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بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

﴿ وَعَايَاتِكُمْ مِّنْ كُلِّ مَّآ سَأَلْتُمُوهُ وَإِن تَعُدُّوا نِعْمَتَ
اللَّهِ لَا تُحْصُوهَا إِنَّ الْإِنْسَانَ لَظَلُومٌ كَفَّارٌ ﴾.

صدق الله العلي العظيم

سورة إبراهيم

آية (٣٤)

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College of Medicine

BACTERIOLOGICAL STUDY REGARDING

ACUTE APPENDICITIS

**A THESIS SUBMITTED TO THE COUNCIL OF THE
COLLEGE OF MEDICINE, UNIVERSITY OF
,BABYLON**

**IN PARTIAL FULFILLMENT OF THE
REQUIREMENTS**

**FOR THE MASTER DEGREE OF SCIENCE IN
MEDICAL MICROBIOLOGY**

SUBMITTED BY

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A.H ١٤٢٧A.D

٢٠٠٦JUN

الخلاصة

تضمنت هذه الدراسة ١١٠ مريضاً كانوا مصابين بالتهاب الزائدة الدودية الحاد وزاروا مستشفى- الفرات الاوسط - وحدة الجراحة البولية وكانت اعمارهم تتراوح من ١-٦٠ سنة خلال فترة ١٠ اشهر(من شهر تشرين الاول ٢٠٠٤- الى تموز ٢٠٠٥) وكان ٦٩ (٦٢.٧%) منهم ذكور و ٤١ (٣٧.٣%) منهم أنثى وأظهرت النتائج إن الفئة العمرية الأكثر عرضة للإصابة بين (١١-٢٠) سنة من كلا الجنسين منهم ٣٥ (٣١ و٨%) ذكوراً و ٢١ (١٩%) أنثى. فمن الناحية السريرية كانوا يعانون الأم الجهة اليمنى مع الم بصورة عامة وحمى وفقدان الشهية وغثيان وإمساك وأحياناً حالات إسهال وصداع وصعوبة التبول. أما التشخيص المختبري فقد تضمن حساب عدد كريات الدم البيضاء و C. Reactive protein وفحص الادارا العام بالإضافة الى فحوصات اخرى مثل الفحص بجهاز السونار. وأظهرت النتائج بأن هناك ١١١ عزلة تم تشخيصها من ١١٠ مريضاً، وكانت البكتريا الممرضة ٨٧ عزلة (٧٨.٣%) هوائية و ٢٤ (٢١.٧%) عزلة لا هوائية، وقد تم الحصول عليها من ١١٠ مسحة، اما بالنسبة لنتائج زرع المسحات كانت موجبة ٩٠ (٨١.٨%) مريض فقط وفي ٢٠ (١٨.٢%) مريضاً كانت سالبة، وكانت انواع البكتريا الظاهرة في الزرع ونسبتها كما يلي ٣٦ (٣٢.٦%) *E. coli* و ٢١ (١٨.٩%) *Bacteroides spp* و ١٨ (١١.٢%) *Klebsiella pneumoniae* و ١١ (٩.٩%) *Pseudomonas aeruginosa* و ٧ (٦.٣%) *Citrobacter freundii* و ٥ (٤.٥%) كل من *Salmonella typhi* و *Proteus mirabilis* و ٤ (٣.٦%) *Enterobacter aerogenes* و ٢ (١.٨%) *Peptostreptococcus spp* و ١ (٠.٩%) كل من *Staphylococcus aureus* و *Clostridium perfringens*. وقد اظهرت الدراسة وجود حالات مزدوجة(ثنائي الاصابة) ٢١ عزلة حيث كانت فيه بكتريا *E.coli* مع بكتريا لاهوائية هي الاكثر سيادة. وقد اظهرت النتائج زيادة في حالات الاصابة بالتهاب الزائدة الدودية خلال اشهر الصيف وقلت خلال أشهر الشتاء. حيث سجلت اعلى إصابة

٢٦ (٢٣.٦%) خلال الشهر السابع و١٨ (١٦.٢%) أصابه خلال الشهر السادس. واقل نسبة ٤ (٣.٦%) خلال شهر كانون الثاني. اما بالنسبة لاختبارات الحساسية تم استخدام طريقتين وهما طريقة الانتشار بالاقراص وطريقة الثخايف الدقيقة لقياس التراكيز المثبط الادنى. اما بالنسبة لتأثير بعض المضادات الحيوية على تلك العزلات البكتيرية. فقد أظهرت النتائج ان مقاومة العزلات للاموكساسولين (٨٤.٦%) وللكرومايسسن (٦٢%) ولللاميكاسين (٥٧.٦%) وللسيروفلوكساسولين (٣٨.٧%) وللسينوتكسام (٤١.٤%) ولل سيفوكسيم (٤٧.٧%) وللمثب ريم (٨٢.٨%) ولل فانكوميسين (٩٦.٣١%) ولل نورفلوكساسولين (٧٦.٥%) وللتتراسايكلين (٧٦.٥%).

قرار لجنة المناقشة

نحن أعضاء لجنة المناقشة نشهد بأننا اطلعنا على الأطروحة الموسومة من الطالب فائز كامل كتاب السلامي وقد ناقشنا الطالب في محتوياتها، وفيما له علاقة بها، ووجدنا بأنها مستوفيه بالقبول بدرجة (أمتياز) لنيل درجة الماجستير في علوم الحياة / الأحياء المجهرية.

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Summary

The subjects of this study were 110 patients having acute appendicitis who were referred to Kufa Teaching Hospital, Department of urology whose ages ranged from 1 to 70 years old. Those subjects were investigated through a period of 10 months from October, 2004 to July, 2005. Patients were 69 (62.7%) males and 41 (37.3%) females. The age group of both sexes being more susceptible for appendicitis ranged from (11-20) years old. The clinical features of patients being observed by physicians were studied. Those were right iliac fossa pain, generalized abdominal pain, fever, loss of appetite, nausea, constipation, diarrhea, headache and dysuria. White blood cells count, C. reactive protein and general urine analysis were also studied, in addition to abdominal ultrasonography and computer tomography (CT). A total of 111 bacterial isolates were identified from inflamed appendix of 110 patients with acute appendicitis. Positive bacterial cultures were detected in 90 (81.8%) patients while 20 (18.2%) patient showed no growth. The aerobic bacteria accounted for 87 (78.3%) isolates whereas anaerobic were 24 (21.6%) isolates. Gram negative bacteria were presented in 107 (96.1%) while Gram positive bacteria were accounted 4 (3.6%). *Escherichia coli* were the predominant pathogens, since it accounted for 36 (32.4%) of all isolates followed

by *Bacteroides* spp 21(18.9%), *Klebsiella pneumoniae* 18(16.2%), *Pseudomonas aeruginosa* 11(9.9%), *Citrobacter freundii* 4(6.3%), *Salmonella typhi* 0(0.0%), *proteus mirabilis* 0(0.0%), *Enterobacter aerogenesa* 4(3.6%), *peptostreptococcus* 2(1.8%), *Staphylococcus aureus* 1(0.9%) and *Clostridium perfringns* 1(0.9%). Mixed cultures were detected in 21 cases, in which more than one organism were detected. Most of mixed bacterial isolates were anaerobic with aerobes (13, 61.9%), in which *Escherichia coli* was the common (10, 76.9%).

A remarkable an increased occurrence of acute appendicitis was observed at warm season, while the incidences were decreased during the winter season. The peak of appendicitis occurrence was observed during July 26(23.6%) and June 18(16.4%), whereas the lowest frequency of appendicitis was observed in January 4(3.6%).

In this study two methods disk diffusion method and minimum inhibitory concentration were used to determine the susceptibility of bacterial isolates to 13 antibiotics. The results showed that (84.6%) of total isolates were resistant to (96.3%), to vancomycin, (90.4%) to, (84%) to, tetracycline (83.8%) to, cefixime, (76.0%), to norfloxacin, (62%) to, amoxycillin, (58.0%) to Garamycin, (46.8%), to trimethoprim (42.3%), to cefotaxime, to amikacin, (38.7%), to ciprofloxacin.

CERTIFICATION

I certify that this thesis was prepared under my supervision at the College of Medicine, University of Babylon, as a partial fulfillment of the requirements for the degree Master of Science in **Medical Microbiology**.

Advisor

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Date: / / ٢٠٠٦

In view of the available recommendation, I forward this thesis for debate by the examining committee

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Examination committee certificate

We, the examining committee, certify that we have read this thesis "Bacteriological study on acute appendicitis" and have examined the student Faiz Kamil Al-Salami in its contents, and that in our opinion it is adequate as a thesis for the degree of master in biology (microbiology).

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HOLY IMAM

AL- HUUJA

TO MY DEAR WIFE

TO MY DEAREST DAUGHTERS

SARA, ZAHRA AND FATMA

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Abbreviations

ART	Antimicrobial resistance testing
BHI	Brain heart infusion
C	Centigrade
CFA	Colonization factor antigen
CFU	Colony forming unit
CM	Cubic millimeter
CT	Computerized tomographic study
EMB	Eosin methylene blue
G	Gram
Hrs	Hours
KIA	Kliglar iron agar
LF	Lactose fermentation
μg	Micro gram
μl	Micro liter
MIC	Minimum Inhibition concentration

ml	Mili liter
mm	Mili meter
NCCLS	National Commetti of Clinical Laboratory Standards
No	Number
Spp	Species
Tmp. Smx	Tri methoprim- sulfamethoxazole
TSI	Triple sugar iron
VP	Vogas proskaur
WBCS	White blood cells
WHO	Word Health Organization

1-1 INTRODUCTION:

Appendix is an organ connected to the digestive system and usually referred to as functionless organ in human. It is now well recognized that this organ may play a role in immunity (Zuercher *et al.*, 2002). The position of this organ lies in the right lower part of the abdomen. In spite of the minor significance of this organ, it is susceptible for infections as other organs such as hepatitis, pneumonitis, cholecystitis, tonsillitis and encephalitis (Schwartz *et al.*, 1996). Appendicitis has been reported world wide. In USA, it was estimated as 200,000 case per / year (Addiss *et al.*, 1990). In Britain

the infection rate was estimated at an average of ۷۰,۰۰۰ case per/ year (Rao *et al.*, ۱۹۹۸). According the (WHO, ۲۰۰۴). The mortality rates, all around the world are summarized in table (۱-۱)

There are no accuratel records about appendicitis in Iraq. The general Cases being registered in Najaf General Hospital were ۵۳۰ case per/ year; while in Kufa hospital were ۴۵۰ case per/ year.

Appendix can be infected by a variety of microbes (parasite, viral and bacterial).A surgical operation should be done to remove this organ and to protect the body and maintain its health from complications.

Table (۱-۱) Country and mortality rate according to WHO, (۲۰۰۴)

Country	Mortality rate per ۱ millon people
Banamas	۶.۶۲ deaths
Cuba	۵.۱ deaths
Malta	۵ deaths
Denmark	۴.۹ deaths
Ecuador	۳.۸ deaths
Nicargua	۳.۴ deaths
Costarica	۲.۹ deaths
Mexico	۲.۵ deaths
Brazil	۲.۴ deaths
Finland	۲.۲ deaths
Germany	۱.۷ deaths
Argentina	۱.۳ deaths

South Africa	١.٢ deaths
Spain	١.١ deaths
Australia	٠.٦٩ deaths
Korea	٠.٦٤ deaths
Japan	٠.٥٣ deaths
Kuwait	٠.٤٢ deaths
Egypt	٠.١٤ deaths
United Kingdom	٠.٠٣ deaths

The great danger following the appendicitis is the rupture of the appendix, leading to release the causative pathogens in to abdominal cavity resulting serious complication (Damjarov *et al.*, ١٩٩٨). Such complications could be avoided by abdominal surgical operation particularly in developing countries. It is worthily mentioned that this human organ has been pointous as a concern of bacteriologists. However most studies focused on the diagnosis of the appendicitis. Other studies deal with parasitic infection as it is a reason of appendicitis, but it seems the rare studies have been done regarding the bacteriological infection. Therefore this study was suggested to

full fill the following aims:

١-Isolation and identification of bacterial profiles associated with
 appendicitis

٢-Comparison of bacterial isolates to determine the common

causes of appendicitis.

۳-Detection the susceptibility of isolated bacteria to antibiotics.

