i>	8	8
<i>S. pyogenes</i>	0	0
Total	84	100

H. influenzae was reported to be the commonest isolate detected in sinusitis followed by *S. pneumoniae*, *S. viridans*, and *K. pneumoniae* Moughthong *et.al.* (۲۰۰۵), while the study done by Brook (۲۰۰۵) found that the predominant cause of sinusitis was alpha hemolytic Streptococci followed by microaerophilic Streptococci and *S. aureus*. The variation in the causative agents of acute sinusitis is either due to the effect of antibiotics therapy on eliminating the sensitive bacteria allowing for the increasing predominance of other (Berg *et.al.*, ۱۹۸۸), or may be related to microorganisms geographical distribution and even may be related to patients conditions (Conrad *et.al.*, ۲۰۰۲).

Table (۳-۵) shows that the most common organisms isolated from chronic sinusitis patients were *S. aureus* (۴۲.۸%), *S. pneumoniae* (۲۳.۴%), *M. catarrhalis* (۱۰.۷%), *K. pneumoniae* (۹%), Viridans streptococci (۵.۳%), *E. coli* (۵.۳%) and *Proteus mirabilis* (۳.۵%). These results were in agreement with the results obtained by Sener *et.al.* (۱۹۹۶) who pointed out that the most common organism in chronic sinusitis was *S. aureus* (۶۰%), followed by *S. pneumoniae* (۴۰%), *M. catarrhalis* (۳۰%), *K. pneumoniae* (۲۵%), Viridans streptococci (۱۵%), (۶%) isolates of *E. coli* and (۴%) of *Proteus mirabilis*. *S. aureus* was reported to be the dominant organisms habitant chronic sinusitis followed by *H. influenzae*, *E. coli* and *S. pneumoniae* (Namyslowski *et.al.*, ۲۰۰۴), while Sarmad (۲۰۰۵) found that the most predominant microorganisms isolated from chronic sinusitis was *H. influenzae*, *S. pneumoniae*, *M. catarrhalis*, *S. aureus*, Viridans streptococci and then some members of Enterobacteriaceae.

Consequently, *S. aureus* seemed to be the common dominant among bacteria resident in chronic sinusitis. This phenomenon may be explained by Bukharin *et.al.* (۱۹۹۸) who stated that the biological properties of *Staphylococci* initiating the inflammation with feature of clinical course of maxillary sinusitis. These virulent characteristics determine the acute

course of the disease whereas the strain with persistent properties causes chronic course of maxillary sinusitis.

Table (३-०) Number and Percentage of Chronic Bacterial

Isolates:

Type of Bacteria	Number	Percentage %
<i>S. aureus</i>	२६	६२.८
<i>S. pneumoniae</i>	१३	२३.६
<i>M. catarrhalis</i>	६	१०.७
<i>K. pneumoniae</i>	०	१
<i>E. coli</i>	३	०.३
<i>S. viridans</i>	३	०.३
<i>Proteus mirabilis</i>	२	३.०
Total	०६	१००

३-१ Seasonal Variation :-

The results shown in Table (३-१) indicate that the occurrence of sinusitis through the year showed an accumulation of cases during the cold season since the peak of incidences was observed in January.

Table (३-१) Number of Positive Obtained Cases Distributed on Months of the Year

years	Months	No. of Cases
२००६	October	६
	November	६
	December	१२
२०००	January	१७
	February	१२
	March	११
	April	१२
	May	७
	June	२

	July	२
	Total	१०

From these results one can conclude that sinusitis occurs as a secondary infection due to sinus ostia obstruction associated with the common cold or allergies and maxillary sinus is the most common (Diaz and Bamberger, 1990). Most cases of common cold usually occur in temperate climate, so it appears in winter more than in summer (Macky and Bull, 1997). These results are also ensured by (Glezen, 1998) who stated that the obstruction in nose due to infection with Rhinovirus or other virus that cause common cold will trap bacteria in the closed space resulting in suppurative sinusitis and otitis media which are the most common complication of URTI (upper respiratory tract infection). The results obtained by this study are in accordance with the results obtained by Stalman *et.al.* (2001) who asserted that sinusitis cases are common in winter rather than in other seasons since (90%) of sinusitis cases were recorded in winter. Statistical analysis by using Chi Square showed that there is a significant relationship between months and number of cases ($p < 0.05$).

३-१ Susceptibility of Bacterial Isolates to Antibiotics:-

For more explanation the effect of different antibiotics on bacterial isolates being identified in this study will now be discussed:-

३-१-१ Penicillins

३-१-१-१ Ampicillin

Figure (3-0) shows that *M. catarrhalis*, *S. aureus* (both MRSA and MSSA) and *Proteus mirabilis* were fully resistant (100%) to ampicillin. These results agree with the results obtained by (Kim *et.al.*, 2004; Rashmi *et.al.*, 2000 and Melo-Cristino *et.al.*(2006) who observed that these bacteria were highly resistant to ampicillin. The resistance frequency

exhibited by *S. pneumoniae* and viridans group streptococci are accounted for (53%) and (69%) respectively while *S. pyogenes* isolates were sensitive. These results were more agreeable with the results obtained by Hsuch *et.al.* (2004) who found that the resistance rate of *S. pneumoniae* to ampicillin was (50%) while it was (60%) for viridians group streptococci whereas *S. pyogenes* was sensitive. Two isolates of *E. coli* (66%) and 9 isolates of *K. pneumoniae* (50%) were resistant to ampicillin. These results were in accordance with the results obtained by Oteo *et.al.*(2005) who found that (59.9%) of *E. coli* were resistant to ampicillin. Moreover Blandino *et.al.*(1990) found that (50%) of *K. pneumoniae* were resistant to ampicillin. 66% of *Staph. epidermidis* was found to be resistant to ampicillin. Hashemi *et.al.* (2005) reported that *S. epidermidis* was less resistant compared with *S. aureus* isolates.

3-8-1-2 Amoxicillin

Figure (3-6) shows that (100%) of *Proteus mirabilis*, (50%) of *E. coli* and (50%) of *K. pneumoniae* were resistant to amoxicillin. These results were in agreement with the results obtained by Quentin *et.al.* (2004) who stated that (53.4%) of *E. coli* was resistant. Stock (2003) stated that *Proteus mirabilis* was naturally resistant to amoxicillin and Toy *et.al.* (1993) who found that (58.6%) of *K. pneumoniae* were resistant to this drug.

The susceptibility of *S. pneumoniae* and *S. pyogenes* are accounted for (100%). These results were in accordance with the results obtained by Vanhoof *et.al.*(2005) who ensured that most of bacterial isolates remained fully susceptible to amoxicillin. Pichichero (1995) found that *S. pyogenes* were highly susceptible to amoxicillin, while (30%) of Viridans streptococci were resistant to amoxicillin. These results were comparable with the results obtained by IKemoto *et.al.* (1996) who found that (40%) of viridians group streptococci were resistant to amoxicillin. All isolates of MRSA (100%) versus (50%) of MSSA were resistant to amoxicillin. These results agreed with the results obtained by Rajaduraipandi *et.al.*(2006)

who obtained that (100%) of MRSA isolates were resistant to amoxicillin. Moreover Kingdom and Swain (2004) pointed out that (98.9%) of MSSA were resistant to amoxicillin. The high resistance for beta-lactam drugs could be explained by the production of beta-lactamases by Staphylococci, gram positive bacteria, gonococci and others, moreover lack of penicillin binding protein (PBPs) or altered (PBPs) in Pneumococci. Sometimes failure of activation of autolytic enzyme in cell wall can result in inhibition without killing bacteria ex. Tolerance of some Staphylococci (Brooks *et.al.*, 2004)

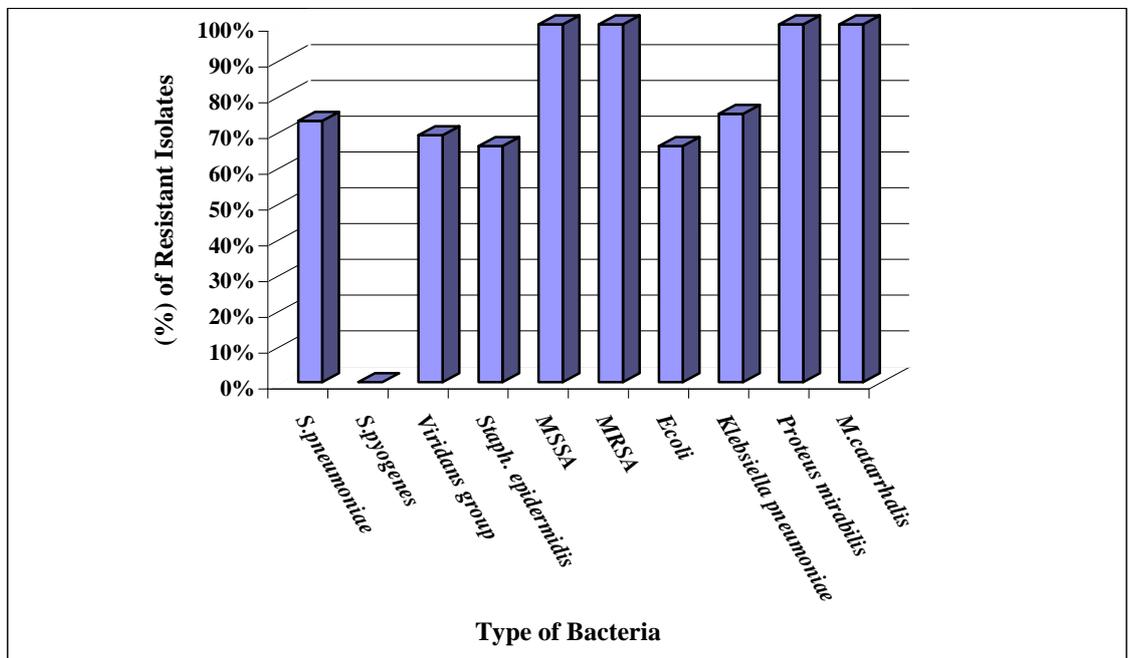


Figure 3.9: The Ratio of Resistant of Bacterial Isolates Against Ampicillin.

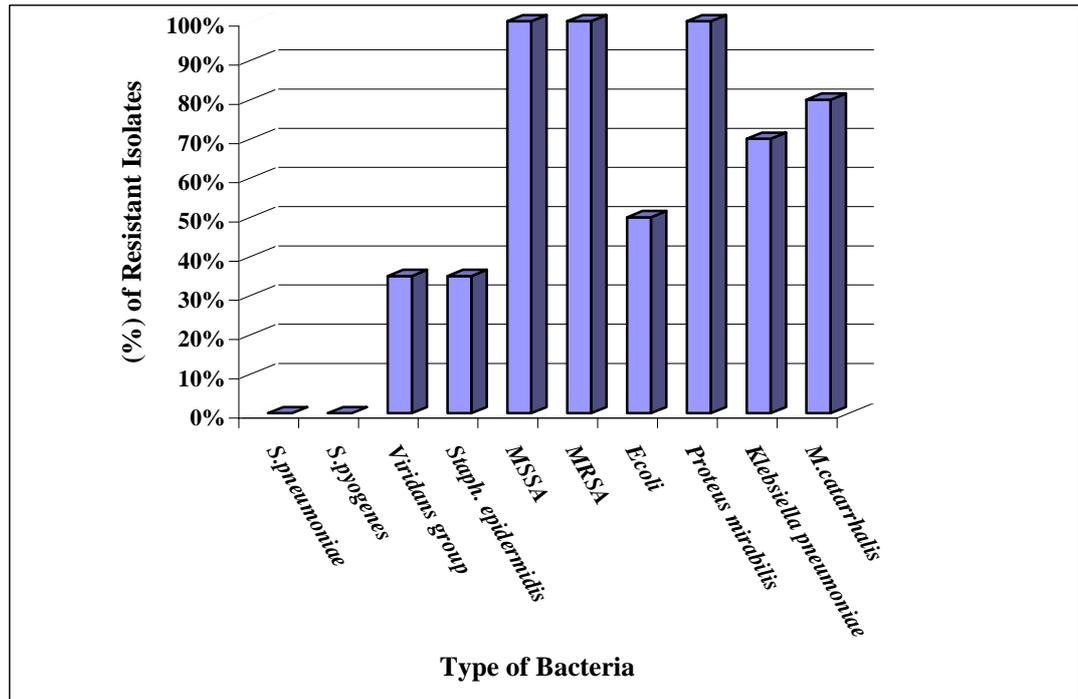


Figure 3.6: The Ratio of Resistant of Bacterial Isolates Against Amoxicillin.

