

## Phytochemical Screening of Methanolic Leaves Extract of *Malva sylvestris*

Abeer Fauzi Al-Rubaye<sup>1</sup>, Ashwak Falih Kaizal<sup>1</sup>, Imad Hadi Hameed<sup>2\*</sup>

<sup>1</sup>Department of Biology, College of Science for women, University of Babylon, Iraq

<sup>2</sup>College of Nursing, University of Babylon, Iraq

Received: 18<sup>th</sup> March, 17; Revised 26<sup>th</sup> March, 17, Accepted: 15<sup>th</sup> April, 17; Available Online: 25<sup>th</sup> April, 2017

### ABSTRACT

The objectives of this study were analysis of the secondary metabolite products and evaluation antibacterial activity. Bioactives are chemical compounds often referred to as secondary metabolites. Thirty six bioactive compounds were identified in the methanolic extract of *Malva sylvestris*. The identification of bioactive chemical compounds is based on the peak area, retention time molecular weight and molecular formula. GC-MS analysis of *Malva sylvestris* revealed the existence of the 1-Propanamine, 2-methyl-N-(2-methylpropylidene)-, Pyrrolidine, 1-(1-oxo-2,5-octadecadienyl)-, Dimethyl sulfoxide, Cyclohexylamine, N-ethyl-, N-(2-Methylbutylidene)isobutylamine, 1-Methyl-2-pyrrolidethanol, 2-(2-Hydroxyethyl)piperidine, 1-Butanamine, 2-methyl-N-(2-methylbutylidene)-, 4-(Pyrrolidin-2-ylcarbonyl)morpholine, Dithiocarbamate, S-methyl-, N-(2-methyl-3-oxobutyl)-, 1-Gala-1-ido-octonic lactone, 1-(5'-methylfurfuryl)pyrrolidine, 2-Methoxy-4-vinylphenol, Pyrrolizin-1,7-dione-6-carboxylic acid, methyl(ester), 1-Naphthaienol, 1,2,3,4-tetrahydro-2,5,8-trimethyl-, Pterin-6-carboxylic acid, N-(2-Acetamido)iminodiacetic acid, N-(1-Hydroxy-4-oxo-1-phenylperhydroquinolizin-3-yl)carbamic, Cyclopropanedodecanoic acid, 2-octyl-, methyl ester, Cholestan-3-ol, 2-methylene-, (3 $\beta$ ,5 $\alpha$ )-, 3-(N,N-Dimethyl laurylammonio)propanesulfonate, Pyrazole[4,5-b]imidazole, 1-formyl-3-ethyl-6- $\beta$ -d-ribofuranosyl-, Octahydrobenzo[b]pyran, 4a-acetoxy-5,5,8a-trimethyl-, Tetraacetyl-d-xylonic nitrile, 4,6-Heptadien-3-one, 1,7-diphenyl-, Pentanoic acid, 2,2,4-trimethyl-3-carboxyisopropyl, isobutyl ester, D-Fructose, diethyl mercaptal, pentaacetate, Phytol, Hexadecanamide, Tributyl acetyl citrate, Cholestan-3-one, cyclic 1,2-ethanediyl acetal, (5 $\beta$ )-, Dasycarpidan-1-methanol, acetate (ester)-, 9-Desoxo-9-x-acetoxy-3,8,12-tri-O-acetylingol, (+)-y-Tocopherol, O-methyl-, Campesterol and Stigmasterol.

**Keyword:** *Malva sylvestris*, Gas chromatography, Phytochemical, Malvaceae.

### INTRODUCTION

*Malva sylvestris* L. (Malvaceae family), annual or biennial herbaceous medicinal plant usually known as common mallow, is native to Asia, North Africa and Europe<sup>1</sup>. The high mucilage content of *Malva sylvestris* makes it an excellent demulcent that can be used for many applications. In the digestive tract the fruit mucilage can be used to heal and soothe inflammations such as gastritis, peptic ulcers, enteritis, and colitis<sup>2</sup>. *Malva sylvestris* L. has been used in folk medicine of Brazil and other countries for the treatment of colitis and stomatitis, in cases of chronic bronchitis, against furuncle and abscess, contusions and haemorrhoids as well as other dolorous and inflammatory processes<sup>3-7</sup>.