۱-۲.LITERATURES REVIEW

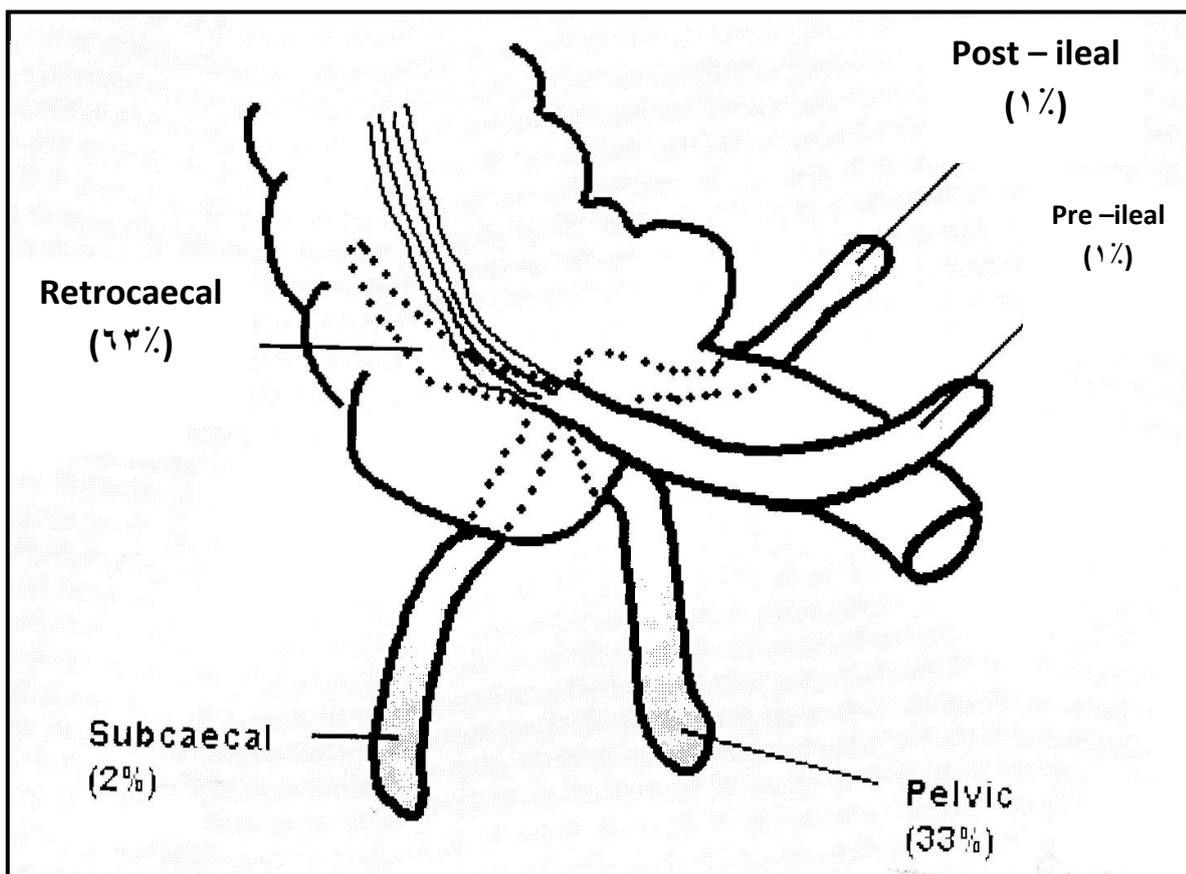
۱.۲.۱. Appendix

۱.۲.۱.۱. Anatomy

The appendix first becomes visible during embryologic development in the eighth week of fetal life as a protuberance off the terminal portion of the cecum. During both antenatal and postnatal development, the growth rate of the cecum exceeds that of the appendix. Displacing the appendix medially toward the ileocecal valve. The relationship of the base of the appendix to the cecum remains constant, whereas the tip can be found in a retrocecal, pelvic, subcecal, preileal, or right pericolic position. These

anatomic considerations have significant clinical importance in the context of acute appendicitis. (Storer *et al.*, 1988; Schwartz *et al.*, 1996).

The appendixes vary in length from less than 1 cm to greater than 30 cm. Most appendixes are 8 to 10 cm in length. Finger like structure or narrow tubular pouch arising from caecum first part of the large intestine. The appendix is normally found in the right lower part of the abdomen, removed without detrimental effect. The appendix can be found anywhere in the abdomen. It may be found retrocaecal in a percentage of 63%, pelvic 33%, subcaecal 2%, postileal and preileal 1% (figure 1-1) (Doherty and Lewis, 1989; Shelton *et al.*, 2003).



(۲٪)

Pelvic
(۳۳٪)

(Fig. ۱-۱) Diagram illustrates the location of appendix (Shelton *et al.*, ۲۰۰۳).

۱.۲.۱.۲. Appendix Function

For many years the appendix was erroneously viewed as a vestigial organ with no known function. It is now well recognized that the appendix is an immunologic organ which actively participates in the secretion of immunoglobulins, particularly IgA (Zuercher *et al.*, ۲۰۰۲). Though the appendix is an integral component of the gut associated lymphoid tissue (GALT) system (Craig *et al.*, ۱۹۹۴) lymphoid tissue first appears in appendix about ۲ weeks after birth. The amount of lymphoid tissue increases throughout puberty. Remains steady for the next decade and there begins a steady decrease with age. After the age ۶۰, virtually no lymphoid tissue

remain with in the appendix and complete obliteration of the appendiceal lumen is common (Schwartz *et al.*, 1996; Wagner *et al.*; 1996).

The appendix attachment to the intestine when the appendix is blocked it becomes inflamed and results in the condition know as appendicitis. If the blockage continues the inflamed tissue become infected with bacteria and begins to necrotize from alack of blood supply which finally result in appendix perforation (Malone and Shetty, 1997; Williams and MarGs., 2003).

The appendixe was regarded as inflamed organ if it has one or more of the following features (Adams *et al.*, 1988; Davis *et al.*, 1990).

- 1- Peritoneum: congestion of the serosal vessels and inflammatory exudates commonly fibrinous or frankly purulent.
- 2- Wall: oedema, congestion and swelling of the muscular wall.
- 3- Lumen: contain an obstructing faecalith and is distended by purulent material.
- 4- Mucosa: hyperaemic swallow and after ulcerated or completely necrotic.

1.2.2. Historical Notes on Appendicitis :

Until the end of the 19th century appendicitis remained unrecognized, before this time it was comparatively rare disease .but these can be no doubt that it extended even in remote time .for an

acutely inflamed, perforated appendix was found preserved in the mummy of a young royal of Egypt (Rains and Capper, 1911). According to McMurrich in 1930, Leonardo da Vinci in Italy was the first who described and illustrated the appendix in 1482. In 1848 Henry Hancock presented a paper entitled the infection of the appendix, the condition which is cured by operation. He was the first surgeon to treat peritonitis due to perforated appendix successfully. In 1898 the infection aetiology of appendicitis was first proposed by Veillon and Zuber. They emphasized the presence of anaerobes as the cause of peritonitis and abscess in appendicitis. Gram negative anaerobic bacilli were the dominant pathogen in 21 cases out of 22 suppurative appendicitis (Veillon & Zuber, 1898). Dudgeon & Sargent, (1900) stated that only one isolate of *Clostridium perfringens* was detected among 27 cases. *E-coli*, *Streptococci* and *Staphylococcus*. Were also isolated as gram negative anaerobic bacilli from 96 cases of peritonitis associated with acute perforated appendicitis. In (1901) Metchuickoff reported three cases of appendicitis due to the presence of *Ascaris lumbricoides* – studies on the incidence of appendical enterobiasis and the possible causal relationship of organism to appendicitis have appeared with increasing frequency since 1900 (Altemeier, 1938; Boulose and Cowie, 1973).

1.2.3. Appendicitis

1.2.3.1 Acute Appendicitis

Acute appendicitis is one of the most common causes of abdominal pain often requiring surgical intervention to avoid any suspected complications (Elhag *et al.*, 1986). Approximately 7% of the population suffer from acute appendicitis during their life time (Wagner *et al.*, 1996). The etiological factors for acute appendicitis may widely vary (Elhag *et al.*, 1986; Al-Joubori, 1994). The most important factor is the obstruction of lumen, the commonest cause of obstruction was reported to be a fecolith (Majed and Al Bakri, 1984). But could be a foreign body (Anderson and Bergdahi, 1978; Green *et al.*, 1994). Appendicitis occurs when the diet is reduced in bulk with diminished cellulose and high protein is taken (Burkitt, 1971; Gibbons and Houte, 1970).

1.2.3.2. Chronic Appendicitis

Although the existence of chronic appendicitis is controversial, most surgeons have managed patients with recurrent abdominal pain that resolved with appendectomy. In some cases the history consists of intermittent right lower quadrant abdominal pain and in fact the appendical pathology demonstrates chronic changes consistent with recurrent inflammation and fibrosis. Some surgeons have suggested that identification of a non filling or partially filling appendix on contrast enema or computer tomographic as well as the failure of contrast to drain from the appendix after few days following contrast administration, is suggestive of chronic or

recurrent appendicitis in the patient with chronic abdominal pain. Although one should perform a thorough evaluation of the patient with chronic abdominal pain prior to operative intervention (Drusano *et al*; Weiner *et al.*, 1990; Schales *et al.*, 1997).

1.2.4 Incidence

1.2.4.1. Age Incidence

Appendicitis is rarely encountered before the age of two years. It is unusual encountered the age of one year and becomes increasingly common during childhood and adolescence. The maximum incidence is between the 20 to 30 years, there after there is a gradual decline but no age is exempt (Dohert *et al.*, 1989; Rain and Capper, 1988).

1.2.4.2 Sex Incidence

Sex ratio in acute appendicitis is about 1:1 prior to puberty the frequency in males increases so that the male/female ratio is about 2:1 between the age of 10 to 20 years after which the male incidence gradually declines until the sex related incidence are again equal (Storer *et al.*, 1988; Benko *et al.*, 1996).The incidence of

appendicitis is approximately 1.5 times greater in men than in women while the incidence of primary appendectomy is approximately equal in both sexes (Doherty and Lewis, 1989; Kosloske, 2004).

Primatesta and Coldacre (1994) studied the demographic and temporal profiles of emergency appendectomy for age and sex specific admission rates were studied subdividing appendectomy into three main categories.

1. Emergency appendectomy for acute appendicitis was more common in males than in females peaked in the 10-19 year age group and declined overtime.

2. Emergency appendectomy as the main operation without appendicitis recorded a diagnosis was more common in women than men (Female male ratio 1.9:1) peaked at age 10-19 years and did not decline overtime.

3. Incidental appendectomy with other operations but without appendicitis was much common in women (female male ratio 3:1). Peaked at older ages than the first two groups and declined significantly overtime.

1.2.5. Etiology of appendicitis

Although inflammatory disease of the appendix has been recognized as such for more than 100 years its aetiology remains a subject of controversy since early in this century consumption of

low fiber diet has been implicated .More recently, this theory has been challenged and the role of infection emphasized (Barker, 1980; Larner, 1988).

1.2.5.1. **Obstruction**

The presumed etiology is obstruction most commonly fecalith with parasite seen in other parts of the world. Obstruction of the lumen is the dominant causal factor in acute appendicitis .Fecaliths are the usual cause of appendiceal obstruction less common are hypertrophy of lymphoid tissue in spissated barium from previous x-ray studies. Vegetable and fruit seeds and intestinal worm particularly *Ascaris* also play role in obstruction (Butter, 1981; Lee *et al*, 1980; Blandino *et al.*, 1990).

Frequency of obstruction rises with the severity of the inflammatory process fecaliths were found in about 40% of cases of simple acute appendicitis and about 60% of cases of gangrenous appendicitis with out rupture while about 9% of cases of gangrenous appendicitis with rupture (Rains & Capper, 1988). Schmidt & Robert (1980) reported that through appendicitis resulted due to mucosal injury of appendix or destruction of tissue by mechanical or chemical means.