3-8-2 Aminoglycosides

3-8-2-1 *Gentamycin*

The results shown in Figure (3-7) indicate that only four types of bacterial isolates in this work were fully sensitive to gentamycin .These bacteria were *S. pyogenes*, *K. pneumoniae*, *E. coli* and *Proteus mirabilis*. These findings correlated with other results being observed by other studies, since *S. pyogenes* exhibited sensitivity rate up to (100%) Zhanel *et.al.* (2002) while the sensitivity rate of *E. coli* and *Proteus mirabilis* to gentamycin was (90%) and more than (90%) of *K. pneumoniae* were also sensitive to this drug (Shavidok, 1987; Petrove *et.al.*, 2000 and Tonkic *et.al.*, 2000). Moreover other types of bacterial isolates exhibited a percentage of resistance ranged from (20%) for *Staph. epidermidis* until they reached (80%) for *M. catarrhalis*. (76%) for Viridans group, (50%)

for *S. pneumoniae*, (46.1%) for MSSA and (66%) for MRSA. These results were in accordance with the results obtained by Hsuch *et.al.* (2004) who ascertained that (8%) of *S. pneumoniae* were resistant to gentamycin. Rashmi *et.al.*(2009) and Smith *et.al.*(2002) showed that Viridans group Streptococci were highly resistant to gentamycin. In addition to that Adwan *et.al.*(2009) mentioned that (64.3%) of MRSA were resistant to gentamycin while Rohani *et.al.*(2000) observed that (40.5%) of MSSA were resistant to gentamycin. Deng *et.al.* (1981) revealed that (8%) of *Staph. epidermidis* were sensitive for this drug. The production of aminoglycoside modifying enzyme is the most important mechanism of its resistance commonly due to plasmid transfer but the alteration in cell wall permeability is another reason for resistance, especially in Streptococci. This mechanism of resistance was described as chromosomally mediated mechanism (Mims *et.al.*, 2004).

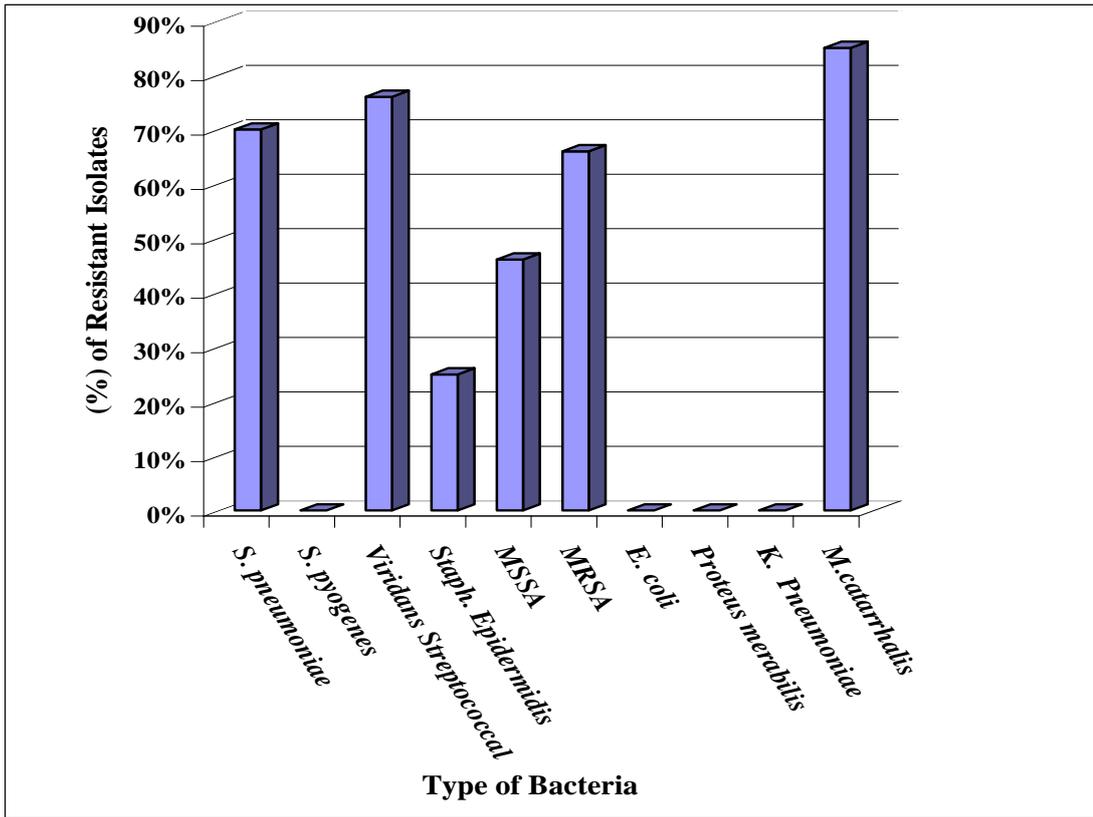


Figure ۳.۷: The Ratio of Resistant of Bacterial Isolates Against Gentamycin

٣-٨-٣ Fluoroquinolon

٣-٨-٣-١ *Ciprofloxacin*

Figure (٣-٨) shows that all types of bacterial isolates found in this test were fully susceptible for ciprofloxacin except (٥٨.٣%) of MRSA, (٢٣%) of MSSA and (١٢%) of *M. catarrhalis* were resistant to ciprofloxacin. These results agreed with the results pointed out by other studies. Tonkic *et.al.*(٢٠٠٥) found that (٤.٤%) of *E. coli* and (٤%) of *K. pneumoniae* were resistant to ciprofloxacin; Brown and Mjrybak (٢٠٠٤) stated that all isolates of *S. pyogenes* were sensitive to ciprofloxacin and more than (٩٧%) of *S. pneumoniae* were sensitive to ciprofloxacin. Inoue *et.al.*(٢٠٠٦) reported that viridians groups streptococci were highly sensitive to ciprofloxacin and Hsuch *et.al.*(٢٠٠٤) observed that (٩٠%) of *M. catarrhalis* were sensitive to ciprofloxacin while Rohani *et.al.*(٢٠٠٠) stated that (٢٩.٢%) of MSSA were resistant to ciprofloxacin. Moreover Adwan *et.al.*(٢٠٠٥) detected that (٣٢.١%) of MRSA were resistant to ciprofloxacin.

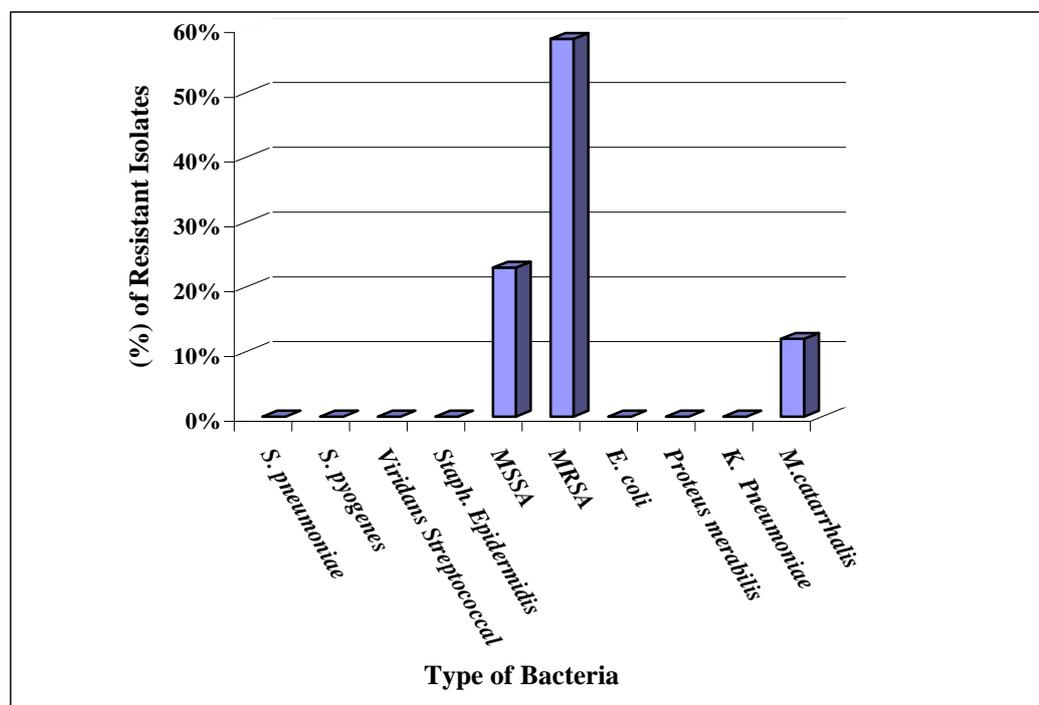


Figure ٣.٨: The Ratio of Resistant of Bacterial Isolates Against Ciprofloxacin.

3-1-4 Cephalosporins

3-1-4-1 *Cefotaxim*

As shown in Figure (3-9) only MRSA, *Proteus mirabilis* and *Klebsiella pneumoniae* exhibited relative resistance represented by (83%), (50%) and (20%) respectively. While all other bacterial types included in this test were fully (100%) sensitive for cefotaxim. These results came to confirm the results being reported by other studies, since Morrissey *et.al.* (2005) found that all isolates of *S. pneumoniae* and *M. catarrhalis* were sensitive to cefotaxim. Mendes *et.al.* (2004) stated that viridans group streptococci were highly susceptible to cefotaxim. Karlowsky *et.al.*, (2004) reported that *E. coli* were sensitive to cefotaxim. Kim *et.al.* (2004) found that a high percentage of *P. mirabilis* was sensitive to cefotaxim. Balandino *et.al.* (1990) and Lee *et.al.*, (2001) mentioned that (19%), (20%) of *K. pneumoniae* were resistant to cefotaxim respectively. Deneg *et.al.* (1981) found that all isolates of *S. epidermidis* and MSSA were sensitive to cefotaxim. *S. pyogenes* was found to be fully susceptible for cefotaxim (Brown and Mjrybak, 2004). Regarding the resistance of MRSA towards cefotaxim in this study, it was quietly in accordance with those results which have been recently reported by Asghar and Momenah (2006) who determined the resistance of MRSA against this antibiotic as high as (80%).

3-1-4-2 *Cephalothin*

The results show in Figure (3-10) the percentage of resistance represented by (13.3%) for *M. catarrhalis* (50%) for *K. pneumoniae* (66%) for *E. coli*, (50%) for *P. mirabilis* and (66%) for MRSA. All other bacterial isolates in this study were sensitive. These results were agreeable with those results obtained by Giglio *et.al.* (1999) who

revealed that the gram positive cocci was highly susceptible to Cephalothin. Stock (۲۰۰۳) observed that moderate percentage of *proteus mirabilis* was resistant to Cephalothin. Yeh and Chi (۲۰۰۱) noticed that (۷۲%) of *E. coli* were resistant to Cephalothin. Furthermore Inglis *et.al.* (۱۹۹۴) stated that (۷۰%) of *K. pneumoniae* were resistant to Cephalothin. Additionally, Morrissey *et.al.* (۲۰۰۵) reported that *M. catarrhalis* and *S. epidermidis* were highly sensitive to Cephalothin. In addition, Rohani *et.al.*(۲۰۰۰) found all strain of MSSA was susceptible to Cephalothin and Durmaz *et.al.*(۱۹۹۷) obtained that (۶۰%) of MRSA were resistant to Cephalothin. Antibiotics resistance is linked to prior exposure to antibacterial drugs because the first and second generation of cephalosporins are commonly prescribed for hospitalized patients (Liu *et.al.*, ۱۹۹۹). Resistance to cephalosporins is due to special β -lactamases in gram negative bacteria and *Staphylococci* that hydrolyze and inactivate these drugs (Brooks *et.al.*, ۲۰۰۴).