Young leaves considered one of the culinary herbs in Palestine and in other Mediterranean countries, they are eaten raw in salads or consumed in soups and as boiled vegetables<sup>8-13</sup>. A pharmaceutical suspension, is thermodynamically unstable, thus, making it necessary to include in the dosage form, a stabilizer or suspending agent which reduces the rate of settling and permits easy redispersion of any settled particulate matter both by protective colloidal action and by increasing the

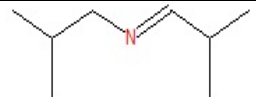
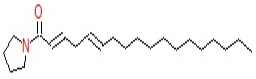
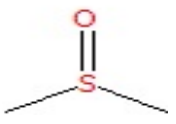
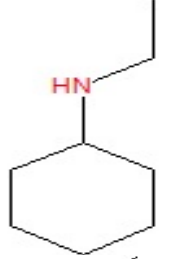
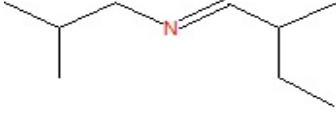
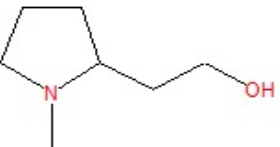
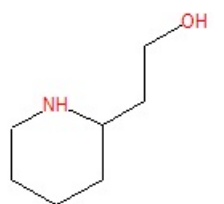
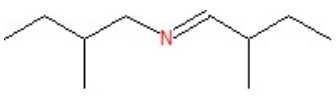
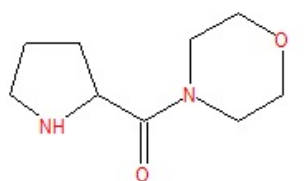
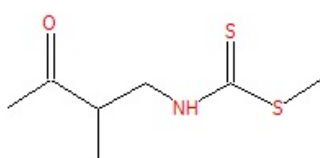
consistency of the suspending medium. It is also widely recognized to have anti-inflammatory properties, some other pharmacological and clinical effects are frequently mentioned such as diuretic, laxative, antiseptic, antispasmodic, lenitive, choleric, bronchodilator, expectorant, antitussive and antiacne activities<sup>14-18</sup>.

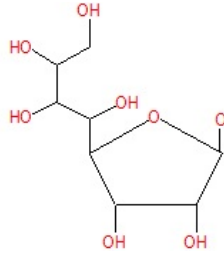
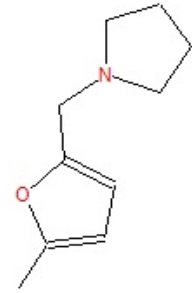
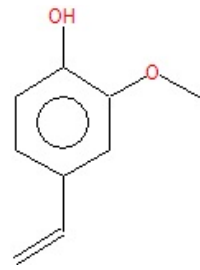
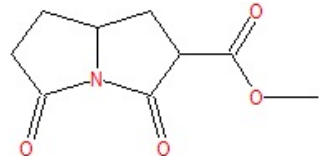
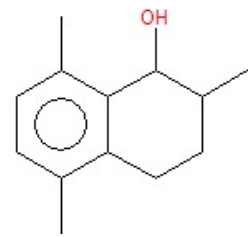
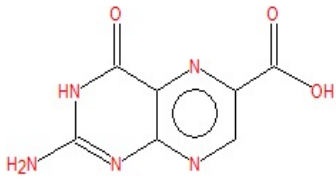
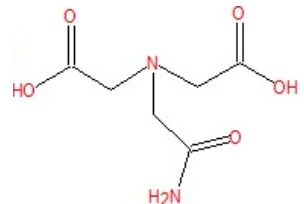
*Malva sylvestris* is an herbaceous plant used in phitotherapy and widely distributed in Terpenoids, phenolic acids and anthocyanins were identified in water leaf extract of the plant. It is used for many purposes as medicine, nutrition, fibrous, green color dye, and cosmetic from centuries. Numbers of medical and pharmacologic researches about nettle are increased day by day. Nettle leaves contain anthocyanin glycosides, quercetin, rutin flavonoids, chlorophyll a, chlorophyll b,  $\beta$ -carotene, and lutein<sup>19, 20</sup>.