1.2.5.2- **Bacterial Appendicitis.**

Bacteria seem to play a major role in appendicitis. It occurs when bacteria in vase and grow in the blocked appendix causing it to become swollen and infected, If the infection progress the swelling can cut off the blood supply to the appendix which result in killing the tissue, the wall of the appendix may rupture and infection spills into abdominal cavity causing peritonitis (Shelton *et al.*, 2003). A variety of bacterial species have been reported to play a major role in the appendicitis .Aerobic and anaerobic bacteria were detected in appendicitis cases and the commonest anaerobe was *Bacteroid fragilis* (Elhag *et al.*, 1986; Gilbert *et al.*, 2002). The significance of beta haemolytic streptococci in appendicitis has been studied by Jakobsen *et al.*(1987) who pointed out that only six cases were identified as streptococcal causes .Okoro (1988) stated that *Yersinia enterocolitica* accounted for 13.8% of the microorganisms impliated in appendicitis . A retrospective study on the microbiology of abdominal pus from acute appendicitis or peritonitis was carried out by Guasco *et al.*, (1991) they found that 40 (84.4%) specimens were positive for bacteriological examination of which (76.3%) were polymicrobial the most represented species were *E.coli* (28.4%), *Bacteroid fragilis* (18%), *Streptococcus milleri* (18%) and *Ps. aeruginosa* (3.9%) The polymicrobial mostly represented by *Bacteroidaceae spp*, *Enterobacteriaceae spp* and *Streptococens spp*. A study on inflamed appendices revealed mixed infections and the most common organisms were *E.coli*, Enterococci, anaerobic

streptococci together with *Clostridium prfringenes* and *Bacteroides* (Rains & Capper 1988). Felner & Dowell (1971) reported that *Bacteroides* species accounted for 10% of the appendiceal lesion while Leigh *et al.*, (1974) isolated *Bacteroides spp* from appendectomy cases in a percentage of 48% where as other Gram-negative bacilli represented by *E. coli* and *Klebsiella* were present in a percentage of 29% and 27% respectively. Gram positive cocci were less frequently.

A bacteriological study for 110 emergency appendectomies in children was reported by Barker, (1980). They suggested that *Bacteroides* considered as an important pathogen in appendicitis. However anaerobic bacteria seemed to be increasingly colonized the appendix and the ileum in cases of appendicitis (Bennion *et al.*, 1990; Park homenlco *et al.*, 1991) studied the characteristic of viral-bacterial lesion in the appendix of children with appendicitis. They obtained antigen of influenza viruses A, B, C, Entero, Adeno, and paramyxo viruses among which influenza c was significantly more frequent (64.1%); viral infection was the majority of cases (79.0%). It was found in association with opportunistic flora more frequently with *E. coli* and less *Ps. aeruginosa* and *Klebsiella*.

Baron *et al.*, (1992) reported that bacteria recovered from appendiceal specimens of patients with acute appendicitis were smaller in number and fewer in species in culture compared with

specimens from patients with more complicated disease. Some appendicitis was found to be caused by *Salmonella spp.* (Kazlow *et al.*, 1991). Moreover Parkhomenko(1998) isolated *Citrobacter frundii* and *Yersinia enterocolitica* from some appendicitis cases.

1-2-5-3 Viral Appendicitis:

Appendicitis may also subject for infection by a group of viruses such adeno viruses, cytomegalo viruses (Lamps, 2004). In such cases appendix becomes accessible by bacteria which invade the appendix rapidly to become inflamed and filled with pus (Lamps, 2004).

1-2-5-4 Parasitic Appendicitis

Parasites have also been reported to be implicated in appendix infection .*Trichuris trichiura*. *Ascaris lumbricoides* .*Entamoeba histolytica*, *Taenia saginata*, *Schistosoma haematopium* and *Strongyloides stercoralis* were isolated and identified from infected appendix (Calchi *et al.*, 1996; Rivero *et al.*, 1996), but the common parasite was *Enterobius vermicularis* (Chang and Fuh, 1987; Dohistrom and Macrthw, 1994).

1-2-6 Risk Factors for Acute Appendicitis

According to the available knowledge, there is no direct factor causing appendicitis. Even though factors seem to be associated in some way with appendicitis, or make the chance of infection higher

but these are not always lead to cause the infection. (Shsrn and Gupta, २००६). Though the following conditions have been recommended to participate with acute appendicitis in various sources include low fiber diet, refined carbohydrate, amebiasis, bacterial gastroenterities and viral gastroenteritis.

१-२-६-१. Diet

A confirmed evidence being concluded from geographical and economical studies revealed that the change from a high to low residue diet is largely associated with appendicitis (Burkitt, १९५१), short (१९२०) showed that if the individuals from African areas migrated to urban areas where appendicitis is common they soon become susceptible to the infection.

This may due to departure from simple diet which rich in cellulose to one relatively rich in meat, but this cannot be the whole explanations, since acute appendicitis may occur in life long vegetarians and even in babies of the breast feeding (Rains & capper, १९८८).

The reduction of dietary cellulose not only slows fecal transit but also results in firmer and more tenacious stools which are considerably reduced in bulk and predispose to fecalith formation.

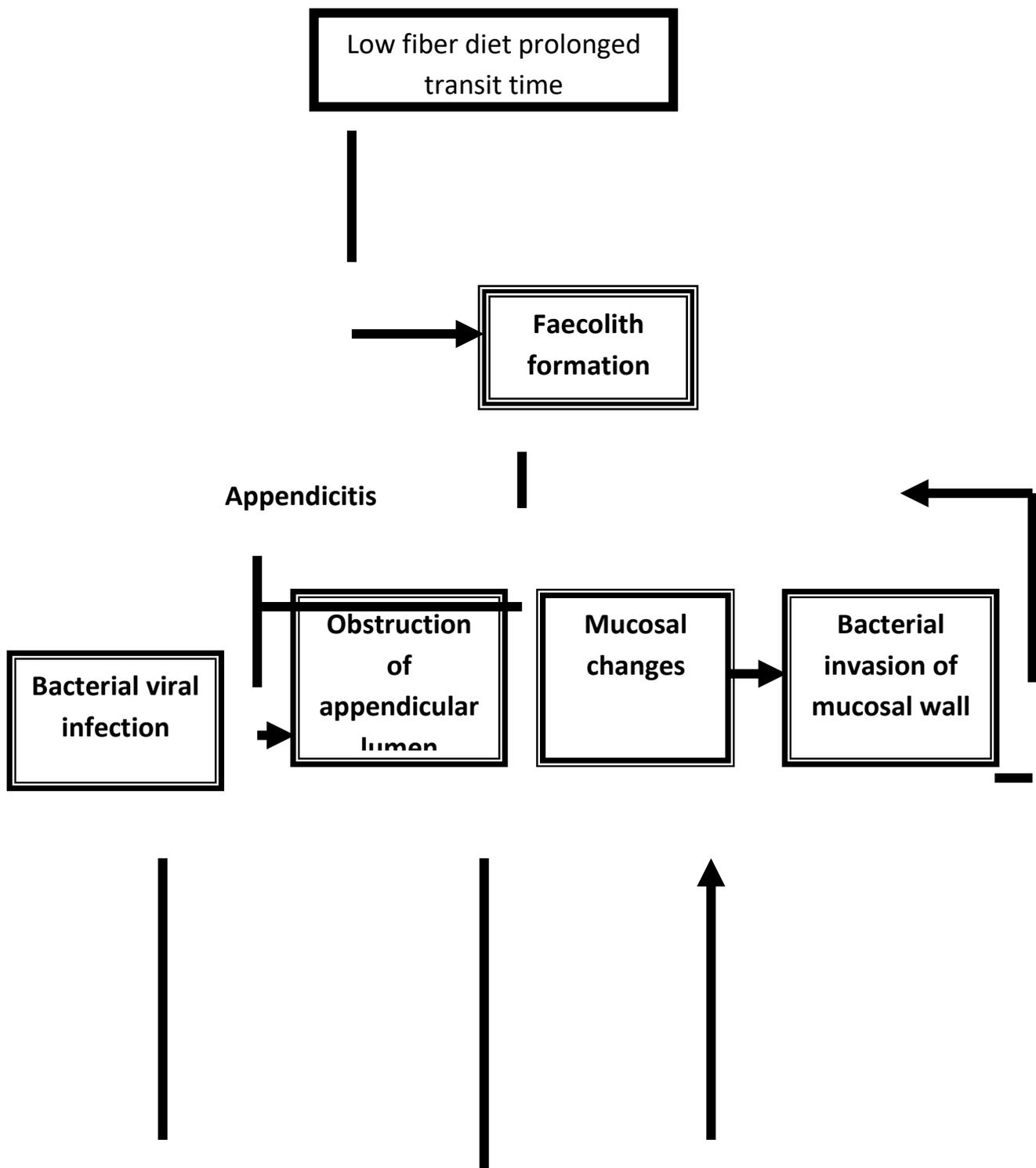
This in turn leads to high intra luminal pressure (Painter, 1969). The urban blacks have been reported to be relatively free of appendicitis (Bruner *et al.*, 1970; Lau *et al.*, 1984; Segal and Walker, 1986).

1-2-6-2 Genetic Factors

Larner (1988) described a family pedigree in which 80% of the members were operated on for appendicitis. Other studies have demonstrated a high incidence of appendicitis among immediate family members (sibling), parent or both not seen in a control group undergoing abdominal operations other appendectomy.

The explanation for this familial tendency is unclear, but the possible reason may include shared dietary habits (Anderson *et al.*, 1979), a genetic difference in resistance to bacterial infection (Larner, 1988), or the inheritance of a fibrous band of peritoneum which kinks the base of the appendix (Fig. 1-2). So partially occluding the lumen and thereby predisposing to appendicitis (Boulose and Cowiem 1993). Of the sporadic evidence currently available, all that can be said is that in certain cases some hereditary factors seem to be involved in the aetiology Larner, (1988). In (1990) Blandino *et al* pointed out the role of a major recessive gene. A follow up study by the same group studying 3rd generation of families using complex segregation analysis supported a polygenic or multifactorial model.

There are no clear evidences to support a major gene although a rare gene could not be ruled out as the cause of small proportion of cases. Specific studies to address genetic and environmental factor in this serious disease seem worth but for now a positive family history of appendicitis might join other evidence leading to improved clinical recognition of acute appendicitis. (Bast *et al*, 1990)



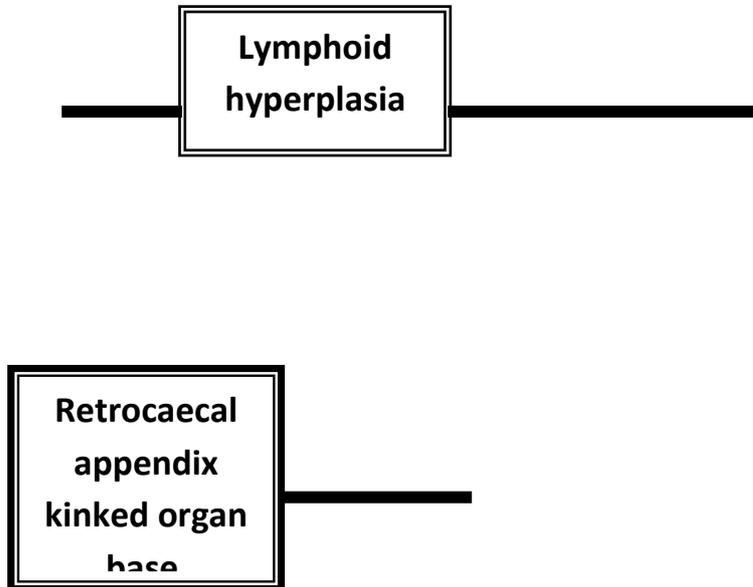


Fig. (1-2). Different theories for the aetiology of appendicitis (Larner, 1988).

1-2-7 Pathophysiology

Pathophysiology of acute appendicitis is most likely the result of closed loop obstruction of its lumen related to fecalith or less commonly resulting from hyperplasia of submucosal lymphoid follicles caused by a viral infection or rarely from obstruction due to pin worm or other helminthic infestation. Occasionally obstruction is caused by carcinoid tumor which if it is less than 1cm in size it requires no further therapy following appendectomy with reason

able margins in the presence of obstruction the mucosa continues to secrete some exudates resulting in an accumulation of mucoid material and increasing intraluminal pressure, bacteria located within the lumen of the appendix proliferate fast. In the presence of stasis and obstruction the flora of the appendix included both aerobic and anaerobic organisms (Lamps, 2004).

However not all luminal obstruction causes acute appendicitis and certainly not all acute appendicitis is caused by obstruction. Following obstruction the appendix continues to secrete into closed lumen causing increased luminal pressure with eventual venous congestion and infarction along the watershed region, eventual perforation is preceded by mucosal ulceration and transmural necrosis. Abscess formation or peritonitis (Hart, 1997).

1-2.8 Investigation

As a result of early diagnosis the case fatality rate has fallen to less than 0.1% for uncomplicated appendicitis (AL-Fahad, 2003). The records for gangrenous and perforated appendicitis are 0.6% and 0% respectively (Chsheri *et al.*, 1978; Sandler *et al.*, 1987). The accurate preoperative diagnosis of acute appendicitis remain a difficult problem, negative appendectomy rate of approximately 20% was commonly reported. In young ovulating women the incidence of gynecologic disease make the clinical diagnosis even more difficult and negative appendectomy rate between 34-46% have been

reported in such group of patient (Bind and Bahlgest, 1986). However diagnosis of acute appendicitis commonly follows two main investigations, clinical investigation and laboratory investigation. These are briefly presented below.