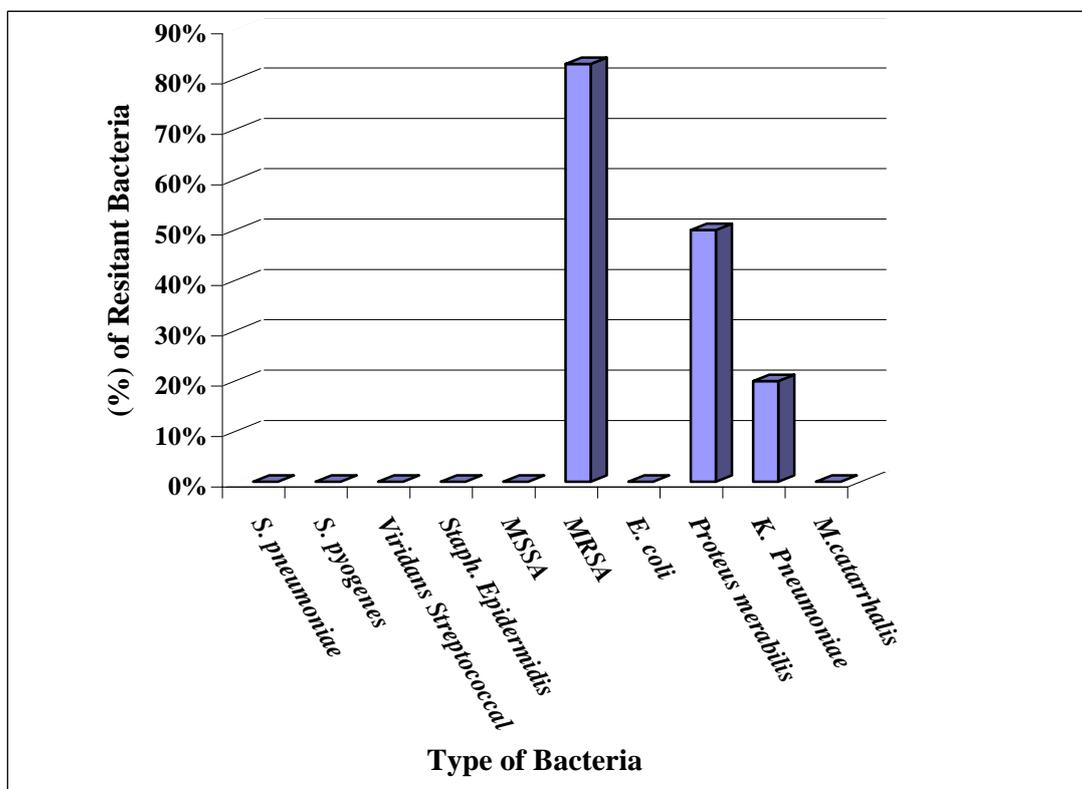


Figure ۳.۹: The Ratio of Resistant of Bacterial Isolates Against Cefotaxim. `

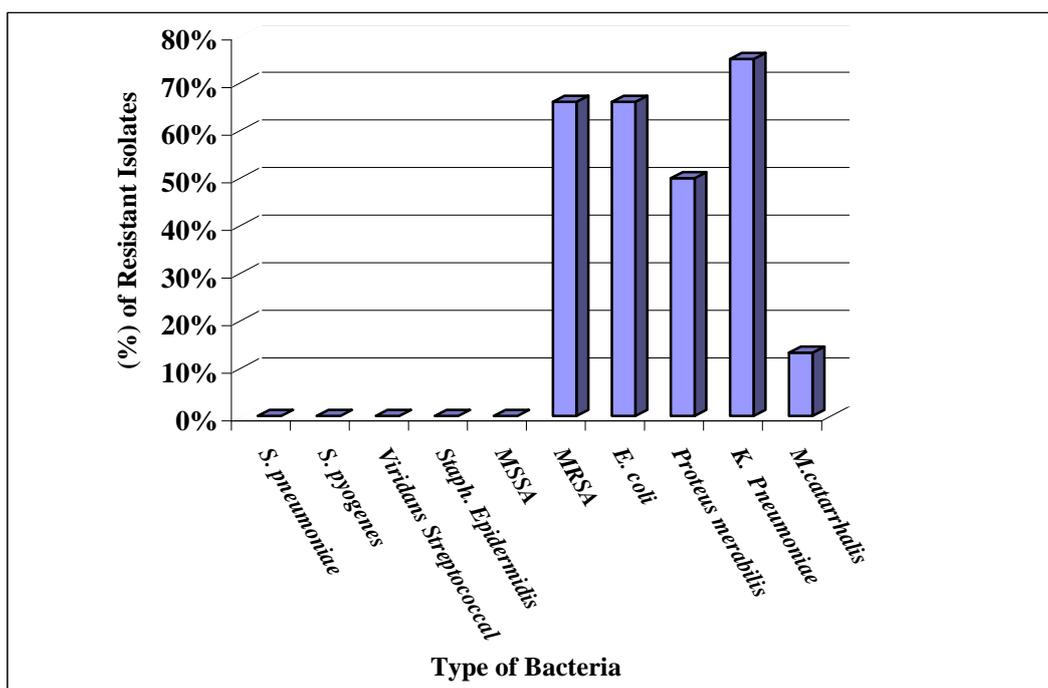


Figure 3.10: The Ratio of Resistant of Bacterial Isolates Against Cephalothin.

3-8-0 Sulfonamide derivative

3-8-0-1 *Trimethoprim*

The results shown in Figure (3-11) indicate that *S. pyogenes* and *Staph. epidermidis* revealed no resistance, while the other types of investigated bacteria exhibited a percentage of resistance ranged from (13.3%) for *K. pneumoniae* up to (91%) for MRSA. A moderate resistance was observed in MSSA and *P. merabilis*. These results were in accordance with Melo-Cristino *et.al.* (2006) who found that (27.1%) of *M. catarrhalis* were resistant and Cardozo *et.al.* (2006) who found that (7.3%) of *S. pneumoniae* were resistant to trimethoprim. It was found that *P. merabilis* resisted Trimethoprim in a frequency of (00%) (Kim *et.al.*, 2004), while *E. coli* resisted Trimethoprim in a frequency of (21%)

(Quentin *et.al.*, 2004). *K. pneumoniae* was observed to be resistant to Trimethoprim in a frequency at (13.3%) (Tonkic *et.al.*, 2009). Rohani *et.al.*(2000) stated that (47.2%) of MSSA were resistant while Pulimood *et.al.*(1996) found that (97%) of MRSA were resistant to trimethoprim. Moreover Rajaduraipandi *et.al.*(2006) obtained (63.2%) of MRSA isolates which were resistant towards Trimethoprim. The resistant to this antibiotic emerged to previous longer use of it leading to reduce cell permeability and pumping it from the resistant bacterial cell (Murray *et.al.*, 1999).

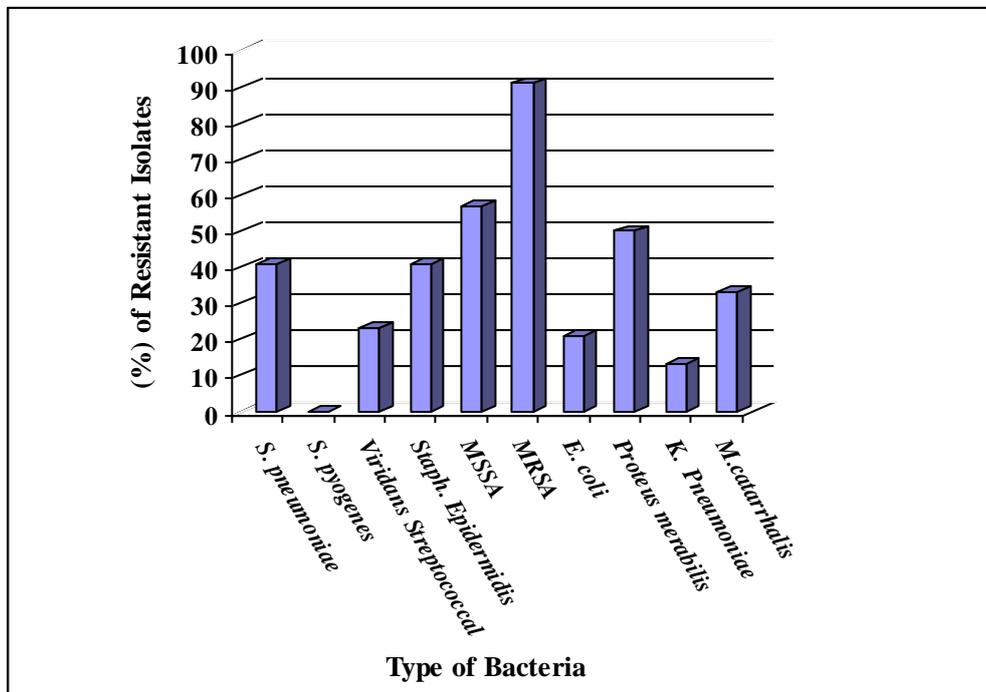


Figure 3.11: The Ratio of Resistant of Bacterial Isolates Against Trimethoprim.

3-7-6 Vancomycin

As shown in Figure (3-12) all types of gram positive isolates were sensitive towards vancomycin, while all the negative isolates were resistant to it. These findings were closely correlated with the results being reported by Cardozo, *et.al.* (2006); Ergin *et.al.* (2006) and Inoue

et.al. (۲۰۰۶) who pointed out that *S. pneumoniae*, *S. pyogenes* and Viridians group streptococci were highly susceptible to vancomycin .Moreover these results Matched the reports of Luh *et.al.*, (۲۰۰۰) and Hamze *et.al.* (۲۰۰۳) who showed that all isolates of MSSA and MRSA were susceptible to vancomycin .In addition Mendes *et.al.*, (۲۰۰۳) stated that *S. epidermidis* was highly susceptible to vancomycin.

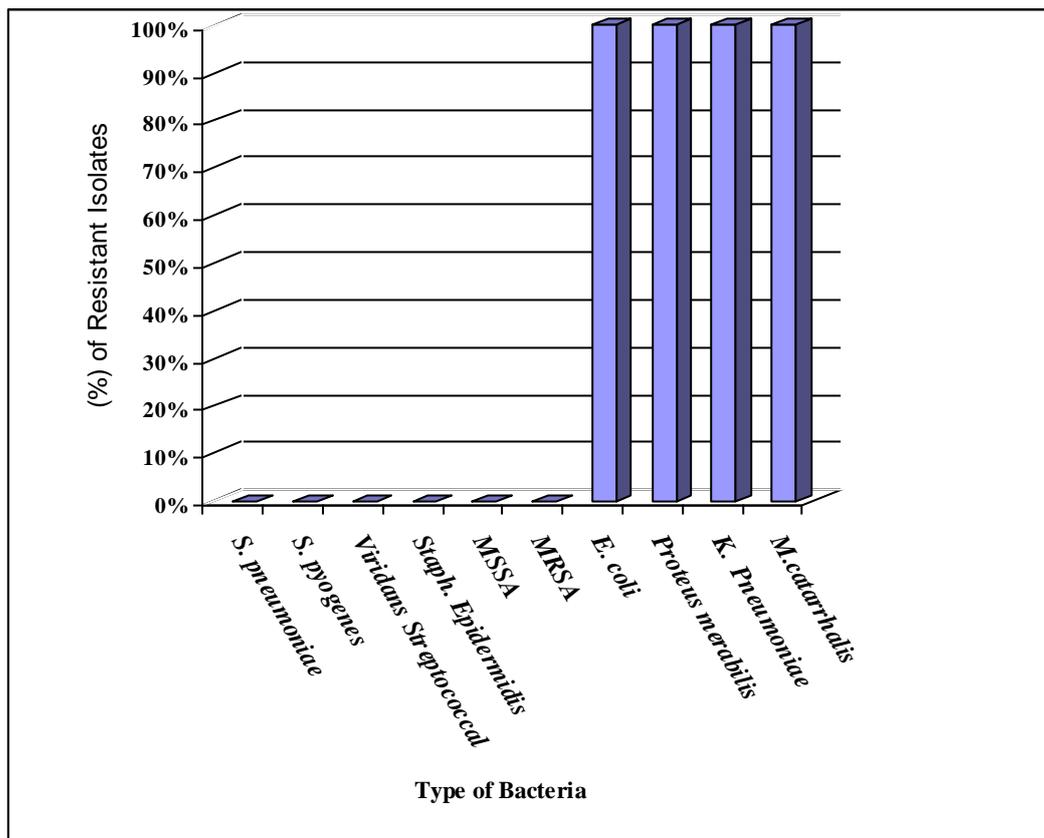


Figure ۳.۱۲: The Ratio of Resistant of Bacterial Isolates Against Vancomycin.