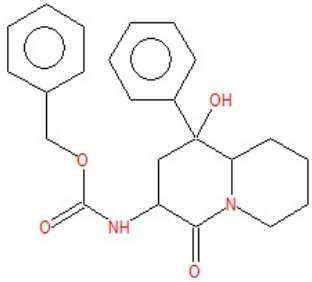
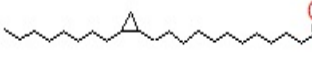
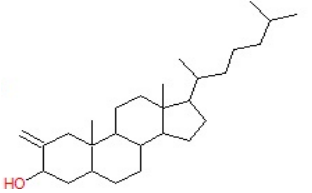
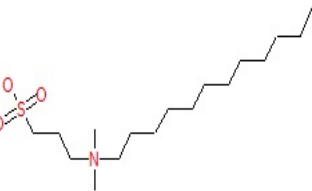
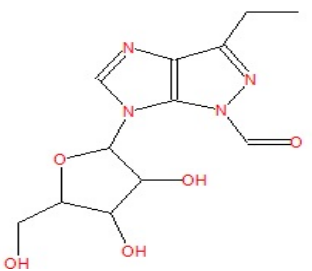
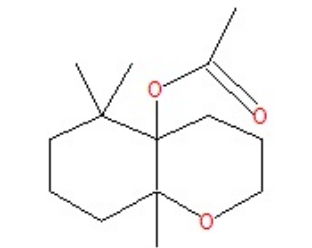
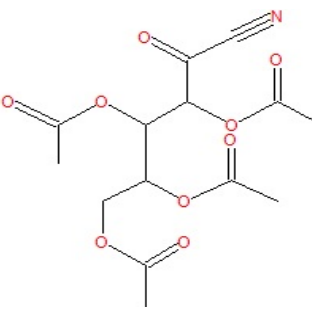
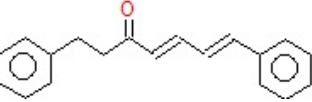
### MATERIALS AND METHODS

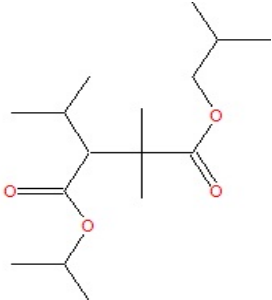
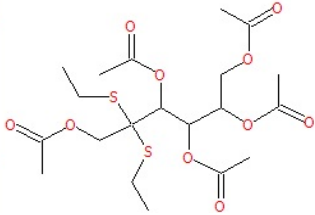
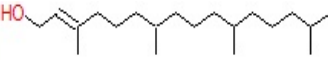
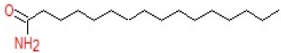
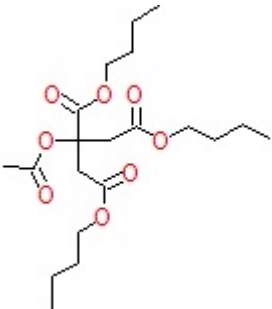
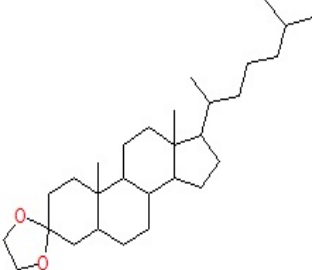
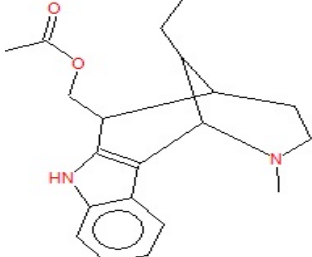
*Malva sylvestris* were collected from local market in Hilla city, middle of Iraq. After thorough cleaning and removal of foreign materials. About eighteen grams of methanolic extract of *Malva sylvestris* powdered were soaked in one hundred mL methanol for ten hours in a

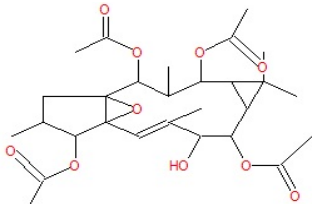
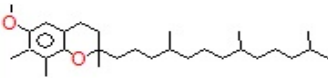
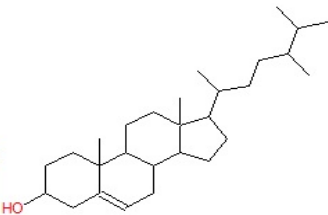
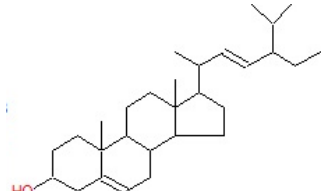
Table 1: Major phytochemical compounds identified in methanolic extract of *Malva sylvestris*.