1.2.8.1 Laboratory Investigation

Laboratory investigation is mostly restricted on the white blood cell count (WBCS) since leukocytosis is seen in 70-80% of patients with acute appendicitis. Studies consistently show that 80-85% of adults with appendicitis have a WBC count density of $10,000$ cell/ cum^3 . Neutrophils greater than 70% occur in 78% of patients and 8% of patients with appendicitis have a WBC count less than $10,000$ cell/ cum^3 and neutrophil less than 70%. Moderate leukocytosis ranging from $10,000$ - $18,000$ cell/ cum^3 accompanied by a moderate polymorphonuclear predominance is the rule in acute uncomplicated appendicitis with normal total and differential while blood cells count, the diagnosis of acute appendicitis in the question through not ruled. (Clark *et al.*, 1992; Schwartz *et al.*, 1996).

If white blood cells count is more than $18,000$ cell/ cum^3 or if shift to left is extreme perforated or an acute inflammatory disease of greater magnitude then appendicitis is probable (Shiffoan and Greenes, 1991; Schwartz *et al.*, 1996).

1.2.8.2. C. Reactive Protein

In acute phase reactant C- reactive proteins synthesized by liver in response to bacterial infection- serum level begins to rise with 6 to 12 hours of acute tissue inflammation. When the symptoms have presented for at least 12 hour especially in children, c. reactive protein level was seen to be increased more than 1 mg/l in 72% of case .This can be considered as evidence of acute appendicitis (Owen and Willims, 1992; Anderson et al., 2004). In addition to the above examinations regarding the diagnosis of appendicitis some other examinations can be recommended such as urine analysis (Arubjornsson, 1988; Schwartz *et al.*, 1996), x ray (Sandler and Pearbery, 1987), Barium enema (Alvarado, 1986).

1.2.8.3. Other Investigation

1.2.8.4. Clinical Investigation

The diagnosis of acute appendicitis is the classic example of application of clinical skills. Ancillary laboratory and radiological tests are not essential in making the diagnosis. Signs and symptom ranking have been reported but these aids have not really improved the overall accuracy of preoperative diagnosis (Kalan *et al.*, 1994; Helmer, 2002).

However that following features may be considered during investigations:

- Symptoms include rightileac fossa pain, generalized pain, and fever, loss of appetite, nausea, constipation, vomiting, diarrhea, headache and dysuria.
- Signs: such as generalized tenderness, rebound tenderness, guarding, mass, rovsings sign and psoas sign.

1.2.9 Intestinal Flora

A vast microflora normally present in the gastrointestinal tract. At birth the baby's intestinal tract is sterile within few days, particularly in breast feclinfants, anaerobic lactobacilli and bifidobacteria appear and predominate 10^9-10^{11} cells / gram of feces along with fewer coliform and enterococci with broadening of diet. Coliforms become more numerous and are joined by *Bacteroides spp* and *Clostridium spp*. (Skinner & Carr, 1976; Fantin *et al.*, 1990). In adults the esophageal mucosa contains only the bacteria ingested with food and saliva. Because of low pH of the gastric juice, few bacteria mainly lactobacilli are found in the stomach, the jejunum has a sparse (10^4-10^6 cells/ ml of fluid predominantly gram positive bacteria the principle components being lactobacilli and various streptococci. The concentration of the microflora in the ileum increases 10^4-10^7 cells/ml with the appearance of coliforms and anaerobic organism $10^{11}-10^{12}$ cells /ml gram of feces, 96- 98% of the

colonic flora consists of anaerobic *Bacteroides spp.* mainly *Bacteroid fragilis*, anaerobic streptococci, *Bifidobacterium* and *Fusobacterium*: only 1 - 4% are aerobes and others are facultative anaerobic bacteria such as coliforms and enterococci. Also a small number of *Porteus spp* *Pseudomonas aeruginosa*, *Staphylococci*, *Spirochetes* and other microorganisms are present (Williams, 1973; Davis *et al.*, 1990).

Moderate changes in the diet do not affect the ratios of dominant bacterial species in feces. This situation is altered when gastrointestinal infection takes place (Moore *et al.*, 1969; Felner and Dowell, 1971; Vanway *et al.*, 1982). The dominant bacteria in human intestinal tract are discussed below.

1-2-9-1. Aerobic Enteric Bacteria

This is the family of a bacterial group commonly referred to as enteric group, they are Gram negative cocci bacilli normally habitat in the intestinal tract of human. Some of them act as a part of the normal flora and incidentally cause diseases while others are aggressive pathogen for human. They possess a complex antigenic structure and produce a variety of enzymes and toxins with other virulence factor. The members of this group are sharing many

characteristics with each other (Mims *et al.* 2004). Some common species among this group and other related bacteria briefly outlined below.

1.2.9.1.1 *E. coli*

Gram negative bacilli, some strains have pili as well as colonization factors antigen (CFA/I, CFA/II, CFA/III). The pili are responsible for adherence so that the adhesion has important role in pathogenesis (Matsumoto *et al.*, 1990). The capsule of *E. coli* represents the K- antigen which helps in the attachment of bacteria to the epithelial cells of the hosts (Eisentein and Azalenznik, 2000). *E. coli* may produce α and β haemolysin as apart of the virulence factors and some strains are urease producing (Collin and Falkow, 1990). (Saxen *et al.*, 1996) reported that colonization of the gut by virulent E-coli type λ c- fimbriated, P- fimbriated O₇₀: K^o: H^o serotype may lead to a perquisite for the development of appendicitis, The producing of green metallic sheen of colonies EMB the specific diagnostic character for this species. Agar is the specific diagnostic character for this species.

1.2.9.1.2 *Klebsiella*

Klebsiella species are Gram negative bacilli non motile, are specifically characterized by forming long and highly mucoid

colonies. The virulence of *Klebsiella* is not well understood, but its antiphagocytosis capsule plays a role in the infection by preventing phagocytosis. It is thought to be an aerobactin, an iron binding protein and the production of β - lactamase enzyme contributes to pathogenicity and resistance to antibiotics. (Thomson *et al.*; Mims, ۲۰۰۴).

۱.۲.۹.۱.۳. *Proteus spp*

This organism is mainly characterized by its active motility which resulting in what it called swarming phenomena it can cause a disease only when leave the intestinal tract and it produces the urease enzyme. The endotoxin liberated by this organism may play the main role in virulence of bacteria (Gilchrist, ۱۹۹۰; Farmer, ۲۰۰۰).

۱.۲.۹.۱.۴ *Enterobacter spp*

Gram- negative bacilli. They have several virulence factor, have a small capsule and siderophore production which contributes to the infection In addition, they are able to produce some enzymes and endotoxins which have an important role in virulence and resistance to antibiotics (Gilchrist, ۱۹۹۰). *Enterobacter spp* can produce antibacterial substance that has antagonistic activity against wide range of bacteria except *Acinetobacter* and *Pseudomonas* (Ya- Ping *et al.*, ۲۰۰۳).

۱.۲.۹.۱.۵ *Salmonella*

Gram negative bacilli, motile by peritrichous flagella. *Salmonella* spp are able to invade the intestinal mucosa and cause intestinal infection which can eventually lead to appear symptoms of typhoid fever. The 2200 known species of *salmonella* are classified according to their surface antigens. The capsular of this organism can cause serious complication in immunosuppressed individuals (Hanberger and Goberndo, 1999; Farmer *et al.*, 2000).

1.2.9.1.6 *Citrobacter freundii*

Gram negative bacilli, motile and invasive. *Citrobacter* infections are associated with high mortality rate. It can cause brain abscess, neonatal sepsis meningitis, blood stream infection and intra abdominal sepsis. *Citrobacter* species are commonly resistant to extended spectrum cephalosporins (Abbott, 1990; Graham and Baud, 1981; Jones, 2001).

1.2.9.1.7 *Pseudomonas aeruginosa*

It is Gram negative, and has several virulence factors very important in pathogenicity. These include capsule or slime layer which associates in adherence and effectively protects the bacteria from phagocytosis. The production of extra cellular protease,

cytotoxins and hemolysin have an important role in virulence in addition to the siderophore which enhances the growth of the organism. The soluble and media diffusible pigments produced by this organism are of important character to diagnose *Ps. aeruginosa* (Macfaddin, ۲۰۰۰).

۱.۲.۴.۱.۸ ***Staphylococcus aureus***

The pathogenicity of staphylococci is contributed to hemolysis of the blood, coagulation of plasma and production of extracellular enzymes and toxins (Mims *et al.*, ۲۰۰۴). *Staph aureus* has a polysaccharide capsule and the cell wall composed of peptidoglycan and teichoic acid moieties that protect it from lyses by osmotic condition and aid the bacteria to attach mucosal surface. The virulence of the bacteria resulted by secretion of toxins and enzymes which act on host cells membranes and mediated the cell destruction (Takahasi *et al.*, ۱۹۹۹).

۱.۲.۹.۳. **Anaerobic Bacteria**

۱.۲.۹.۳.۱. ***Clostridium***

Gram positive obligately anaerobic rod with center or subterminal heat resistant spores (Calo *et al.*, 1989). Although *Clostridium* species are ubiquitous in nature their principle habitats are the soil and the intestinal tracts of human and animal. The genus *clostridium* includes many species but the most three common species are *Clostridium perfringens* ,*Clostridium tetani* and *Clostridium botulinum* .The *Clostridium perfringens* is the species most commonly isolated from human clinical specimens excluding feces in the normal adult colon *clostridium perfringens* $10^3 - 10^6$ /g. It is encountered in a wide variety of clinical settings ranging from simple contamination of wound to traumatic or nontraumatic myonecrosis, clostridial cellulites, intra abdominal sepsis and gangrenous cholecystitis.

The clostridia form many virulent factors such as spores, many types of toxins and other extracellular products include collagenase and proteases. *Clostridium* species are commonly encountered in a variety of polymicrobial infection involving the abdomen including peritonitis, intraabdominal abscesses and septicemia in patient with obstructive or perforating lesions of the terminal ileum or large bowel (Gorbach *et al.*, 1998).

1.2.9.3.2- ***Peptostreptococcus* spp.**

They are gram positive anaerobic non spore forming cocci. These organisms are the most part components of the normal flora of the skin or mucosal surface of human. As with many other anaerobes they may become opportunistic pathogens when they are allowed to leave their normal habitat and gain access to normally sterile body tissue (Murray *et al.*, 1999).

Most (96%) *peptostreptococcus* isolates are susceptible to B. lactam antibiotics, clindomycin and metronidazole are slightly less active since it was reported that 84% - 88% of isolates were found to be resistant to these antibiotic respectively (Reig *et al.*, 1992; James *et al.* 2001).

1.2.9.2.3- Bacteroidaceae

It is obligately anaerobic non spore forming bacilli. The common species among this family is *Bacteroid fragillis* .The members of this family are straight, curve or helical. Some strains are motile. The Bacteroidaceae are prevalent among the indogenous flora of mucosal membranes of human. Anaerobic or mixed infection result from disruption of mucosal surface and introduction of anaerobes into (normally) sterile site (Baron and Mifinegold, 1992). Many of these organisms produce virulent factors that contribute to pathogenicity. This includes catalase, superoxide dismutase, collagenase,

neuraminidase, deoxyribonuclease (DNase), heparinase, protease and Immoglobulinase (Briselden *et al.*, 1992). Several strains produce capsule which promotes the abscess formation (Brook *et al.*, 1998). Enterotoxin production by some strains of *Bacteroids fragilis* has been recently described (Van *et al.*, 1992; Kenneth *et al.*, 2005) *Bacteroides* are resistant to penicillin and ampicillin. The anti pseudomonal penicillins, ticarcillin and mezlocillin are some what more active than penicillins (Sedlock *et al.* 1990; Cuchural *et al.*, 1996). Among the metronidazol, cephalosporins and cephamycin, cefoxitin remain very active against *Bacteroids* species (Aldridge *et al.*, 1997).

1.2.10 **Complication of Acute Appendicitis**

It has been pointed out that appendicitis and appendectomy may lead to produce some complications from of which the patients remain complain through the life. These complications are outlined below as concluded by (Paulson, 2003):

- Post operative infection.
- Intra abdominal abscess.
- Intestinal obstruction.
- Increase incidence of infertility in female.