3-8-7 Tetracyclines

3-8-7-1 *Tetracyclin*

The results shown in Figure (3-13) indicate that only two types of bacterial isolates represented by *S. pyogenes* and *S. epidermidis* were fully (100%) sensitive for tetracycline. Other types of bacterial isolates showed different rates of resistance ranging from (23%) for Viridians group streptococci up to (100%) for enteric group represented by (*Proteus mirabilis*, *E. coli* and *K. Pneumoniae*). On the other hand, the resistance rate against tetracycline for *S. pneumoniae* was (29%), *M. catarrhalis* (23.8%) and (46.1%) of MSSA isolates and (91.6%) of MRSA isolates. These results were in accordance with the results obtained by Stock (2003) who found that *Proteus mirabilis* was naturally resistant to all tested tetracycline. Ansari and Khatoun (1999) reported that *E. coli* was resistant against tetracycline, while *K. pneumoniae* showed no response towards tetracycline (Toy *et.al.*, 1993). Moreover Vanhoof *et.al.* (2000) mentioned that (30.0%) of *S. pneumoniae* were resistant to tetracycline, while Inoue *et.al.* (2006) expressed 30% of Viridians group streptococci were resistant to tetracycline and all isolates of *S. pyogenes* were sensitive. MSSA resisted to tetracycline in a frequency of (44.2%) Inoue *et.al.* (2006), while MRSA resisted to tetracycline in a frequency of more than (90%) (Rohani *et.al.*, 2000 and Asghr and Momenah 2006). The resistance of bacteria towards this antibiotic could be explained by efflux pump by an active transport protein pump (Jacopy, 1993).

3-1-1 Macrolides

3-1-1-1 *Erythromycin*

As shown in Figure (3-14), all types of bacterial isolates investigated by this study showed resistance to erythromycin in variable rates starting from (17%) for *S. pneumoniae* and (20%) for *S. pyogenes* up to (100%) for the members of Enterobacteriaceae represented by (*proteus mirabilis*, *E. coli* and *K. pneumoniae*). Moreover (83.3%) of MRSA isolates were resistant to erythromycin, while a moderate resistance rate was observed in MSSA, *S. epidermidis* and Viridans group streptococci. Consequently these results can be compared with the results obtained by Melo-Cristino *et.al.* (2006) who showed that (18.8%) of *S. pneumoniae* and (18.9%) of *S. pyogenes* were resistant to erythromycin. Moreover Mendes *et.al.* (2003) stated that (32.6%) of viridans group streptococci were resistant to erythromycin. Additionally, Rohani *et.al.* (2000) revealed that only (20.9%) of MSSA isolates were resistant to erythromycin and the results obtained by Asghar and Momenah (2006) mentioned that (82.1%) of MRSA isolates were resistant to erythromycin. Toye *et.al.* (1993), Zhang *et.al.* (2006) and Stock (2003) also obtained such results. Moreover, *M. catarrhalis* resisted to erythromycin in frequency of (13%) (Inoue *et.al.*, 2006), while *S. epidermidis* resisted to this antibiotic in a frequency of (30%) (Pichiche, 1990). The resistance of gram positive bacteria for these antibiotics resulted by the production of Erythromycin-resistant methylase resultant structural changes to rRNA prevent macrolide binding and allows synthesis of bacterial proteins to continue or by efflux (Pechere, 2001), while the resistance of gram negative bacteria particularly (Enterobacteriaceae) can be ascribed to the production of esterase that hydrolyze macrolides (Chambers, 2001).

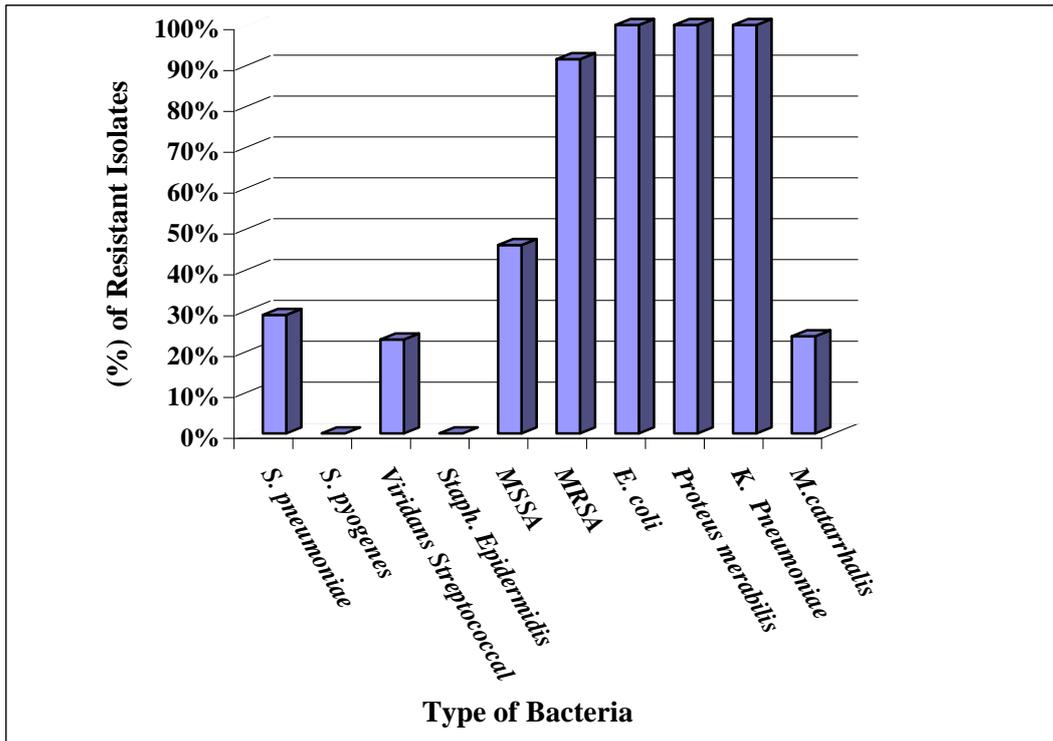


Figure 3.13: The Ratio of Resistance of Bacterial Isolates Against Tetracycline.

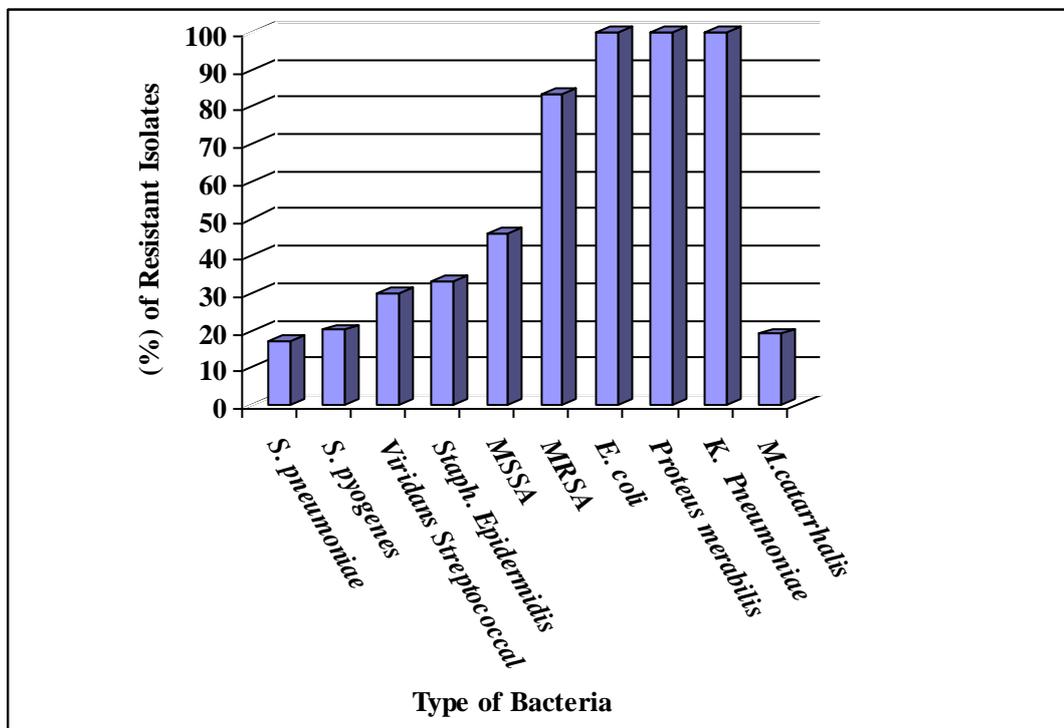


Figure 3.14: The Ratio of Resistant of Bacterial Isolates Against Erythromycin

Conclusions & Recommendations

Conclusions:-

According to this study, the following could be concluded :

- ١- ٢٠-٢٩ years old persons are more suffering form acute and chronic sinusitis than other age groups.
- ٢- Females are more exposed for acute and chronic sinusitis.
- ٣- Nasal septal deviation and /or nasal polyps are predisposing for chronic sinusitis.
- ٤- Climatic allergens, food allergens, environmental allergens and common cold are mostly resulting in sinusitis.
- ٥- The proportionally increase in Methicillin Resistance *Staphylococcus aureus* (MRSA) is a serious risk for sever nosocomial infections.
- ٦- The most common type of bacterial isolates which isolated from acute sinusitis which were *S. pneumoniae* followed by *M. catarrhalis*, *S. aureus*, *S. epidermidis*, Viridans streptococci, *K. pneumoniae* and *S. pyogenes* .
- ٧- The most common type of bacterial isolates which isolated from chronic sinusitis which were *S. aureus* followed by *S. pneumoniae*, *M. catarrhalis* *K. pneumoniae*, *E. coli*, Viridans streptococci and *Proteus mirabilis*

Recommendations:-

According to this study, the following could be recommended:

- ١-For proper diagnosis of the microbial etiology of sinusitis, culturing of sinus specimens is quietly necessary.
- ٢-Properly picking up the nasal swabs or maxillary sinus aspiration for laboratory diagnosis of the causative agents.
- ٣-Management of the local predisposing causes such as polyp or nasal septal deviation which affect the normal sinus physiology.
- ٤-Ciproflaxacin and cefotaxim can be recommended as antimicrobial therapy for sinusitis.
- ٥-Prospective studies can be conducted to detect the role of microorganisms in sinusitis complications, the physical characters of sinus secretions and organisms causes sinusitis.
- ٦-Immunological studies could also be conducted to detect the level of serum antibody like IgA and IgG .

Appendices

Case Sheet Used in This Study

Pt. name	Sex:	Age	Address	date	
Preliminary clinical diagnosis					
Associated history	A. common cold	b. Allergy	c-Polyp	D. Nasal septal deviation	E. Dental problem
F. Diabetes mellitus	J. Per orbital pain	H. Fever	I. Nasal obstruction	X ray finding and CT- Scan	
Type of sample	A. Nasal swab	B. Sinus antral wash			
Notes:-					

References

- Abuhammour ,W.M., Abdle-Haq N.M., Asmar B.I. and Dajani A.S.(١٩٩٩). *Moraxella catarrhalis* bacterimia: a ١٠ year experience .South. Med. J. ٩٢(٦١):١٠٧١-١٠٧٤.
- Adwan ,A., Abu-Hasen N., Adwan G., Jarrar N. and Zant A.(٢٠٠٥). Nosocomial infection caused by methicillin-resistant *Staphylococcus aureus* in Palestine. Microb. Drug. Resist., ١١(١):٧٥-٧٧.
- Almadori ,G., Bastianini L., Bistoni F., Maurizi M., Ottaviani F., Paludetti G. and Scuteri F.(١٩٨٦). Microbial flora of nose and paranasal sinuses .Rhinology, ٢٤(٤):٢٥٧-٢٦٤.
- Aneke ,E.C. And Ezeanolue B.C.(٢٠٠٤). Profile of aerobic bacteria isolated in chronic maxillary sinusitis patients. Niger. Postgrad. Med. J., ١١(٢):١١٦-١٢٠.
- [Ansari ,F.A.](#), [Khatoon H.](#)(١٩٩٩). A survey of antibiotic resistance among *E. coli* strains isolated from poultry in Karachi. [Pak. J. Pharm. Sci.](#), ١٢(١):٧-١٤.
- [Asghar ,A.H.](#) And [Momenah A.M.](#)(٢٠٠٦). [Methicillin resistance among *Staphylococcus aureus* isolates from Saudi hospitals. Med. Princ. Pract.](#), ١٥(١):٥٢-٥٥.
- Axelsson ,A. and Brorson J.E.(١٩٧٣). The correlation between bacteriological findings in the nose and maxillary sinus in acute maxillary sinusitis. Laryngoscope, ٨٣:٢٠٠٣-٢٠١١.
- Baily ,QR.(١٩٨١). Chronic sinusitis .J. Laryngol. Otol., ٩٥(٥٦):٥٥-٦٠.
- Baily ,B., Johnson J. and Kohut R.(١٩٩٣). Head and Neck surgery – Otolaryngology. Nose and paranasal sinuses. ٢nd ed. Philadelphia ,Lippincott. Pp ١٥٣-١٦٠.
- Baron ,E.J. and Finegold S.M.(١٩٩١). Bailey and Scotts diagnostic microbiology. ٨th ed. C.V. Mosby Co., St. Louis, Baltimore, Philadelphia.