1.2.11 **Management**

Appendectomy is seemed to be the suitable chosen management for most patients with acute appendicitis. The surgery often decided laparoscopically, is not complicated and carries little risk unless the appendix has ruptured. Generally the patient can resume normal activities quickly and will have no lingering problems. (Gorecki *et al.*, 1999) An inflamed appendix some times ruptures within the first 24 hours of symptoms. If the appendix does rupture, infection can spread to the neighboring tissue lining the abdomen (peritoneum) (Shelton, 2003).

The possibility of rupture is highest in young children and older adults. The diagnosis may be delayed for those people because their symptoms may not be as obvious as the symptoms of older children and younger adults. A ruptured appendix can be fatal if it is not managed early (Williams and MarGs, 2003).

Antibiotic could be administered preoperatively to control any local or generalized sepsis that may occur and to reduce the incidence of postoperative infection. The most common antibiotics being recommended for metronidazols the treatment of acute appendicitis are aminoglycosides, cephalosporins and metronidazols (Launrence *et al.*, 1997; Katazung, 2003), but sensitivity test may help in determine the appropriate antibiotic to be used for treatment .

1.2.12. Antibiotics Used to Treatment of Appendicitis

1.2.12.1. Aminoglycosides

Aminoglycosides antibiotic are in general active against staphylococci and aerobic Gram negative organism including almost all the members and Enterobacteriaceae even though some bacterial species. Emerged mutants resistant to aminoglycosides (Launrence *et al.*, 1997; Jones *et al.*, 2002).

1-2-12-2 Cephalosporins

All first three generation of cephalosporin have oral preparations that have used for treatment of most bacterial infection (Wilhelm and Edson, 1987; Forrest *et al.*, 1993).

First generation cephalosporins act by inhibiting bacterial cell wall synthesis the bactericidal activity is mainly against gram positive bacteria and the administration is either orally cefadroxil, cephacexin and cephradine or parenteral such as cephalothin, cephalazolin and cephradine (martinez *et al.*, 1990).

Now the usage of first generation cephalosporin. Is limited because of high resistance to it (Matinez *et al.*, 1990). Second generation cephalosporins have bactericidal activity that inhibits bacterial cell wall synthesis. It has greater activity against anaerobic bacteria and the administration is either oral (include cefaclor and cefuroxime) or parenteral second generation of cephalosporin has limited use that (Dumpis *et al.*, 2003).

Third generation of cephalosporins have bactericidal action that inhibits cell wall synthesis. They are highly stable in the presence of β -lactamase enzyme and they are effective in wide range of hospital acquired and nosocomial bacterial infections. The administration is either oral (cefixime, cefpodoxime and ceftibuten) or parenteral (cefotaxime, cefotriaxone, cefazidime and ceftizoxime) (Katzung, 2003).

1-2-12-3 **Metronidazol**

Metranidozol is antiprotozoal drug that also has potent antibacterial activity against anaerobes including *Bacteroides* and *Clostridium* species. It is well absorbed after oral administration widely distributed in tissue and reach serum level of 4-6 $\mu\text{g/ml}$ after 200 mg Oral dose metronidazole was suggested for treatment of anaerobic or mixed intra abdominal infections (Ya-ping *et al.*, 2003; Mims *et al.*, 2004).

1-2-13 Antibiotic Susceptibility Test Methods

Methods of bacteria antibiotic susceptibility test vary as regards accuracy of evaluating bacteria response to antibiotics and determining the boundary between sensitive and resistance against these antibiotics. Quantitative evaluation is frequently used to measure the bacteria susceptibility to antibiotics as it is more accurate and definitive than qualitative evaluation. Discovering the bacteria resistance to bacteria susceptibility represented by disk diffusion method (Murray *et al.*, 1999). Although the last method is not costly and quick, it is some times still unable to show the real level of bacteria susceptibility to antibiotics as it is the case in examining the anaerobic bacteria and the weakly growing bacteria in the solid culture media. Moreover the disk method is affected by some factors such as the growth density the type of bacteria used in the test, the type of antibiotics, concentration of the culture media and its degree of the hydrogenic exponent, beside the incubation period and temperature (WHO, 1987). The average of antibiotic spread in the culture medium is varies from one antibiotic to another (Murray *et al.*, 1999). All these factors may decrease the test efficiency using this method still the limitation of the minimum inhibitory concentration MIC of the antibiotic which is the lowest antibiotic concentration that inhibit the bacteria growth after a period of bacteria incubation in a series of dilution antibiotic in the

culture medium (Baron and Jone, 1994). The method is very important especially to the antibiotic rapidly resisting bacteria to observe the range of the resistance development through noticing the increase in the values of MIC (Murray *et al.*, 1999; Collee *et al.*, 1996).

MIC test is one of the significant tests used to determine the bacteria susceptibility when more than one antibiotic are mixed (Murray *et al.*, 1999).

This test is often used in surveying studies to examine susceptibility to antibiotic on a certain kind of bacteria specifically after determining the values of MIC₅₀ or MIC₉₀ of the antibiotic. Recently means devices and computer systems have been used to estimate the bacteria growth and to measure susceptibility to antibiotics and charging their results. This field called automated microbiology has witnessed great progress. Therefore, these technicalities have been used in microbiological Laboratories because they are quickly processed and give accurate results.

Results, learning out the old methods of susceptibility tests which are affected by some personal factors as eyesight problems, color blindness, fatigue, exhaustion and some technical mistakes by the people working in this field (Collee *et al.*, 1996). Sensitivity to antibiotics test by instrumental methods depends on different principles such as the turbidimetric bacterial growth in a liquid

medium or measuring the optical density. Other time if depends on colour and glittering reagents to decide the bacterial growth. Due to the difficulty in determining the bacterial growth in the MIC test. The photometric has been followed to obtain results compared other methods (Check, ۲۰۰۰). Moreover it was uneasy to do the MIC test in microbiological laboratories test (E) was used. It is one of the recent technologies used during the last years to decide the value of the MIC as soon as possible and accurately (E) test was designed in a way leads to discover antimicrobial resistance testing (ART) quickly and accurately (ART) test is also a means to discover the minimum changes at the level of the antibiotic bacterial susceptibility. Other applications of this test are the determination of the MIC value of both non aerobic and fastidious bacteria because it is not affected by factors as the bacterial growth density and the cells growth level. As the bacterial growth density and the cells growth level results can be read immediately from the test tape .It is also possible to use more than one type for more than one antibiotic in one test (Collee *et al.*, ۱۹۹۶; Barron and Jones, ۱۹۹۴).

۲.۱.۱. Patients

From October 2004 to July 2005, a total of 110 acute appendicitis patients who referred to Al- Furate Teaching hospital were investigated. Their ages ranged between 4-60 years. 69 of them were males and 41 were females. All patients were under went the following Laboratory investigations: General urine examination (GUE),white blood cell count (WBC) and c-reactive protein .Radiological investigation at abdominal ultrasonography and computerized tomographic (CT) scan being decided by radiologists were also considered.

2.1.2. Materials

Many types of instruments and chemical materials in addition to biological material materials were used in this study. The traditional apparatus devices being necessary for the appropriate experiments were carefully used. Culture media being used in this study are shown in table (2-1). Chemical substances and antibiotics disk are inserted in table (2-2) and (2-3) respectively. Table (2-4) and table (2-5) show the selected antibiotics for MIC and API diagnostic kit respectively.

Table (٢-١) List of cultures media

Culture medium	Company (origin)
Blood agar base	Mast diagnostic(USA)
Brain –Heart infusion broth	Mast diagnostic(USA)
Cooked meat broth	B.D.H (UK)
Eosine methylen blue	Mast diagnostic(USA)
Gas generating kit	Al- Raze (Iraq)
Kliglar Iron agar	Difco (USA)
Manitol Salt agar	Oxoid (UK)
MacConkey agar	Mast (UK)
MR- VP broth	Difco (USA)
Mueller. Hintor broth	Difco (USA)
Muneller. Hinton agar	Difco (USA)
Nutrient agar	Mast (USA)
Nutrient broth	Mast (USA)
Peptone water	Biolife (Italy)
Simmon citrate agar	Difco(USA)
Thioglyconate broth	B.D.H (UK)
Urea agar base	Difco(USA)

Table (۲-۲) List of chemical material

Chemical-materials	Company
Absolute Ethanol	BDH
BaCl₂	BDH
Crystal violet	Fluka
Formalin (4.0%)	BDH
Glycerol	BDH
HCl	BDH
H₂O₂ (3.0%)	Fluka
Iodin	BDH
KI	BDH
KOH	BDH
Kovac's reagent	BDH
Methyl red	Fluka
α -naphthol	BDH
Safranin	Fluka
Tetramethyl- p- paraphenylene diamine dihydrochloride	BDH
Urea. Solution	Fluka

Table (۲-۳) List of Antimicrobial agent

No	Antibiotic	Potency (μg)	Antibiotic	Company
۱	Amoxicillin	۱۰	Amx	Oxoid
۲	Gentamycin	۳۰	Gm	Oxoid
۳	Amikacin	۱۰	Ak	Oxoid
۴	Cefotriaxom	۳۰	CTX	RA
۵	Cephalexin	۳۰	CL	Oxoid
۶	Cefoxitin	۱۰	Cxm	Oxoid
۷	Ciprofloxacin	۲۰	CIP	RA
۸	Norfloxacin	۲۰	NF	RA
۹	Chlormphenicol	۱۰	C	Oxoid
۱۰	Augmentin	۳۰	AC	Oxoid
۱۱	Refampicin	۳۰	RD	Oxoid
۱۲	Vancomycin	۳۰	VA	Oxoid
۱۳	Tetracycline	۳۰	TE	Oxoid
۱۴	Co- Trimoxazole	۲۵	SXT	RD

Table (५-६) Antibiotic Used in MIC Test

Antibiotic	Company (origin)
Ceftriaxone	RA (India)
Cetotaxime	RA (India)
Ciprofloxacin	RA (India)
Amikacin	Oxoid (UK)
Augmentin	Oxoid (UK)

Table (५-७) Other materials

Material L	Company
API <i>staph</i>	Bio Merieux (franca)
API ५ · E and reagents (TDA, UP^१, UP^२, NIT)	Bio Merieux (franca)

2.2. Reagents

2.2.1. Methyl Red Reagent: It was prepared by dissolving 0.1 gm of methyl red was dissolved in 300 ml of 90 % ethanol and then the volume was completed to 500 ml with distilled water this reagent was used as indicator in methyl red (MacFaddin, 2000).

2.2.2. Voges-Proskauer Reagent: it included two reagents; A and B. Reagent A: 0gm of α -naphthol was dissolved in 100 ml of 99 % ethanol. Reagent B: 40 gm of KOH was dissolved in 100 ml of distilled water these reagents were used as indicators in voges- proskaur test (Collee *et al.*, 1996).

2.2.3. Oxidase Reagent: It was prepared by dissolving of 0.1 gm of tetra -p- paraphenylenediamine dihydrochloride in 10 ml of distill water and stored in a dark container this reagent was used as indicator in catalase test (Baron *et al.*, 1996).

2.2.4. Catalase Reagent: It was prepared by dissolving 3ml of H_2O_2 to 100 ml of distill water (Baron *et al.*, 1996).

۲.۳. Biochemical tests:

۲.۳.۱. Catalase test: a colony of the organism was transferred to a drop of ۳% H_2O_2 on a microscope slide the formation of gas bubbles indicate for the positive test (Collee *et al.*, ۱۹۹۶).

۲.۳.۲. Oxidase test:

A piece of filter paper was saturated with oxidase reagent. Then the colony to be tested was picked up with a sterile wooden stick and smeared over filter paper the organism under test was spread on this paper .when the colour around the smear turned from rose to deep purple color within ۱۰- ۱۵ second the oxidase test is positive (Collee *et al.*, ۱۹۹۶).

۲.۳.۳. Coagulase test:

Several colonies of bacteria were transferred with the a loop to test tube containing ۰.۵ ml of rabbit plasma. The tube was covered to prevent evaporation and incubated at ۳۷C for overnight .The test was read by tilting the tube and Observing for clot formation in the plasma. Negative test results in the plasma remained free flowing with no evidence of clot (Collee *et al.*, ۱۹۹۶).

۲.۳.۴. Indol test:

Peptone water was inoculated with a young bacterial colonies and incubated at ۳۷c° for ۲۴- ۴۸. Kovacs reagent (۰.۵ ml) was added.

A red color in the alcohol layer indicated a positive reaction (MacFaddin, 2000).

2.3.5. Methyl Red test:

The test was performed on 5 ml of MR-VP broth which inoculated by the organism and incubated at 37°C for 24 h. There 6-8 drops of methyl red reagent were added. To the culture, mixed and the result was red immediately the change of color to bright red indicates as a positive reaction (Collee *et al.*, 1996).