- Baron ,E.J., Peterson L.R. and finegold S.M.(١٩٩٦). Bailey and Scotts diagnostic microbiology .٩th ed. C.V. Mosby company.
- Bauer ,A.W., Kirby W.M. Sherris J.C. and Turk M.(١٩٦٦). Antibiotic susceptibility testing by standardized single disk method. Am. J. Clin. Pathol., ٤٥:٤٩٣-٤٩٦.
- Benninger ,M., Anon J. and Marby R.(١٩٩٧). The medical management of rhinosinusitis. Otolaryngology Head Neck surgery ,١١٧(٢٣):٤١-٤٩.
- Berg ,O., Carenfelt C. , Rystedt G. and Enggderd A.(١٩٨٦). Occurrence of asymptomatic sinusitis in common cold and other acute ENT-infections. Rhinology , ٢٤ :٢٢٣-٢٣٠.
- Berg ,O., Carenfelt C. and Kronvall G.(١٩٨٨). Bacteriology of maxillary sinusitis in relation to character of inflammation and prior treatment. Scan. J. Infect. Dis., ٢٠ (٢٣):٥١١-٥١٦.
- Bile ,MD., Carl A., Brown M.D., Richard M., Levinson M.D., Gary E., Garvis M.D., Hyman M., Paisner D., Melvine E., Sigle E., Michil S. and Tedford M.D.(١٩٩٨). Evaluation the microbiology of chronic maxillary sinusitis. Ann. Otol. Rhinol. Laryngol. ,٥٦(١١):٩٤٢-٩٤٥.
- Bjorkwall ,T.(١٩٥٠). Bacteria and inflammatory cell in maxillary sinusitis . Acta. otolaryng .Stockh., ٢٣٩(٢):١٧٣-١٨٠.
- Blandino ,G., Caccamo F., Dimarco R., Speciale A. and Nicoletti G.(١٩٩٠). Epidemiology of antibiotic resistance in human isolates of Enterobacteriaceae in Sicily .J. Cemother., ٢(١):٤٠-٤٤.
- Boyl ,M.D.P.(١٩٩٥). Variation of multifunctional surface binding proteins-A virulence strategy for group A *Streptococci*. J. Theor. Biol. ١٧٣(٢٣):٤١٥-٤٢٦.
- Bridger ,R.C.(١٩٨٠).Sinusitis :An improved regimen of investigation for the clinical laboratory. J. Clin. Pathol., ٣٣:٢٧٦-٢٨١.

- [Brook ,I.](#)(١٩٩٦). Microbiology and management of sinusitis. J. Otolaryngol., ٢٥(٤):٢٤٩-٥٦.
- Brook ,I., Yocum P. and Frazier E.H.(١٩٩٦). Bacteriology and beta-lactamase activity in acute and chronic maxillary sinusitis. Arch Otolaryngol Head Neck Surg., ١٢٢(٤):٤١٨-٤٢٢.
- Brook ,I., Gooch W.M. and Jenkins S.G.(٢٠٠٠). Medical management of acute bacterial sinusitis: recommendation of a clinical advisory committee on pediatric and adult sinusitis. Ann. Otol. Rhinol. Laryngology, ١٠٩:٢-٢٠.
- Brook ,I.(٢٠٠٢). Bacteriology of acute and chronic sphenoid sinusitis. Ann. Otol. Rhinol. Laryngol. , ١١١(١١):١٠٠٢-١٠٠٤ .
- Brook ,I. and Frazier E.H.(٢٠٠٤). Microbiology of recurrent acute rhinosinusitis. Laryngoscope , ١١٤(١):١٢٩-١٣١.
- Brook ,I.(٢٠٠٤). Discrepancies in the recovery of bacteria from multiple sinuses in acute and chronic sinusitis. J. Med. Microbiol., ٥٣(١٠):٨٧٩-٨٨٥.
- Brooks ,G.F., Butel J.S. and Morse S.A.(٢٠٠٤). Staphylococci. Medical Microbiology. ٢٣rd ed. McGraw-Hill Company. Pp ٢٢٣-٢٢٦.
- Brooks ,J.G., Butel J.S. and Morse S.A.(٢٠٠٤). Antimicrobial Chemotherapy. In Medical Microbiology. ٢٣rd ed. Lange. McGraw-Hill Medical Publishing Division. Pp ١٦١-١٩٠.
- Brook ,I. and Frazier E.H.(٢٠٠٥). Microbiology of acute and chronic maxillary sinusitis associated with an odontogenic origin. Laryngoscope , ١١٥(٥):٨٢٣-٥.
- Brook ,I. and Frazier E.H.(٢٠٠٥). Bacteriology of chronic maxillary sinusitis associated with nasal polyposis . J. Med. Microbiol., ٥٤: ٥٩٥-٥٩٧.
- Brown ,S.D. and Mjrybak M.J.(٢٠٠٤). Antimicrobial susceptibility of *Streptococcus pneumoniae*, *Streptococcus pyogenes* and

- Haemophilus influenzae* collected from patients across the USA, in 2001-2002, as part of the PROTEKT US study .J. Antimicrob. Chemother., 54(1): 7-10.
- Buchem ,L., Peter M. and Beaumont J.(1990). Acute maxillary sinusitis in general practice: the relation between clinical picture and objective finding. Eur. J. Gen. Pract., 1:100-106.
- Buchman ,C.A., Doyle W.J. and Skoner D.(1994). Otolgic manifestations of experimental Rhinovirus infection. Laryngoscope, 104(32):1290-1300.
- Bukharin ,O.V., Chernova O.L., Matiushina S.B., Raitselis I.V. and Zabirov R.A.(1998). The association of biological properties of staphylococci with the process of purulent sinusitis. Vestn. Otorhinolaringol , 12(5):30-37 .
- Calhoun ,K.(1992). Diagnosis and management of sinusitis in the allergic patient. Otolaryngol Head Neck Surgery, 107(6):800-804.
- [Cardozo ,D.M.](#), [Nascimento-Carvalho C.M.](#), [Brandao M.A.](#), [Azevedo G.M.](#), [De Souza F.R.](#), [Silva N.M.](#), [Brandao A.P.](#), [De Andrade A.P.](#) and [Brandileone M.C.](#)(2006). Antimicrobial Resistance and Serotypes of Nasopharyngeal Strains of *Streptococcus pneumoniae* in Brazilian Adolescents. [Microb. Drug Resist.](#), 12(1):29-32.
- Chambers ,H.F.(2001). Chloramphenicol, Tetracycline, Macrolides, Clindamycin & Streptomycin. In Basic and Clinical Pharmacology. 8th ed. International edition McGraw-Hill. Pp 778-780.
- Chapman ,A.J., Musher D.M., Johnsson S., Clarridge J.E.and Wallace R.J.(1980). Development of bactericidal antibody during *Branhamella catarrhalis* infection. J. Infect. Dis., 101:S878-S882.
- Chart ,H.(1998). Toxogenic *E. coli*. Journal of Applied Microbiology, 84:77-86.

- Chart ,H. and Tenkin S.(1999).Serodiagnosis of infection with verotoxin-producing *E. coli* .Journal of Applied Microbiology, 86:731-741.
- Chilvers ,M., Hirst R., Baker N. and Andrew O.(2000). Novel mechanism of pneumonia: the differential effects of *Streptococcus pneumoniae* toxin on human respiratory cilia. Arch. Dis. Child., 82(44):162-166.
- Chow ,A.W., Hall C.B. and Klein J.(1992). Evaluation of new anti-infective drugs for the treat ment of respiratory tract infections. Clin. Infect. Dis., 15(1):62-88.
- Chow ,A.W.(1998). Infections of the sinuses and parameningeal structures. In Grobach S.L., Bartlett J.G. and Blacklow N.R. Infectious Diseases .2nd ed. W.B. Saunders. Pp 517-523.
- Collee ,J.G., Fraser A.G., Marmion B.P. and Simmons A.(1996). Test for identification of bacteria Mackie and McCartney practical medical microbiology . 1st ed., Churchill Livingstone ,USA.
- Conrad ,B., Dennis A., Jenson F. and Hal B.(2002). Management of acute bacterial rhinosinusitis. Curr. Opin. Pediatr., 14 (1) :86-91 .
- Courtiss ,E., Gargan T. and courtiss G.(1984). Nasal physiology. Ann. Plast. Surg., 13(22):214-216.
- Cowan ,A.S.T.(1980). Cowan and steel's manual for identification of medical bacteria .2nd ed. London , Cambridge University Press ,UK.
- Cruickshank ,R., Duguid J.P., Marmion B.P. and Swain R.H.(1990). Medical Microbiology. Vol.2, 1st ed. Churchill Livingstone , New York.
- Cullman ,W.(1997). *Moraxella catarrhalis* : virulence and resistance mechanisms. J. Med. Microbe., 92(3):162-166.
- Davidson ,T., Brahme F. and Gallagher M.(1989). Radiographic evaluation for nasal dysfunction: computed tomography versus plain films. Head Neck Surgery, 11:400-409.

- [Deng ,L.J.](#), [Luh](#) K.T., [Hsieh](#) W.C. and [Ho](#) S.W.(1981). Antibiotic susceptibility patterns of *Staphylococcus aureus* and *Staphylococcus epidermidis*. [Zhonghua Min Guo Wei Sheng Wu Ji Mian Yi Xue Za Zhi](#). 14(1):1-9.
- [Desrosiers ,M.](#), [Frenkiel](#) S., [Hamid](#) Q.A., [Low](#) D., [Small](#) P., [Carr](#) S., [Hawke](#) M., [Kirkpatrick](#) D., [Lavigne](#) F., [Mandell](#) H.E., [Stevens](#) K., [Weiss](#) I.J., [Witterick](#) E.D., [Wright](#) L. and [Davidson](#) R.(2002). Acute bacterial sinusitis in adults: management in the primary care setting. *J. Otolaryngol.*, 31 (2): 2-14.
- [Diaz ,I.](#) and [Bamberger](#) D.M.(1990). Acute sinusitis. *Semin. Respir. Infect.* 10(1):14-20.
- [Dishock ,H.](#) and [Franssen](#) M.(1907). The role of anaerobic bacteria in recurrent episodes of sinusitis and tonsillitis. *J. Med. Microbiol.*, 19(8): 002-003.
- [Douglas ,C.W.](#), [Heath](#) J., [Hampton](#) K.K. and [Perston](#) F.E.(1993). Identify of Viridans streptococci isolated from cases of infective endocarditis .*J. Med. Microbiol.*, 39:179-182.
- [Drake-Lee ,A.](#)(1997). The physiology of the nose and paranasal sinuses. In Alan G. Kerr and Michael Gleeson. *Scott-Brown's Otolaryngology*. 6th ed. Pp 171.
- [Durmaz ,B.](#), [Durmaz](#) R. and [Sahin](#) K.(1997). Methicillin-resistance among Turkish isolates of *Staphylococcus aureus* strains from nosocomial and community infections and their resistance patterns using various antimicrobial agents. [J. Hosp. Infect.](#), 37(4):320-9.
- [Dykewicz ,Ms.](#)(2003). Rhinitis and sinusitis. *J. Allergy. Clin. Immunol.*, 111(42):020-029.
- [Eisenstin ,B.I.](#) and [zaleznik](#) D.F.(2000). Enterobacteriaceae. In [Mandell](#) M.D., [Bennett](#) J.E. and [Dolin](#) R. *Principles and Practice of Infectious Diseases*. 0th ed. Churchill Livingstone. Pp 2294-2298.