2.3.6. Simmon Citrate Test:

Simmons citrate slant was inoculated with bacterial culture and incubated 24-48 hours at 37°C. The positive result appears as change in the color of media from green to blue. The unchanging of the color indicates for a negative reaction (Benson, 1998).

2.3.7. Urease Test:

It was used to determine the ability of an organism to split urea, forming two molecules of ammonia by the action of the enzyme urease. It was prepared by adding 2.3 gram of urea base to 100 ml distilled water, then it was sterilized by autoclaving and cooled to about 50°C then 5 ml of 20% solution of sterile urea. Appearance of deep pink color indicates for positive result urea agar slant was

inoculated heavily over the entire slant surface and incubated at 37°C for 24 h. Urease test is positive if the indicator was changed to purple- pink (Benson, 1998).

2.3.8. Kligers Iron Agar Test:

A heavy inoculum was streaked over the surface of the slope and stabbed in to the buttm, incubated at 37°C for 24 h. Results were unformatted as follows:

<u>Slant/ Butt</u>	<u>Color</u>
Alkaline/ Acid	Red/ Yellow
Acid/ Acid	Yellow/ Yellow
Alkaline/ Alkaline	Red/ Red
H ₂ S	Black precipitate

It was used to detect three primary characteristics of bacterium: The ability to produce gas from the fermentation of sugars, the production of H₂S gas and the ability to ferment Lactose (Baron and Jones, 1994).

2.3.9. Methal- Red Vogas Proskauer:

MR.VP medium was inoculated with the colony of tested bacteria and incubated at 37°C for 48 hours. One ml of 40% KOH and 3ml of 0% solution of α – naphthol were added 1.16 and α- naphthol.

After 10 minutes, the formation of red color is indicative for the presence of acetoin (acetyl methyl carbinol) (Baron *et al.*, 1996).

2.4. Preparation of Culture and Diagnostic Media.

2.4.1. Ready- Prepared Media.

Media used in this study (listed in table 2-1) were prepared in accordance with the manufacturers instructions fixed on their container. All the above media were sterilized in the autoclave at 121°C for 15 min. After sterilization blood agar base was supplemented with 5% defibrinated human blood, and urea agar base was supplemented with 20% sterile urea solution all media were used for isolation and diagnosis of bacteria.

2.4.2. Laboratory Prepared Media.

2.4.2.1. Neomycin Blood Agar.

It was used for cultivation of *Clostridium perfringens*. It was prepared by adding Neomycin 10 mg / 100 ml of blood agar (Brook *et al.*, 1998).

2.4.2.2. Pepton Glucose Broth.

It was used as transport medium for the samples. It was prepared by dissolving 9 gram peptone glucose into 100 ml distilled water (Fredriksen *et al.*, 1996).

2.4.2.3. Kanamycin - Vancomycin Blood Agar:

It was used for cultivation of *Bacteroids* spp. It was prepared by kanamycin 100 mg and vancomycin 5.0 gm/1000 ml of blood agar (Brook *et al.*, 1998).

2.5. Culture Media:

2.5.1. MacConky Agar:

It was used for cultivation of gram negative bacteria and to differentiate the lactose fermenter and non Lactose fermenter gram negative bacilli (Baron *et al.*, 1994).

2.5.2. Nutrient Agar:

It is general medium. It was used for isolation of bacteria from their sources and for studies their morphological characteristics (Collee *et al.*, 1996).

2.5.3 Mannitol Salt Agar

It was used as selective and differential medium for *staphylococci* (Collee *et al.*, 1996).

۲.۵.۴. Eosine Methylene Blue (EMB) Agar

It was used as a differential medium for *E. coli* (Brooks *et al.*, ۱۹۹۸).

۲.۵.۵. Blood Agar

It was used for cultivation of fastidious microorganism and determining the hemolytic activity (Baron *et al.*, ۱۹۹۴). This medium was prepared by adding ۵% human blood to autoclaved blood agar base (Collee *et al.*, ۱۹۹۶).

۲.۵.۶. Gas Generating Kit

It was used for cultivation of anaerobic Microbes (Collee. *et al.*, ۱۹۹۶).

۲.۵.۷. Nutrient Broth

It was used in general experiments and maintain the bacterial isolates.

۲.۵.۸. Cooked Meat Broth (CMB).

It was used for cultivating of anaerobes (Baron *et al.*, ۱۹۹۴).

۲.۵.۹. Muller-Hinton Agar:

It was used to determine the antimicrobial sensitivity patterns (Bauer *et al.*, ۱۹۶۶).

2.5.10. Thioglyconate Broth:

It was used for supporting growth of anaerobes (Baron *et al.*, 1996).

2.5.11. McFarland Standard Solution (tube no 0.5)

It was used for antimicrobials susceptibility testing. It was prepared according to the method suggested by Collee *et al.*, (1996).

as follows -BaCl₂ 1% 0.05%ml

-H₂SO₄ 10% 9.90 ml

2.6. Specimens Collection

One hundred ten excised appendixes were collected from 110 patients who were referred to AL-Furate AL-Awsate Teaching Hospital for (suspected) acute appendicitis through a period of ten months (from October 2004 to July 2005). The collected specimens were transported straight a way to Laboratory for bacteriological analysis as it is illustrated below.

2.7. Sterilization of the Outer Surface of the Appendix.

The outer surface of appendix was sterilized by three methods, ethyl alcohol 70%, iodine solution and heat cauterization (Fredriksen *et al.*, 1996).

2.7.1. Ethyl Alcohol 70%

The appendix was totally immersed in beaker contained ethyl alcohol 70% and immediately cultured on the proper media under aerobic and anaerobic conditions (Fredriksen *et al.*, 1996).

2.7.2. Iodin Solution 10%.

In this method, the appendix was totally immersed in the iodine solution (10%) for 5 minutes. The excessive solution was then absorbed by sterile filter paper. The tissue was then sectioned by sterile lancet and a swab from the inner part was cultured aerobically on blood agar, MacConky agar and anaerobically on kanamycin-vancomycin blood agar (KvA) and neomycin blood agar (NBA).

2.7.3. Heat Sterilization.

In this method, a metal spatula was used. The spatula was flamed first and then appendix was cauterized by it. The appendix which is supposed to be sterile was then anatomized by sharp Lancet and a swab from the inner tissue was cultured aerobic and anaerobic on the appropriate culture media.

2.8. Identification of Bacteria:

For identification and diagnosis of bacterial isolates, the following criteria were considered, colonial, cellular morphology, and biochemical reaction. Diagnosis of some isolates was further confirmed by application of API-system. The results of tests were

compared with standard result according to Bergy's manual for determinative bacteriology (Holt *et al.*, 1994; Collee *et al.*, 1996; MacFaddin, 2000; Baron *et al.*, 1996).

2.9. Antibiotic Sensitivity Test.

2.9.1. Disk Diffusion Technique.

It was performed following Kirby-Bauer method by using a pure culture of previously identified bacterial isolates. The inocula to be used in this test was prepared by suspending selected colonies grown on blood agar plate in 2 ml of broth to a cells density (2×10^6 cell/ml) equal to the turbidity of McFarland tube No 0.5. A sterile cotton swab was used to spread a proper inoculum on the standardized Muller Hinton plate. The antibiotic discs were placed on the surface of the medium at evenly spaced intervals with flamed forceps. After 24 hrs incubation at 37 °C the antibiotics inhibition zones were measured in comparison with standard results measurement of NCCLS (1999) to determine the susceptibility of the tested organisms to each antibiotic.

2.9.2. Determination of Minimum Inhibitory Concentration (MIC).

The MICs tests were determined according to the method suggested by (Barry, 1976; Murray *et al.*, 1999) as follows.

۲.۹.۲.۱. Preparation of Bacterial Suspension.

Bacterial suspension was prepared from liquid culture (۱۸-۲۰) hrs. old for each bacterium(^ types) the suspension was diluted to ۱/۱۰ in BHI broth for Gram-negative bacilli and ۱/۴ for Gram positive ۱۰⁷ cfu/ml (Murray & cocci for obtaining cell density approximately ۱۰^۶ *et al.*, ۱۹۹۹).

۲.۹.۲.۲. Preparation of Antibiotic Solution

Determination of minimum inhibition concentration was studies against the initial dilutions of the antibiotic used in this study were prepared from the original stock solution by using the two folds technique for five antibiotics.

۲.۹.۲.۳. MIC Test.

By using a plastic microdilution tray, ۱۰۰ μl from the required concentration of the Muller- Hinton broth were inoculated by ۱۰۰ μl from the required bacterial suspension then ۱۰۰ μl from antibiotic solution were added to the first well and other serial dilutions were made in order to obtain double concentration for all wells .A control was also prepared by ۱۰ ml of each bacterial suspension, antibiotic solution and muller- Hinton broth. A control positive containing of broth without antimicrobe, a control negative inoculated and helped in the refrigerator over night to serve as a negative control. The tray

was closed with its special lid and incubated at 37°C for 16- 18 hr, the concentration which gives no bacterial growth inhibition of bacteria 100%, is regarded as MIC.

2.9. Statistical Analysis

The data were analyzed statistically using Chi-square(χ^2) test and Z-test at the 1% level (Daniel, 1978).

3.1. Samples Collection

The study concerned first with the detection of the appropriate medium that can enhance the growth of bacteria being present in excised appendix even in few numbers. Consequently, three types of culture media were suggested. Those were BHI-broth, nutrient broth supplemented with 0.9% NaCl and peptone water supplemented with 0% glucose solution (PGM). Experimentally, the medium (PGM) revealed quite satisfactory results compared with other two media, since it remarkably promoted the growth of all bacterial types. This result was in agreement with (Hussien, 1999). Accordingly PGM was chosen as preliminary supportive medium for prepare the infective bacteria in further necessary experiments.

3.2 . Sterilization of Appendix

As with culture media, previously described, three methods were tried for sterilization of the outer surface of appendix in order to, investigate the samples properly with out any suspected contamination being gained during the process of appendectomy. Those methods were application of ethyl alcohol 70%, application of iodine 10% and heated spatula (cauterization).The results revealed that cauterization was the most effective and useful method for the sterilization of appendix. The results of this experiment were in agreement with that obtained by Gibbous & Houte (1970), Beachey (1981), Brodsky *et al* (1991) and Hussien (1999).

3.3 . Clinical Features Related to Acute Appendicitis

The subjects included in were complaining from various symptoms are shown in table (3-1), 70(63.6%) showed right iliac fossa pain, increased fever was seen in 30(31.8%) patients, while 31(28.1%) patient were with nausea. Other features were as follows; headache 17(15.5%), constipation 16(14.5%), generalized pain 14(12.7%), diarrhea 11(10%), loss of appetite 6(5.5%), vomiting 6(5.5%) and dysuria 2 (1.8%). From the results described above one can concluded that the right iliac fosse pain is the commonest symptom which may draw attention to the case as suspected appendicitis. Those results were in accordance with related study

Kosloske (୧୦୦୧) who pointed out that the right iliac fosse pain was the common feature of appendicitis.

Table (୩-୧) Distribution of the acute appendicitis according to the clinical features (n=୧୧୦)

Clinical feature	No. (%)
Right iliac fossa pain	୮୦ (୭୩.୬)
Fever	୩୦ (୩୧.୮)
Nausea	୩୧ (୨୮.୧)
Headache	୧୮ (୧୬.୫)
Constipation	୧୬ (୧୫.୫)
Generalized pain	୧୫ (୧୩.୬)
Diarrhea	୧୧ (୧୦)

Loss of appetite	٦ (٥.٥)
Vomitting	٦(٥.٥)
Dysuria	٢ (١.٨)

٣.٤ Risk Factors

٣.٤.١ Age Related Disease

The distributions of patients with acute appendicitis according to their ages are shown in table (٣-٢). The results revealed a significant differences upon age factor [cal. $\chi^2 = ١٨.٨$, tab $x = ٧.٨١$, $p < (٠.٠٥)$] since the results indicated that the peak of incidences observed in the age group (١١-٢٠) years old, that the frequency of infection accounted for ٥٧(٥١.٨%) patients, ٣٩(٣٥.٤%) of them were males and ١٨(١٦.٣%) were females. The results indicate that there was an

increase in the incidences with the advancement of age, since the lowest rate 2(1.8%) was seen within the age group of (01-10) old.

The patients of 11-20 years old being more susceptible for appendicitis may ascribed to the nature of physiological and anatomical factors of appendix tissue(Jones, 2002), since lymphoid tissue is the most susceptible for infection gradually increase up to the 20 years of age. The lymphoid tissue begins to decrease with advancement of the age up to 60 years old when this tissue is totally disappear. In addition to the configuration of the appendix at the age of group of (11-20) years due to increase infection with acute appendicitis (Jones, 2001).

Table (3-2) Distribution of acute appendicitis according to the age and sex

Age(year)	No of patient (%)	No of males (%)	No of females (%)
1-10	3 (2.7)	1 (0.9)	2 (1.8)
11-20	57 (51.8)	39 (35.4)	18 (16.3)
21-30	33 (30)	20 (18.1)	13 (11.8)

٣١-٤٠	٩ (٨.١)	٥ (٤.٥)	٤ (٣.٦)
٤١-٥٠	٦ (٥.٤)	٣ (٢.٧)	٣ (٢.٧)
٥١-٦٠	٢ (١.٨)	١ (٠.٩)	١ (٠.٩)
Total	١١٠	٦٩(٦٢.٧)	٤١(٣٧.٣)

٣.٤.٢. Sex Related Disease

The occurrence of acute appendicitis was found to be higher in males than females in (Table ٣-٢). The results indicated that males (٠.٠٩) more <are significantly (Cal. $Z=٢.١٢٦$, tab $Z=٠.٩٨٤$, p

susceptible 69(62.7%) than females 31(37.3%) with ratio of male to female accounted for 1.7:1. The same results were being reported by Addiss *et al.*, (1990) and al Fahad (2003) since, it has been found the diagnosis of acute appendicitis in males is more reliable compared with females that, the females remain a difficult to diagnose and misdiagnosis may result due to other diseases such as gynecologic diseases.

3.4.3. Seasonal Variations and Acute Appendicitis

Table (3-3) illustrates the occurrence of appendicitis according to the seasonal variations. The results indicated that the prevalence were considerably increased during the hot season, beginning through May which account 14(12.7%) cases then June 18(16.4%) cases and July 26(23.6%), whereas a remarkable decreased incidences were seen during the cold season represent by January, 4(3.6%) cases and February 0(0.0) cases. Rothrock *et al.*, (2000) reported closely similar results but the explanation for such results are rare. However this study may conclude that and extreme change in temperatures may have a role in an increasing the incidences during the warm season in addition to other factors such as type of diet and water (for drinking and swimming) may cause the occurring of bacterial and viral infections which may lead to appendicitis (LuckMann and Davis, 1991; Rothrock *et al.*, 2000).

Table (3-3) Seasonal Variations of Acute Appendicitis

Month		NO. Of patient (%)
2004	October	10 (9.1)
	November	7 (6.4)
	December	6 (5.5)
April	January	4 (3.6)
	February	5 (4.5)
	March	9 (8.2)
	April	11 (10)
	May	14 (12.7)
	June	18 (16.4)
	July	26 (23.6)
Total (%)		110 (100%)

3.5. Isolation and Identification

In this study a total of eleven bacterial genera were found to be implicated in acute appendicitis. Three genera were Gram positive

(Table 3-4) and eight genera were Gram negative (table 3-5). Those isolates were isolated and identified according to the cellular morphology, cultural characteristics and biochemical tests for each organism and compared with the referential results recommended by Holt *et al.*, (1994) Baron *et al.*, (1996); Benson (1998) and McFadden (2000).

3.6. Isolation Bacteria From Acute Appendicitis

Table (3-6) shows that out of 110 specimens (appendix swabs), of which 90 (81.8%) specimens yielded positive results for bacterial growth, while 20 (18.2%) specimens yielded no growth. The same results seen by Lau- WY *et al.*, (1994) and Cuss& Richard, (2000) who stated that most of bacterial cultures for suspected appendicitis were positive. In this study, a total of eleven bacterial appendicitis. Eight genera were Gram- negative and 3 genera were Gram- positive as shown in table (3-7), Gram- negative bacteria were the common

cause of appendicitis compared with that of Gram positive, they were 107(96.4%) isolates and 4(3.6%) isolates respectively. Moreover 21(18.9%) cases revealed mixed growth (more than one caustive agent).The most frequent pathogen was *E. coli* which accounted for 36(32.4%) isolates, followed by *Bacteroides* spp isolates (21, 19%).

Table (3-6) Results and Swab Culture in Males and Females Suffering From Appendicitis

Cultural- result	No of cases	No.(%) of	
		Males	Females
Positive	90(81.8)	60(59.1)	20(22.7)
Negative	20(18.2)	4(3.6)	16(14.6)
Total	110(100)	64(62.7)	41(37.3)

Table (3-7) Types and Frequency of Bacteria Isolated From In 90 Patients Suffering From Appendicitis.

Type of bacteria	No. (%) of Bacteria
Gram- negative n=107	
<i>Esherichia coli</i>	36(32.4)
<i>Bacteriodes spp</i>	21(18.9)
<i>Klebsiella pneumoniae</i>	18(16.2)
<i>Pseudomonas aeruginosa</i>	11(9.9)
<i>Citrobacter frundii</i>	7(6.3)

<i>Salmonella typhi</i>	0(4.0)
<i>Proteus mirabilis</i>	0(4.0)
<i>Enterobacter aerogenes</i>	4(3.6)
Gram positive n=4	
<i>Peptostreptococcus sp</i>	2(1.8)
<i>Clostridium perfringnes</i>	1(0.9)
<i>Staphylococcus aureus</i>	1(0.9)
Mixed growth	21(18.9)
Total	131

18(16.2%) isolates of *Klebsiella pneumoniae*, 11(10%) isolates of *Pseudomonas aeruginosa*, 7(6.3%) isolates of *Citrobacter freundii*, 0(4.0%) isolates of *Salmonella typhi*, 0(4.0%) of *Proteus mirabilis* and 4(3.6%) isolates of *Enterobacter aerogenes*.

Peptostreptococcus spp was the most frequent microorganism among Gram-positive bacteria which accounted as 2(1.8%). Whereas each of *staphylococcus aureus* and *Clostridium perfringnes* were accounted as 1(0.9%) as presented in table (3-8).

The results being found were in accordance with other results being reported by Elhag *et al.*, (1986) and Baste *et al.*, (1999). Those results were accepted and suspected since *E. coli* is the common

organism present in intestine fastly proliferates and quickly adheres to the tissue surfaces (Baron *et al.*, 1992).

The adhesion of microorganism to the epithelial cells of the tissue is the first stage of infection followed by the invasive stage. Moreover *E. coli* has others virulence factors such as host cell surface- modifying factors, toxins, hemolysin and cytotoxin necrotizing factor type I (CNFI). (Martirosian *et al.*, 2001).

Bacteroides has been also described to possess several factors capable of participating in intrabdominal infection. The *Bacteroides* Contribute to develop an infections in three ways, stimulation of abscess formation, reduction of phagocytes by polymorphnuclear leukocytes (PMNS) because of the capsule of *Bacteroidis* and inactivation of antibiotics by B-lactamase production(Kenneth *et al.*, 2003).

Klebsiella pneumonia isolates were 18(16.3%). This organism has the Capsule which plays an important role during the initial steps of the pathogenesis by interacting with mucus producing cells, the colonization of mucus membranes by bacteria is liked to an adhesion process involving specific adhesions on the bacterial surface. In addition to several pilli involved in adhesion to epithelial cell of intestine (Podschun and Sahly 1991; mims, *et al.*, 2004).

The explanation for detection of *Pseudomonas aeruginosa* in appendix as causative agent of appendicitis may be attributed to the ability of this organism to adhere and to colonize the epithelial tissues probably by pili and by the alginate (slime) layer surrounding the cells of this bacterium (Zhu *et al.*, 2004). Furthermore *Pseudomonas aeruginosa* possesses other virulence factors (enzymes and toxins, which enable it to cause infection. Other Gram-negative *Citrobacter freundii*, *Salmonella typhi*, *Proteus mirabilis* and *Enterobacter aerogenes* were also detected in appendicitis although in low frequencies compared with other Gram negative (table, 3-7).

However, the implication of these bacteria in appendicitis is suspected, since they belong to the enteric group and frequently present in intestine and all have virulence factors enabling them to cause disease (Abbott *et al.*, 1999). In this study, some of Gram positive members also represented by *Staphylococcus aureus*, *Peptostreptococcus* sp and *Clostridium perfringens* were also isolated and identified from appendicitis cases, but in low frequencies in relative with that of Gram negative.

Members (Table 3-8) Gram positive appendicitis are rarely reported at the present time. This may be due to adhesive and colonizer factors being less in Gram positive compared with that of Gram negative- moreover most of Gram positive bacteria are fastidious require for special growth factors (amino acids, vitamin, etc) and

growth condition (O₂, CO₂ ...etc) (Mims *et al.*, 2004). However qualitatively, Gram positive infections are more serious and the detection of *Clostridium perfringens* in appendicitis by this study may can be the announcement that even the strict obligate anaerobic necrotizer gas gangrene producer *Clostridium perfringens* can implicate in appendix infection.

3.7. Mixed Bacterial Growth

In this study, anaerobes, Gram negative and Gram positive were frequently isolated from mixed infection with facultative aerobic bacteria and presented in 21 isolates (18.9%). Table (3-8). This pattern of mixed culture between aerobic and anaerobic bacteria may be attributed to the synergistic relationship between aerobes and anaerobes, in which the aerobic bacteria grow first, exhausting oxygen and producing intermediates products may help in growing of the anaerobic bacteria .(James *et al.*, 2001).

Table: 3-8 Appendicitis by Mixed Bacterial Types.

Type of Bacteria	NO. (%)
<i>E .coli</i> <i>Bacteroides sp</i>	7(33.3)
<i>Klebsiella pneumoniae</i> <i>Bacteroides spp</i>	4(19)

<i>E. coli</i> <i>Citrobacter frundii</i>	3 (14.2)
<i>Klebsiella pneumoniae</i> <i>E. coli</i>	2 (9.0)
<i>E. coli</i> <i>Enterobacter aerogenes</i>	2 (9.0)
<i>Proteus mirabilis</i> <i>Pseudomonas aeruginosa</i>	1 (4.5)
<i>E. coli</i> <i>Peptostreptococcus spp</i>	1 (4.5)
<i>Pseudomonas aeruginosa</i> <i>Clostridium perfringens</i>	1 (4.5)
Total	21 (100)

3.8. Susceptibility to Antimicrobial Agents

The susceptibility of all isolates being detected in this study to ten (10) traditional antimicrobial agents was determined and are shown in table (3-10). The results indicated that *Pseudomonas aeruginosa* was the most important bacteria resistant for these antibiotics. All 11 isolates (52%) exhibited considerable resistance toward amoxicillin, Gentamycin, Cefotaxime, Cefixime, trimethoprim, vancomycin, norfloxacin and tetracyclin, while 10 isolates (47.6%)

were resistance to ciprofloxacin and 9 isolates (88.8%) resistance to amikacin. 21 isolates (100%) of *Bacteroides* sp resistance to trimethoprim, vancomycin, norfloxacin and tetracycline, while 19 isolates (90.4%) showed resistance to amoxicillin, Gentamycin and amikacin. However isolates 0 cases (23.8%) exhibited resistance to ciprofloxacin, cefotaxime and cefixme, 0 isolates (100%) of *Salmonella typhi* resistance for Gramycin, amikacin, cefotaxime, cefixme, vancomycin norfloxacin and tetracycline, while 4 isolates (80%) were resistance to amoxycillin and trimethoprim and only 1 isolates (20%) resisted ciprofoxacin. 18 cases of *Klebsiella pneumonia* were resistant to vancomycin and norfloxacin, while 14 isolates (94.4%) resistance to amoxicillin, trimethoprim and tetracycline. In addition to 4 isolates (38.8%) were resistance to Gentamycin and amikacin and 0 isolates (27.7%) were resistant to ciprofloxacin, cefotaxime and cefixie. Five isolates (100%) of *Proteus mirabilis* were resistant of vancomycin and tetracycline, while 3 isolates (60%) exhibited resistance for amoxicillin, Gentamycin, amikacin, trimethoprim and norfloxacin in addition to 1 isolates (20%) resistance to ciprofloxacin, cefotaxime, most isolate of *E. coli* revealed high sensitive to most of tested antibiotics. Such results were true for *Enterobacter aeruginosa* as well are 2 isolates (00%) resistance for trimethoprim and norfloxacin while 3 isolates (60%) resistance to amoxycillin, Gentamycin, amikacin, ciprofloxacin, cefotaxime and cefixme, 1 isolates (100%) of *Clostridium perfringes*

resistance for amoxicillin, Gentamycin, amikacin, trimethoprim, norfloxacin and tetracycline. The isolates of *S. aureus* was resistant to trimethoprim, vancomycin and amoxycillin. The results showed that *Pseudomonas aeruginosa* was the most pathogen resistant toward the most tested antibiotics. Since it resisted amoxicillin, Garamycin, cefotaxime, cefixime, trimethoprim, vancomycin, norfloxacin and tetracycline, in a remarkable level. Resistance mediated by *Pseudomonas aeruginosa* can be attributed both to an inducible, chromosomally mediated β lactamase that render broad spectrum cephalosporins to inactive, and to plasmid mediated beta lactamase that can lead to resistance to several penicillins and older cephalosporins (Cross and Campbell, 1999). In addition other factors may enable *Pseudomonas aeruginosa* to resist antimicrobial agents such as, altered and membrane site, nature of cell wall and alginate layer surrounding the bacterial cell (Mandell, 1999; Norrby, 1997). Moreover *Pseudomonas aeruginosa* having lipopolysaccharide (LPS) permeability barrier connected with complex proteins in their outer membrane, which are preventable passing of antimicrobial agents to the inside of the bacterial cell (Moore *et al.*, 1969; Jones *et al.*, 2001). Gram negative bacilli (*Klebsiella pneumoniae* and *Citrobacter freundii*) have broad spectrum resistance for antimicrobial agents. The production of β lactamase is the main mechanism for this resistance (Yamaguchi *et al.*, 1999; Ya *et al.*, 1999; Robert, 1999).