- [Ergin ,A.](#), [Ercis S.](#) and [Hascelik G.](#)(۲۰۰۵). Macrolide resistance mechanisms and in vitro susceptibility patterns of viridans group streptococci isolated from blood cultures. [J. Antimicrob. Chemother.](#), ۵۷(۱):۱۳۹-۴۱.
- Evan, F.O., Syndor J.B., Moore F.C., Moore G.R., Manwaring J.L., Brill R.T., Jackson A.H., Hanna J.S., Skaar A., Holdman L.V, Fitz-Hugh G.S., Sande M.A. and Gwaltney J.M.(۱۹۷۵). Sinusitis of the maxillary antrum .N. Engl. J. Med., ۲۹۳(۲۳): ۷۳۵-۷۳۹.
- [Evans ,KL.](#)(۱۹۹۸).Recognition and management of sinusitis. [Drugs](#), ۵۶(۱):۵۹-۷۱.
- Facklam ,R.R. and Elliott T.A.(۱۹۹۵). Identification, classification and clinical relevance of catalase negative gram positive cocci excluding the *Streptococci* and *Enterococci*. [Clin. Microbiol. Rev.](#), ۸:۴۷۹-۴۹۵.
- Fagnan ,L.J.(۱۹۹۸). Acute sinusitis: Ascot- Effective Approach to Diagnosis and Treatment. [Am. Family Physician](#), ۵۸(۸):۱-۱۲.
- Farhadi ,M., Nejad G.B., Fathol-Lahzadeh B., Moazami N., Holakoei K. (۱۹۸۹). Bacterial etiology and antibiotic resistance in sinusitis: Astudy of ۲۶۴ cases. [M. J. I. R. L.](#), ۲(۲): ۱-۷.
- Farmer ,J.J.(۲۰۰۰). Introduction and identification of Enterobacteriaceae. [Amer. Society for Microbiol.](#), Pp :۲۱۲.
- Ferguson , B.J.(۱۹۹۵). Acute and chronic sinusitis. How to ease symptoms and locate the cause. [Postgrad. Med.](#) ۹۷(۵):۴۵-۴۸.
- Finegold ,S. M., Flynn M. J., Rose F. V., Jousimies- Somer C., Jakielaszek H., McTeague M., Wexler H. M., Berkowitz E. and Wynne B. (۲۰۰۲). Bacteriologic Findings Associated with Chronic Bacterial Maxillary Sinusitis in Adults. [Clinical Infectious Diseases](#), ۳۵:۴۲۸-۴۳۳.
- Frederich ,J. and Braude A.(۱۹۷۴). Anaerobic infection of the paranasal sinuses. [New England journal of medicine](#), ۲۳(۷۶):۲۹۰-۱۵۳.

- Geurkink ,N.(1983). Nasal anatomy, physiology, and function .J. Allergy. Clin. Immunol., 72(12):123-127.
- [Giglio ,M.S.](#), [Farias](#) O. and [Pinto](#) M.E.(1999). Surveillance of gram positive cocci susceptibility to betalactams, glycopeptides and other antimicrobials. [Rev. Med. Chil.](#), 127(8):919-920.
- Glezen ,W.P.(1998). The Common Cold. In Gorbach S.L., Bartlett J.G. and Blacklow N.R. Infectious Diseases. 2nd ed. W.B. Saunders Company. Pp 500-501.
- Gooch ,M.W.(1998). Potential infectious diseases complications of upper respiratory tract infections. Pediatric. Infect. Dis. J., 17(8):579-582.
- Gronroos ,J.A. and Palva A.(1962). The microbiology and management of acute and chronic rhinosinusitis. Acta. otolaryng.Stockh., 54(18):109.
- Gwaltney ,J.M., Sydnor A. and Sande M.A.(1981). Etiology and antimicrobial treatment of acute sinusitis. Ann. Otol. Rhinol. Laryngol., 90(84):78-71.
- Gwaltney ,J.M., cheld W.M., Sande M.A. and sydnor A.(1992). The microbial etiology and antimicrobial therapy of adults with acute community-acquired sinusitis: A fifteen year experience at the university of Virginia and review of other selected studies. J. Allergy. Clin. Immunol., 80(31):457-462.
- Gwaltney ,J.M.(1996). Acute-community-acquired sinusitis .Clin. Infect. Dis., 23(66):1209-1212.
- Gwaltney ,J.M.(1999). Acute community acquired bacterial sinusitis :to treat or not to treat. Can. Respire. J., 46(32):56-60.
- Hamilos ,D.L.(2000). Chronic sinusitis. J. Allergy. Clin. Immunol., 106(2):213-227.

- Hamory ,J.H., Sande M.A., Sydnor A., Seale D.L. and Gwaltney J.M. (١٩٧٩). Etiology and antimicrobial therapy of acute maxillary sinusitis. J. Infect. Dis., ١٣٩:١٩٧-٢٠٢.
- [Hamze ,M.](#), [Dabboussi F.](#), [Daher W.](#) and [Izard D.](#)(٢٠٠٣). Antibiotic resistance of *Staphylococcus aureus* at north Lebanon: place of the methicillin resistance and comparison of detection methods. [Pathol. Biol.](#), ٥١(١):٢١-٦.
- Hannele ,R., Seppo S. and Ylikoski S.(١٩٨٨). Bacteriological findings of acute maxillary sinusitis in young adults. J. Clin. Microbiol. ٢٦:١٩-٢٥.
- Hannele ,R., Jousimies S., Seppo C. and Ylikoski S.(١٩٨٩). Comparison of the nasal bacterial flora in two groups of healthy subject and in patients with acute sinusitis .J. Clin. Microbiology, ٢٧:٢٧٣٦-٢٧٤٣.
- Harabuchi ,Y. Faden H. and Yamanaka N.(١٩٩٤). Nasopharyngeal colonization with nontypeable *Haemophilus influenzae* and recurrent otitis media. J. Infect. Dis., ١٧٠:٨٦٢-٨٦٦.
- Harper ,J.J. and Tilse M.H. (١٩٩١). Biotypes of *Haemophilus influenzae* that are associated with non invasive infections .J. Clin. Microbiol., ٢٩:٢٥٣٩-٢٥٤٢.
- Hartog ,B., Degener J.E., Van Benthem P.P. and Hordijk G.(١٩٩٥). Microbiology of chronic maxillary sinusitis in adults: isolated aerobic and anaerobic bacteria and their susceptibility to twenty antibiotics. Acta. Otolaryngology, ١١٥(٥):٦٧٢-٦٧٧.
- Hashemi ,M.D., Sadeghi M.D., Omrani M.D. and Torabi M.D.(٢٠٠٥) Microbiology and antimicrobial resistance in chronic resistant rhino sinusitis with or without polyp after functional endoscopic sinus surgery. Journal of Research in Medical Sciences, ١٠(٣):١٦٧-١٧١.

- Heikens ,E.A., Fleer A.H., flor D.A. and Pavw N.B.(၂၀၀၀). Comparison of genotypic and phenotypic methods for identification of Coagulase-negative Staphylococcus. J. Clin. Microb., ၄၃:၂၂၈၆-၂၂၉၀.
- Hengerer ,A.(၁၉၈၄). Embryologic development of the sinuses. Ear ,Nose ,Throat journal, ၆၃(၄၂):၁၃၄-၁၄၄.
- Henry ,D. and Richard F.(၁၉၉၈). Gram negative bacilli. In Sherwood L., John G. and Niel R. Infectious Diseases. ၂nd ed. W.B. Saunders Company. Pp. ၁၂၆၉
- Herwald ,L.A.(၁၉၉၉). Control of methicillin resistant *staphylococcus aureus* in hospital setting. Am. J. Med., ၁၀၆(၁၁):၄၈-၀၂.
- Hsuch ,P.R., Huang W.K., Shyr J.M., Lau Y.J., Liu Y.C. and Luh K.T. (၂၀၀၄). Multicenter surveillance of antimicrobial resistance of *Streptococcus pyogenes* ,*Streptococcus pneumoniae*, *Haemophilus influenzae* and *Moraxella catarrhalis* to ၁၄ oral antibiotics. J. Formos. Med. Assoc. ၁၀၃ (၉):၆၆၄-၆၇၀.
- [Ikemoto, H.](#), [Watanabe K.](#), [Mori T.](#), [Igari J .](#), [Oguri T .](#), *et.al.*(၁၉၉၆). Susceptibilities of bacteria isolated from patients with respiratory infectious diseases to antibiotics (၁၉၉၄). [Jpn. J. Antibiot.](#), ၄၇(၀):၄၁၉-၄၀၀.
- [Inglis ,T.](#), [Kumarasinghe G.](#), [Chow C.](#) and [Liew H.Y.](#)(၁၉၉၄). Multiple antibiotic resistances in *Klebsiella* sop. and other Enterobacteriaceae isolated in Singapore. [Singapore Med J.](#) ၃၀(၆):၆၀၂-၆၀၄.
- Inoue ,M. ,Kaneko K., Akizawa K., Fujita S., Kaku M. , Yamaguchi J., Kohno S., Yamanaka K., Iinuma Y., Murase M., Yokoyama T., Asari S. and Hirakata Y.(၂၀၀၆). Antimicrobial susceptibility of respiratory tract pathogens in Japan during PROTEKT years ၁-၃ (၁၉၉၉-၂၀၀၂). J. Infect. Chemother., ၁၂(၁):၉-၂၁ .

- Jack ,M. and Gwaltney J.R.(۲۰۰۰). Sinusitis. In Gerald L., Mandell M.D., Jhon E., Bennett MD. and Dolin R. Principle and practice of Infectious Diseases. ۰th ed. Churchill Livingstone. Pp.۶۷۶.
- Jacops ,J.A., Schouten H.C., Stobberingh E.E. and Soeters P.B.(۱۹۹۰).Viridans streptococci isolated from the blood stream revelance of species identification. Diag. Microbiol. Infect. Dis., ۲۲:۲۶۷-۲۷۳.
- Jacopy ,G.A.(۱۹۹۳). Prevalence and resistance mechanisms of common bacterial respiratory pathogen. Clin. Infect. Dis., ۱۸:(۳۲):۹۰۱-۹۰۰.
- Janda ,W.M. and Knapp J.S.(۲۰۰۳). *Nisseria* and *Moraxella catarrhalis* . In Baron E.J., Jorgensen J.H., Pfaller M.A. and Tenover F.C. Manual of Clinical Microbiology .^۸th. Washington D.C. PP ۰۸۰.
- Karalus ,R. and Campagnari A.(۲۰۰۰). *Moraxella catarrhalis* :a review of an important human mucosal pathogen. Microes. Infect. ۲:۰۴۷-۰۰۹.
- Karlowsky ,J.A., Jonse M.E., Draghi D.C., Thomsberry C., Sahn D.F. and Volturo G.A.(۲۰۰۴). Prevalence and antimicrobial susceptibilities of bacteria isolated from blood cultures of hospitalized patients in the United States in ۲۰۰۲. Ann. Clin. Microbiol. Antimicrob., ۲(۱۲):۳-۷.
- Killian ,M.(۲۰۰۲). Streptococcus and enterococcus .In Greenwood D., Slack R.C.B., Peutherer J.F. Medical Microbiology . ۶th ed. Churchill Livingstone. Pp ۱۷۴.
- Kim ,P., Musher D.M. and Glezen W.P.(۱۹۹۰). Association of pneumococcal disease with season, atmospheric conditions, air pollution and the isolation of respiratory viruses. Clin. Infect. Dis., ۲۲:۱۰۰-۱۰۶.
- Kim ,C.S., Jeon S.Y., Min Y.G., Rhyoo C., Kim J.W., Yun J.B., Park S.W. and Kwon T.Y.(۲۰۰۰). Effects of beta-toxin of *Staphylococcus*

- aureus* on ciliary activity of nasal epithelial cells. Laryngoscope, 111(12):2080-2088.
- Kim ,J.Y., Park Y.J., Kim S.I., Kang M.W., Lee S.O. and Lee K.Y.(2004). Nosocomial outbreak by *Proteus mirabilis* producing extend-spectrum beta-lactamase VEB-1 in a Korean university Hospital. J.Antimicrob. Chemother., 54(6):1144-1147.
- [Kingdom ,T.T.](#) and [Swain R.J.](#)(2004). The microbiology and antimicrobial resistance patterns in chronic rhinosinusitis. [Am. J. Otolaryngol.](#) 25(5):323-8.
- Kinman ,J., Chang WL. and Seung HP.(1967). Acta. otolaryngo., 46:374.
- Kirby ,R. and Ruoff K.L.(1990). Cost effective clinically relevant method for rapid identification of beta hemolytic *Streptococci* and *Enterococci*. J.Clin. Microbiol., 33:1104-1107.
- Kloss ,W.E. and Bannerman T.L.(1994). Up date on clinical significance of coagulase negative *Staphylococci*. Cli. Microbiolo. Rev. 7(11):117-120.
- Kortekangas ,A.E.(1964). Antibiotic in the treatment of maxillary sinusitis .Acta. Otolaryngol. 188:379-389.
- Krishna ,V.(2004).The head and neck. Text Book of pathology. Orient Longman. Pp 622.
- Laine ,F.J. and smoker W.R.(1994). The ostiomeatal unit and endoscopic surgery:anatomy, variations, and imaging finding in inflammatory diseases. Am. J. Roentgenol ,159(4):849-857.
- Lanza ,D.C. and Kennedy D,W.(1992). Current concepts in the surgical management of chronic and recurrent acute sinusitis. J Allergy Clin Immunol., 9:500-511.
- Lazar ,R.(1993). Functional endoscopic sinus surgery in adults and children. Laryngoscope, 103(67):1-5.