Additionally other mechanisms may enable the bacteria to decrease the permeability of cell membrane (Katzung, ۲۰۰۳; Mims *et al.*, ۲۰۰۴). This may be true for *Protous mirabilis* and *E. coli*, since they showed relative resistance for many of the antimicrobial agents.

The high sensitivity of *Staphelococcus aureus* to most antimicrobial agents (except trimethoprim and tetracycline) is an expected result to these bacteria, since *S. aureus* belong to the Gram positive bacteria which has been characterized to be sensitive to most common antibiotics in comparison with Gram negative bacteria due to the difference in the outer membrane structure which appears to be permeable to most antibiotics in gram positive bacteria than gram negative bacteria (Brook *et al.*, ۱۹۹۸).

۳.۹ Determination of Minimum Inhibitory Concentration (MIC)

In this study, five antimicrobial agents were used to determine the MIC against bacterial isolates. Those antimicrobial agents were cefotaxime, amikacin, amoxiclave, cefotriaxon and ciprofloxacin as shown in table (3-10). The results were recorded according to NCCLS (1999). It was found that amoxiclave and amikacin were most effective antibiotics against *E. coli*, since the MIC ranged from 0.5 - 16 µg/ml for amoxiclave and 0.5 - 8 µg/ml for ciprofloxacin. *Proteus mirabilis* exhibited more sensitive for cefotriaxon and ciprofloxacin, since the MIC ranged from 2-32 µg/ml for cefotriaxon and 1-8 µg/ml for ciprofloxacin.

Klebsiella pneumoniae revealed high sensitivity for cefotaxime and ciprofloxacin, since their MIC ranged from 8-128 µg/ml and 1-8 µg/ml respectively. *Enterobacter aerogenes* was highly sensitive to cefotriaxon, cefotaxime and ciprofloxacin, since their MIC ranged from 0.125- 8 µg/ml, 0.125-8 µg/ml and 0.125 - 0.5 µg/ml respectively.

Citrobacter freundii exhibited more sensitivity for amoxiclave and cefotaxime, since their MIC ranged from 0.25- 128 µg/ml and 0.008- 128 µg/ml respectively, while ciprofloxacin and cefotriaxon were the most effective against *Salmonella typhi*, since their MIC ranged from (0.5-32 and 0.5-2 µg/ml respectively).

Pseudomonas aeruginosa was the highest resistant for most antibiotics, except the ciprofloxacin which was the more effective

against this bacteria (Jones *et al.*, 2002), since the MIC of this antibiotic ranged from (0.06-1 µg / ml) for ciprofloxacin. The result of this method which in accordance the result of disk diffusion method used in this study.

Conclusion

- ١- **Appendicitis can be resulted due to bacterial infection of appendix.**
- ٢- **Gram positive & negative (aerobic & anaerobic) were implicated in appendicitis.**
- ٣- **Gram negative bacteria were more common in appendicitis than Gram positive.**
- ٤- ***E. coli* constitutes the predominant bacteria in appendix.**

- ④- The age group ranges from 11 to 20 years are more susceptible for appendicitis.
- ⑤- The occurrence of appendicitis increases during the warm season.
- ⑥- Women are some times misdiagnosed as appendicitis leading to unnecessary appendectomy.

Recommendations

- ١ - Achievement of further studies to detect serologically the
early diagnosis of appendicitis.
- ٢ - Achievement of further studies to detect the role of diet type
and nature in appendicitis occurrence
- ٣ - Careful management with women complaining from
symptoms suspected for appendicitis
- ٤ - Search for other member of quinolones and new generation
cephalosporins that are used to prevent complication.
 - ٥ - Working on viral appendicitis.

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Table (३-४) Biochemical tests used for the diagnosis of Gram- positive bacteria

Tests Types of Bacteria (No.)	Aerobic growth	Anaerobic growth	Motility	Oxidase test	Catalase test	Hemolysis	Pigment formation	Fermentation of		Coagulase test
	Glucose	manitol								
<i>S. aureus</i> (1)	+		-	-	+	+	+	+	+	+
<i>Peptostreptococcus sp</i> (2)	-	+	-	-	-d	-	-			
<i>Cl. Perfringes</i> (1)	-	+	-	-	-	+				
d. different * not done										

Table (3-0) Biochemical Tests used for the Diagnosis of Gram- negative bacteria

Aero- bic	Anae	Oxide	Indol	Meth	Voges	Citra- te	Ureas e	Catal	Reaction On KIA:	Motil- ity	Hemo- lysis	Grow- th on	O - F gluco	Fermentation of
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Tests											Slant/ bupp	Gas	H ₂ S											Glucose	Lactose	Manitol	Arabinose	Rhamnose	Trehiaose
	Type of (No.)																												
<i>E. coli</i> (27)	+	+	-	+	+	-	-	-	+	A/A	+	-	+	+	+	F	+	+	+	+	+								
<i>K. pneumoniae</i> (18)	+	+	-	-	-	+	+	+	+	A/A	+	-	-	-	+	F	+	+	+	+	+								
<i>P. aeruginosa</i> (11)	+	-	+	-	-	-	+	-	+	K/K	-	-	+	+	+	O	+	-											
<i>Mirabilis</i> (2)	+	+	-	-	+	-			+	K/A	V	+	+		+	F	+	-	-	-	-								
<i>St. frundii</i> (1)	+	+	-	-	+	-	+	-	+	K/A		+	+	-	+		+	V	+	+									
<i>S. typhi</i> (2)	+	-	-	-	+	-	-	-	+	K/A	-	+	+	-	+														
<i>Proteobacter. genes</i> (2)	+	+	-					V	+	A/A	-	-	+		+		+												
<i>Acidobacter. Spp</i> (21)	-	+	-	-				V		A/A	-	V	-	+	-	F	+	+					V						

KIA= kligler iron agar K= alkaline A= acid V= variable *= not done

Table (3-9) The Numbers and Percentage Resistance of 111 Bacterial to 10 Antibacterial Agents

Type of bacteria (No)	NO of isolate	AMX (%)	GM (%)	AK (%)	CF (%)	CE (%)	CXM (%)	TMX (%)
<i>E. coli</i>	27	29(100)	14 (38.8)	11 (30.5)	16 (44.4)	10 (29.6)	10 (29.6)	7 (20.6)
<i>K. pneumoniae</i>	18	17 (94.4)	7 (38.8)	7 (38.8)	0 (27.7)	0 (27.7)	0 (27.7)	0 (27.7)
<i>P. aeruginosa</i>	11	11 (100)	11 (100)	9 (81.8)	10 (90.9)	11 (100)	11 (100)	10 (90.9)

<i>C. frundii</i>	7	6 (85.7)	5 (71.4)	5 (71.4)	4 (57.1)	4 (57.1)	5 (71.4)	(85.7)
<i>P. mirabilis</i>	5	3 (60)	3 (60)	3 (60)	1 (20)	1 (20)	2 (40)	(60)
<i>S. typhi</i>	5	4 (80)	5 (100)	5 (100)	1 (20)	5 (100)	4 (80)	(100)
<i>E. aerogenes</i>	4	3 (75)	3 (75)	3 (75)	0	0	3 (75)	(75)
<i>S. aureus</i>	1	0	0	0	0	0	0	0
<i>Bacteroides spp</i>	21	19 (90.4)	19 (90.4)	19 (90.4)	0 (23.8)	0 (23.8)	0 (23.8)	(90.4)
<i>C.perfringes</i>	1	1 (100)	1 (100)	1 (100)	0	0	0	(100)
<i>Peptstreptococcus spp</i>	2	1 (50)	1 (50)	2 (100)	1 (50)	1 (50)	2 (100)	(100)
Total (%)	11	(84.6) 94	(62.1) 69	(58.5) 65	(38.7) 43	(42.3) 47	(46.8) 52	(83.6)

Type of Bacteria	No of Isolates	Antimicrobial agent	Approximate MIC		Correlation	Range of MIC (µg/ml)	Susceptibility %
			Amoxiclav	Ciprofloxacin			
			I (µg/ml)	R			
<i>P. mirabilis</i>	(0)	Amoxiclav	≤4(1)	16(1)	16(1)	≥32(3)	100(1)
		Cefotriaxone	<1(3)	16(1)	16(1)	>32(1)	100(1)
<i>E. coli</i>	(36)	Amoxiclav	0.5-16(18)	16(1)	16(3)	0.5-64	88.8
		Cefotriaxone	4(32)	16-	>32(14)	1-32	11.1
		Cefotaxime	≤4(4)	32(18)	>32(14)	0.5-32	11.1
		Amikacin	≤4(4)	≥16(18)	>32(11)	1-64	0
		Ciprofloxacin	1-16(18)	>16(7)	≥4(16)	0.5-8	41.6
<i>K. pneumoniae</i>	(18)	Amoxiclav	≤2(0)	≥8(4)	>32(9)	1-64	27.7
		Cefotriaxone	≤8(9)	≥32(4)	≥64(0)	4-128	0
		Cefotaxime	8(10)	≥16(3)	≥64(0)	8-128	02.0
		Amikacin	≤16(8)	32(3)	≥64(7)	8-128	44.4
		Ciprofloxacin	1-8(10)	≥2(2)	≥4(6)	1-8	00.0
<i>P. aeruginosa</i>	(11)	Amoxiclav	4(1)	16(1)	≥32(9)	1-64	20
		Cefotriaxone	≤8(2)	16(1)	64-	4-128	60
		Cefotaxime	≤8(2)	16(1)	128(8)	4-128	60
		Amikacin	<16(2)	32(1)	≥64(8)	2-64	20
		Ciprofloxacin	<1(8)	2(1)	8>(8)	0.5-8	80
<i>C. freundii</i>	(7)	Amoxiclav	<4(3)	≥8(1)	≥32(3)	0.25-16	42.8
		Cefotriaxone	≤8(2)	16(1)	≥64(4)	128	28.5
		Cefotaxime	≤8(3)	————	≥64(4)	0.004-16	42.8
		Amikacin	≤16(1)	32(1)	≥64(0)	128	14.2
		Ciprofloxacin	1(2)	2(1)	≥4(4)	0.004-16	28.5

Table (3 - 10) Antimicrobial susceptibility of bacteria isolated from patients with acute appendicitis, MICs of five antibiotics to wards isolated bacteria

<i>S. typhi</i>	(๑)	Amoxiclave	<๙(๑)	๙(๑)	≥๑๖(๓)	
		Cefotriaxone	≤๙(๓)	๑๖(๑)	๓๓(๑)	
		Cefotaxime	————	๓๓(๑)	≥๓๕(๕)	
		Amikacin	————	————	≥๓๓(๑)	
		Ciprofloxacin	<๑(๕)	๓(๑)	>๓(๑)	
<i>E. aerogenes</i>	(๕)	Amoxiclave	≤๕(๓)	๙(๑)	————	
		Cefotriaxone	≤๕(๕)	————	————	
		Cefotaxime	≤๕(๕)	————	————	
		Amikacin	๙(๓)	————	>๓๓(๓)	
		Ciprofloxacin	<๑(๕)	————	————	
<ul style="list-style-type: none"> • S= Sensitive • I= Intermediate • R= Resistance 						