- Lee , K., Lee HS., Jang S.J., Park A.J., Lee M.H., Song W.K. and Chong Y. (2001). Antimicrobials drugs . Med. Sci. 16:262-67.
- Lindbeak ,M., Melby K., Schoyen R. and Hjortdahl P.(2001). Bacteriological finding in nasopharynx specimens from patients with clinical diagnosis of acute sinusitis .Scan. J. Prim. Healthcare, 19(2):126-131.
- Little ,D., Mann B. and Sherk D.(1998). Factors influencing the clinical diagnosis of sinusitis. Journal of Family Practice, 46(2):147-152.
- [Liu ,Z.](#), [Gao Q.](#) and [Cui Y.](#)(1998).Bacteriological study of chronic maxillary sinusitis in adults and observation of susceptibility to antibiotics. Lin Chuang Er Bi Yan Hou Ke Za Zhi, 12(12):545-8.
- Liu Y. C., Huang W. K., Huang T. S.m, and Kunin C. M.(1999). Detection of antimicrobial activity in urine for epidemiological studies of antibiotic use. J. Clin. Epidemiol., 52:539-545.
- Liu ,F., Zhou S., Zhang S., Deng Q., Zhang L., Xu Y. and Wang J.(2001). Research of drug resistance and detection of methicillin-resistant *Staphylococcus aureus* in chronic maxillary sinusitis. Lin Chuang Er Bi Yan Hou Ke Za Zhi. 15(8):341-343.
- [Low ,D.E.](#), [Desrosiers M.](#), [McSherry J.](#), [Garber G.](#), [Remy H.](#), [Fenton R.S.](#), [Forte V.](#), [Balter M.](#), [Rotstein C.](#), [Craft C.](#), [Dubois J.](#), [Harding G.](#), [Schloss M.](#), [Miller M.](#), [McIvor R.A.](#) and [Davidson R.J.](#)(1997). A practical guide for the diagnosis and treatment of acute sinusitis. C.M.A.J., 15(6):S1-14.
- Lowy ,F.D.(1998). *Staphylococcus aureus* infections. N. Engl. Med. 339:521-522.
- Luh , K.T., Hsueh P.R., Teng L.G., Pan H.J., Chen Y.C., Lu J.J., Wu J.J. and Ho S.W.(2000) . Quinupristin- dalfopristin resistance among gram-positive bacteria in Taiwan .Antimicrob-Agents-Chemother., 44(12):3374-3381 .

- Lund ,V.J. and Loyd G.A.S.(1983). Radiological change associated with benign nasal polyps. J. of Laryngology and Otology, 97:503-510.
- Lundberg ,C., Carenfelt S., Engquest R. and Nord C.E.(1979). Anaerobic bacteria in maxillary sinusitis. Scand. J. Infect. Dis., 19:74-76.
- Lystad ,A.P., Berdal P. and Lund-Iversen L.(1964). The bacterial flora of sinusitis with an in vitro study of bacterial resistance to antibiotics. Acta. Otolaryngol., 188:390-400.
- Mackfaddin ,J.F.(2000). Biochemical test for identification of medical bacteria . 3rd ed. Williams and Wilkins- Baltimor. PP 321-400.
- Macky ,I.S. and Bull T.R.(1997). Infective rhinitis and sinusitis. Scott-Browne's Otolaryngology . 6th ed. Vol. 4. Butterworth-Hieneman. Pp 120.
- Manarey ,C.R., Anand V.K. and Huang C.(2004). Incidence of methicillin-resistant *Staphylococcus aureus* causing rinosinusitis. Laryngoscope , 114(5): 939-941.
- Mann ,W.K., Petz K., Schlenter W. and Niebling W.(1982). The role of *Streptococci* and *Haemophilus* in sinusitis .Laryngoscope, 33(4):17-21.
- Maresh ,M.(1989). Paranasal sinuses from birth to late adolescence. Am. J. dis. Child., 60(43):55-56.
- Melo-Cristino , J., Santo L., Ramirez M. and Respiratorias G.D. (2006).The Viriato Study : Update of antimicrobial susceptibility data of bacterial pathogens from community-acquired respiratory tract infections in Portugal in 2003 and 2004 . Rev. Port. Pneumol . 12(1):9-30.
- [Mendes , C.](#), [Marin](#) M.E., [Quinones](#) F., [Sifuentes-Osornio](#) J., [Siller](#) CC., *et.al.*(2004). Antibacterial resistance of community-acquired respiratory tract pathogens recovered from patients in Latin

- America: results from the PROTEKT surveillance study (1999-2000). [Braz. J. Infect. Dis.](#), 4(1):44-61.
- Miller. D.L. and Jones R.(1964). The bacterial flora of the upper respiratory tract and sputum of working men. *J. Pathol. Bacteriol.*, 87:182-186.
- Mims ,C., Dockneii H.M., Goering R.V., and Roitt I.(2004). *Medical microbiology* . 3rd ed , Elsevier Limited .
- Moller ,L.V.M., Regelink A.G. and Garsselier H.(1990). Multiple *Haemophilus influenzae* strains and strain variants coexist in the respiratory tract of patient with cystic fibrosis. *J. Infect. Dis.*, 172(02):1388-1392.
- Monora ,J.K., Monora R. and Paton J.C.(1997). Molecular and genetic characterization of the capsule biosynthesis locus of *Streptococcus pneumoniae* .*J. Bacteriol.*, 179(62):4903-4908.
- Monox ,E.R. and Murphy T.F.(2000). *Haemophilus influenzae*. In Mandell G. L., Bennett J.E., Dolin R. *Principles and practice of Infectious Diseases* . 6th ed. Churchill Livingstone. Pp 2369
- Morrissey ,I. , Robbins M., Viljoen L. and Brown DF.(2000). Antimicrobial susceptibility of community-acquired respiratory tract pathogens in the UK during 2002\3 determined locally and centrally by BSA methods. *J. Antimicrob Chemother.*, 00(2):200-208.
- [Moungthong ,G.](#) , [Suwas A.](#), [Jaruchida S.](#), [Chantaratchada S.](#), [Phonphok Y.](#) and [Rangsin R.](#)(2000). Prevalence of etiologic bacteria and beta-lactamase-producing bacteria in acute and chronic maxillary sinusitis at Phramongkutklao Hospital. *J. Med. Assoc. Thai.*, 88(4):478-483.
- Mounier-Kuhn ,P.(1903). Bacteriological study of maxillary sinusitis. *J.Fr. Otolaryng.*, 11(1):32-33.

- Murphy ,T.F.(1996). *Branhanella catarrhalis* :Epidemiology, surface antigenic structure and immune response .Microbial. Rev. J., 60(2):267-279.
- Murray ,B.E., Greenwood D. and Gilbert D.N.(1999). Manual of clinical microbiology. 9th ed. Amer. Society for Microbiology, PP:99.
- Namyslowski, W., Namyslowski G., Buszman E. and Misiolek M.(2004). Microbiology of acute exacerbation chronic sinusitis in adults. Otolaryngol. Pol., 58(2):331-337.
- NCCLS. (2000). Performance Standards for Antimicrobial Disk Susceptibility Tests, Approved Standard, 9th ed, document M7-A9. Villanova, PA: National Committee for Clinical Laboratory Standards.
- Nizet ,V., Colina K.F. and Almquist J.R.(1996). A virulent non encapsulated *Haemophilus influenzae* . J. Infect. Dis., 173(19):180-186.
- Norrby R.(1983).Clinical aspect on bacterial infection in the upper respiratory tract. Scand. J. infect.Dis., 39:14-18.
- O'Hara ,C.M., Brenner F.W. and Miller J.M.(2000). Classification, identification and clinical significance of *proteus*, *Providencia* and *Morganella*. Clinical microbiology reviews,13:534-546.
- Ologe ,F.E. and Nwabisi C.(2003). Bacteriology of chronic sinusitis in Ilron, Nigeria.African journal of Clinical and Experimental Microbiology, 4(2):91-97.
- Osguthorpe ,J. and Hadley J.(1999). Rhinosinusitis current concepts in evaluation and management . Med. Clin. North. Am., 83(32):27-33.
- Oteo ,J., Lazoro E., de Abajo F.J., Baquero F. and Campos J.(2000). Antimicrobial resistant invasive *Escherichia coli*. Emerg. Infect. Dis., 11:540-553.

- Pechere ,J.C.(٢٠٠١). Macrolide resistance mechanisms in Gram-positive cocci. Int. Antimicrob. Agents, ١(١٨):٢٥-٢٨.
- Penttila ,M., Savolainen S., Kiukaanniemi H., Forsblom B. and Jousimies-Somer H.(١٩٩٧). Bacterial findings in acute maxillary sinusitis European study. Acta Otolaryngol., ٥٢٩:١٦٥-١٦٨.
- Peter ,J.(٢٠٠٠). Treating acute sinusitis. Australian prescriber Journal. ٢٣(٢١):٣٩-٤٢.
- Petrov, M., Hadjiev N., Kantardjieva T., Velinov T.Z. and Bachvarova A. (٢٠٠٥). Surveillance of antimicrobial resistance in Bulgaria. Euro. surveillance, monthly release .١٠ .
- Pfaller ,MA., Ehrhardt A.F. and Jones A.F.(٢٠٠١). Frequency of pathogen occurrence and antimicrobial susceptibility among community-acquired respiratory tract infection in the respiratory surveillance program study : microbiology from the medical office practice environment . Am. J. Med., ٩:٤-١٢.
- Pichichero ,Mo.(١٩٩٥). Resistant respiratory pathogens and extended-spectrum antibiotic. Am. Fam. Physician. ٥٢(٦):١٧٣٩-١٧٤٦ .
- Podschum ,R. and Ullman U.(١٩٩٨). *Klebsiella* spp. as nosocomial pathogens ,epidemiology, taxonomy, typing methods and pathogenicity factors. Clinical microbiology review.١١:٥٨٩-٦٠٣.
- [Poole ,M.D.](#) (١٩٩٩).A focus on acute sinusitis in adults: changes in disease management. Am J Med. ١٠٦(٥):٣٨S-٤٧S.
- Prescott, L.M.,J.P. Havely and D.A. Klein .١٩٩٠. microbiology .wm. C. Bron Publisher ,USA.
- Pulimood , T.B., Lalitha M.K., Jesudason M.V., Pandian R., Selwyn J. and John T.J.(١٩٩٦). The spectrum of antimicrobial resistance among methicillin resistant *Staphylococcus aureus* (MRSA) in a tertiary care center in India. Indian J. Med. Res. ١٠٣:٢١٢-٢١٥.

[Quentin ,C.](#), [Arpin C.](#), [Dubois V.](#), [Andre C.](#), [Lagrange I.](#), [Fischer I.](#), [Brochet J.P.](#), [Grobost F.](#), [Jullin J.](#), [Dutilh B.](#), [Larribet G.](#) and [Noury P.](#)(٢٠٠٤). Antibiotic resistance rates and phenotypes among isolates of Enterobacteriaceae in French extra-hospital practice. [Eur J Clin Microbiol Infect Dis.](#) ٢٣(٣):١٨٥-١٩٣.

Radosz-Komoniewska ,H., Kapp-Burzynska Z., Klaptocz B., Wilk I. and Ekiel A.(١٩٩٧). Aerobic and anaerobic bacteria in chronic sinusitis in adults . Med. Dosw. Microbiol., ٤٩(١-٢):٨٩-٩٤.

Rajaduraipandi , K., Mani K.R., Panneerselvam K., Mani M., Bhaskar N. and Manikandan P.(٢٠٠٦). Prevalence and antimicrobial susceptibility pattern of methicillin resistant *Staphylococcus aureus* : amulticentre study . Indian J. Med. Microbiol., ٢٤(١):٣٤-٣٨ .

Rashmi ,S., Lal S. and Bhubneshwar K.(٢٠٠٥). Antibacterial resistance current problem and possible solution. J. Med. Sci., ٥٩:١٢٠-١٢٩

Robert ,D.H.(١٩٩١). Acute sinusitis: diagnosis and treatment update. J. American family physician, ٢٠(٣):١٧-٢٠.

Rohani ,M.Y., Raudzah A., Lau M.G., Zaidatul A.A., Salbiah M.N., Keah K.C., Noraini A. and Zainuldin T.(٢٠٠٠). Susceptibility pattern of *Staphylococcus aureus* isolated in Malaysian hospitals. [Int. J. Antimicrob. Agents.](#) ١٣(٣):٢٠٩-٢١٣.

Ron ,G., Shashy M.D., Eric J., Moore M.D. and Weaver M.S.(٢٠٠٤). Prevalence of the Chronic Sinusitis Diagnosis in Olmsted County, Minnesota. Arch. Otolaryngol. Head Neck Surg. ١٣٠:٣٢٠-٣٢٣.

Sarmad ,M.H.(٢٠٠٥). Some microorganisms isolated from maxillary sinusitis with some immunological aspects. A thesis submitted to the council of the college of Baghdad medicin.

Sener , B., Hascelik G., Onerci M. and Tunckanat F.(١٩٩٦). Evaluation of the microbiology of chronic sinusitis. J. Laryngol. Otol. ١١٠(٦):٥٤٧-٥٥٠.

- Shavidok ,I.G.(١٩٨٧). Sensitivity to beta-lactam and aminoglycoside antibiotics of clinical *Proteus* strains as dependent upon on their species classification and the source of their isolation. Antibiot. Med. Bioteknol., ٢٣(١١): ٨٥٠-٨٥٥.
- Shikani ,A.H.(١٩٩٧). Sinusitis : Ahead and Neck surgeon's perspechive. Medscape pulmonary medicines Journal, ١(٦):٦٦-٧١.
- Smith ,H.R. and Scotland S.M.(١٩٩٣). Isolation and identification methods for *E.coli* O١٥٧ and other verocytotoxic producing strain. J. Clin. Pathol., ٤٦:١٠-١٧.
- Smith ,A., Jackson M.S., Kennedy H.(٢٠٠٢). Antimicrobial susceptibility of Viridans group streptococci blood isolat to eight antimicrobial agents. Scand. J. Infect. Dis., ٣٦(٥٤):٢٥٩-٢٦٣.
- Snow ,V., Mottur-Pilson C. and Hickner J.M.(٢٠٠١). Principle of appropriate antibiotic use for acute sinusitis in adults. Ann. Intern. Med., ١٣٤:٤٩٥-٤٩٧.
- Spector ,S.L., Bernstien I.L. and Li J.J. (١٩٩٨). Parameter for the diagnosis and management of sinusitis. J. Allergy. Clin .Immunol., ١٠٢:S١٠٧-S١٤٤.
- Stalman ,W.A.B., Essen G.A. and Vander G.Y.(٢٠٠١). Determinats for the course of acute sinusitis in adult general practice patients. Postgrad Med. J. Des., ٧٧:٧٧٨-٧٨٢.
- Stamberger ,H.(١٩٨٦). Endoscopic endonasal surgery-concept and treatment of recurring rhinosinusitis. Part ١. Anatomic and pathophysiologic consideration. Otolaryngology Head Neck Surgery, ٩٤:١٤٣-١٤٧.
- Stankiewicz ,J., Newell D. and Parki A.(١٩٩٣). Complication of inflammatory diseases of the sinuses. Otolaryngol Clin. North. Am., ٢٦(٣٢):٦٣٩-٦٥٠.

- Stephenson ,W.P., Doern J., Gantz N., Lipworth L. and Chapin K.(1980). Pharyngeal carriage rates of *Haemophilus influenzae* type b and non-b, and prevalence of ampicillin-resistant *Haemophilus influenzae* among healthy day-care children in central Massachusetts. Am. J. Epidemiol., 122(00):868-870.
- Stock ,I.(2003). Natural antibiotic susceptibility of *Proteus spp.*, with special reference to *Proteus mirabilis* and *Proteus penneri* strains. J. Chemother., 10(1):12-26.
- Sutter ,V.L., D.M. Citron and Finegold S.M.(1980) .Wadsworth anaerobic bacteriology manual. 3rd ed. CV Mosby ,St. louis.
- Temple, M. and Nahata M.(2000). Pharmacotherapy of acute sinusitis in children. American Journal of Health system pharmacy, 57:663-668.
- Tinkelman ,DG. And Silk H.J.(1989). Clinical and bacteriological features of chronic sinusitis in children. Am. J. Dis. Child., 123(8):938-941.
- Tonkic ,M., Barisic I.G. and Punda-Polic V.(2000). Prevalence and antimicrobial resistance of extended-spectrum B-Lactamases producing *Escherichia coli* and *Klebsiella pneumoniae* strains isolated in university hospital in Split,Croatia. Int.Microbiol., 8:119-124.
- Torzillo ,P., Hanna J., Morey F.(1990). Invasive pneumococcal disease in central Australian. Med. j. Aust., 162:182-186.
- [Toye ,B.W.](#), [Scriver](#) S.R. and [Low](#) D.E.(1993). Canadian survey of antimicrobial resistance in *Klebsiella spp.* and *Enterobacter spp.* The Canadian Antimicrobial Resistance Study Group. [J. Antimicrob. Chemother.](#), 21:81-86.
- Turner ,B.W., Coil W.S., and Hendley J.O.(1992). Physiologic abnormalities in the paranasal sinuses during experimental Rhinovirus colds. J. Allergy. Clin. Immunol., 90(04):474-478.

- VanCauwenbergen ,P., kluyskens p. and Van Reterghem L.(1976).
Bacteriological findings in sinusitis (1963-1970). Scand. J.
infectious. Dis., 9:72-77.
- Vandipitte ,J., Engbeak K., Piot P. and Heuck C.C.(1991). Basic
Laboratory procedure in clinical bacteriology. 1st ed. WHO
Publication Geneva.
- Vanhoof ,R., Brouillard J., Damee S., Hondt N., Haccourt A., Mansoor I.,
Marchal J.F., Philippart I., Trigaux F., Van B., Bosterhaut C., Rossi
H. and Van Bossuyt E.(2000). High prevalence of penicillin
resistance and comparative in vitro activity of various antibiotic in
clinical isolation of *Streptococcus pneumoniae* isolated in the
Province of Hianaut during winter 2000. Acta. Clin. Belg.,
6.(6):340-349.
- [Varonen ,H.](#), Savolainen S., Kunnamo I., Hiekkinen R., and Revonta
M.(2003). Acute rhinosinusitis in primary care: a comparison, signs,
Ultrasound and radiography. Rhinology, 41(1):37-43.
- Wagenmann ,M. and Naclerio R.M.(1992). Anatomic and physiologic
consideration in sinusitis. J. Allergy. Clin. Immunol., 9.(22):419-
423.
- Wald ,E.R., Milmos G.J. and Bowen A.(1981). Acute maxillary sinusitis
in children. N. Engl. J. Med., 3.4(32):749-704.
- Wald ,E.R., Reilly J.S., Casselbrant M., Bluestone C.D. and Cliponic
D.(1984). Treatment of acute sinusitis in childhood: a comparative
study of amoxicillin and cefaclor. J. Pediatr., 1.4:297-302.
- Wald ,E.R.(1998). Microbiology of acute and chronic sinusitis in
children and adults. Am. J., 76(87):1-22.

- [Williams J.W.](#), [Simel DL.](#), [Roberts L.](#) and [Samsa G.P.](#)(1993). Clinical evaluation for sinusitis. Making the diagnosis by history and physical examination. [Ann. Intern. Med.](#), 119(1):92-97.
- Wu J., Tao Z., XU Y., Kong Y. and WU Y.(2000). The relationship between perennial allergic rhinitis and chronic sinusitis. *Lin.Chuang Er Bi Yan Hou Ke Za Zhi.*, 19(17):790-791.
- Yeh L.L. and Chi C.L.(2001). Another look at difference in susceptibility of *E. coli* and *Klebsiella pneumoniae* to cephalothin and cefazolin. *Int. Antimicrob. Agents*, 17(62):521-524.
- Zahnel G.G., Palatnick L., Nichol K.A., Low D.A. and Hoban D.J.(2002). Antimicrobial resistance in *Haemophilus influenzae* and *Moraxella catarrhalis* respiratory tract isolates: results of the Canadian Respiratory Organisms Susceptibility Study, 1997 to 2002. *Antimicrob. Agents. Chemother.*, 46(66):1870-1881.
- [Zhang Y.Y.](#), [Zhu D.M.](#), [Hu F.P.](#), [Wu S.](#) and [Wang F.](#)(2006). Changes of antimicrobial resistance among clinical isolates of *Escherichia coli* in Shanghai 1990-2004. [Zhonghua Yi Xue Za Zhi.](#), 86(1):12-16.
- Zinreich S.J.(1992). Imaging of chronic sinusitis in adults: X-ray, Computed Tomography and Magnetic Resonance Imaging. *J. Allergy. Clin. Immunol.*, 90(67):440-451.
- Zogaj X., Bokranz W., Nimitz M. and Romling U.(2003). Production of cellulose and curli fimbriae by members of the family Enterobacteriaceae isolated from the human gastrointestinal tract. *Infect. Immun.*, 71:4101-4108.

سيفوتاكسيم و سيفالوثين و سيبروفلوكساسين و اريثروميسين و تيتراسيكلين و فانكوميسين و ترميثوبريم) للعزلات البكتيرية أن المضادين الحيويين سبروفلوكساسين و سيفوتاكسيم كانا الأكثر تأثيراً ضد العزلات البكتيرية المختبرة.

الخلاصة

تم خلال هذا البحث جمع و دراسة ١٠٠ عينه (مسحات و غسيل الجيوب الانفية) من ١٠٠ مريض (٦٥ منهم إناث و ٣٥ ذكور) مصابين بالتهاب الجيوب الانفية (حسب التشخيص السريري الاولي من قبل اختصاصي الأنف والأذن والحنجرة) وممن أحيلوا إلى مستشفى الحلة التعليمي العام والعيادات ألتخصصية خلال فترة ١٠ اشهر (من تشرين الأول ٢٠٠٤ إلى تموز ٢٠٠٥) . تراوحت أعمار المرضى الذين شملتهم الدراسة من ١٠-٦٠ سنة كان من بينهم ٥٨ حالة مرضية حادة و ٤٢ حالة مزمنة لالتهاب الجيوب الانفية .

اظهرت النتائج أن أكثر فئة عمرية تعرضا لالتهاب الجيوب الانفية هي الفئة العمرية من ٢٠ إلى ٢٩ سنة وان النساء أكثر عرضه واستجابة للمرض مقارنة بالرجال.

اظهر ٥٠ مريضا (٨٦,٢%) ممن كانوا يعانون من الالتهاب الحاد نتائج موجبة اتجاه الزرع البكتيري مقابل نتائج سالبة ل ٨ مرضى (١٣,٨%) في حين اظهر ٣٥ مريضا (٨٣,٣%) من بين الحالات المزمنة نتائج موجبة للزرع البكتيري مقابل نتائج سالبة تمثلت ب ٧ مرضى (١٦,٧%) .

الأنواع البكتيرية التي تم عزلها وتشخيصها خلال هذه الدراسة من الحالات المرضية الحادة شملت أنواعا بكتيرية موجبة وسالبة لصبغة غرام جاءت في مقدمتها بكتريا *Streptococcus pneumoniae* حيث شكلت نسبة مئوية مقدارها ٢٥%، جاءت بعدها بكتريا *Moraxella catarrhalis* (١٨%) ، ثم *Staphylococcus aureus* (١٦,٧%) ، ثم Coagulase- negative Staphylococci (١٤%) ، ثم Viridians streptococci (١٢%) ، ثم *Klebsiella pneumoniae* (٨%) ، وأخيرا *Streptococcus pyogenes* (٦%) . أما في الحالات المرضية المزمنة فقد جاءت في مقدمتها بكتريا *Staphylococcus aureus* (٤٢,٨%) تلتها *Streptococcus pneumoniae* (٢٣,٤%) ، ثم *Moraxella catarrhalis* (١٠,٧%) ، ثم *Klebsiella pneumoniae* (٩%) ، ثم *E. coli* (٥,٣%) ، ثم Viridians streptococci (٥,٣%) ، وأخيرا *Proteus mirabilis* (٣,٥%) .

كان النوع المقاوم للمضاد الحيوي الميثيسيلين من بين عزلات العنقوديات الذهبية Methicillin Resistance *Staphylococcus aureus* (MRSA) في كلا الحالتين الحادة والمزمنة ، فقد مثل نسبة مئوية مقدارها ١٤% و ٣٧% على التوالي. اظهرت نتائج اختبار فحص الحساسية للمضادات (امبيسيلين و اموكسيسيلين و جنتاميسين و



جامعة بابل
كلية الطب

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رسالة

مقدمة الى مجلس كلية الطب/جامعة بابل
كجزء من متطلبات نيل درجة الماجستير
في علم الأحياء المجهرية الطبية

من قبل

تيسير صفاء

٢٠٠٧ كانون الثاني

ذو الحجة ١٤٢٧