Serial No.	Phytochemical compound	RT (min)	Molecular Weight	Exact Mass	Chemical structure	MS Fragmentations	Pharmacological actions
1.	1-Propanamine, 2-methyl-N-(2-methylpropylidene)-	3.218	127	127.1360993		57,84,112	Unknown
2.	Pyrrolidine,1-(1-oxo-2,5-octadecadienyl)-	3.396	333	333.303165		55,81,98,13,150,220,264,333	Unknown
3.	Dimethyl sulfoxide	3.476	78	78.013936		63,78	anti-inflammatory, and an antioxidant
4.	Cyclohexylamine, N-ethyl-	3.590	127	127.1360993		55,71,84,98,127	anti-inflammatory and antioxidant
5.	N-(2-Methylbutylidene) isobutylamine	3.877	141	141.15175		57,69,98,13,126	anti-stereochemistry
6.	1-Methyl-2-pyrrolidideethanol	4.306	129	129.115364		55,84,98,129	Unknown
7.	2-(2-Hydroxyethyl) piperidine	4.449	129	129.115364		56,84,98,128	Antimicrobial, Anti-malarial, Antibacterial
8.	1-Butanamine, 2-methyl-N-(2-methylbutylidene)-	4.563	155	155.167399		56,70,84,98,113,127,140,154	antimicrobial activity
9.	4-(Pyrrolidin-2-ylcarbonyl)morpholine	4.649	184	184.121178		56,70,86,114,142	antimicrobial activity
10.	Dithiocarbamate, S-methyl-, N-(2-methyl-3-oxobutyl)-	5.215	191	191.043856		57,85,143,191	anti-bacterial activity

11.	l-Gala-l-ido-octonic lactone	6.057	238	238.068868		61,73,84,1 12,127,142 ,159,189,2 20	anti-bacterial activity
12	1-(5'-methylfurfuryl)pyrrolidine	6.371	165	165.115364		95,122,165	Anti-oxidant
13	2-Methoxy-4-vinylphenol	7.041	150	150.06808		51,77,89,1 07,135	Antioxidant, anti microbial and anti-inflammatory
14	Pyrrolizin-1,7-dione-6-carboxylic acid , methyl(ester)	7.361	197	197.068808		55,69,84,9 8,142,197	anti-tumor activity, anti-diabetic activity
15	1-Naphthaienol , 1,2,3,4-tetrahydro-2,5,8-trimethyl-	7.853	190	190.135765		51,77,11,1 42,157,172 ,190	Unknown
16	Pterin-6-carboxylic acid	7.916	207	207.039239		57,69,105, 149,163,20 7	Anti-cancer, anti-viral
17	N-(2-Acetamido)imino diacetic acid	8.025	190	190.058971		71,101,127 ,146,172,1 90	Unknown

18	N-(1-Hydroxy-4-oxo-1-phenylperhydroquinolin-3-yl)carbamic	8.248	394	394.189257		55,84,105,138,166,196,238,282,394	anti-tumor activity, anti-diabetic activity
19	Cyclopropanedodecanoic acid, 2-octyl-, methyl ester	8.563	366	366.349781		5,69,118,66,334,366	antioxidants, anti-microbial
20	Cholestan-3-ol,2-methylene-, (3β,5α)-	8.626	400	400.370516		69,81,95,149,227,315,400	anti-inflammatory
21	3-(N,N-Dimethylaurylammonio)propanesulfonate	8.906	335	335.249414		58,69,84,97,122,152,179,213	activity anti-oxidant and anti-inflammatory activities
22	Pyrazole[4,5-b]imidazole, 1-formyl-3-ethyl-6-β-d-ribofuranosyl-	9.101	296	296.11207		55,149,281	antimicrobial, anticancer
23	Octahydrobenzo[b]pyran, 4a-acetoxy-5,5,8a-trimethyl-	9.427	240	240.1725445		55,69,111,180,197,240	anti-inflammatory
24	Tetraacetyl-d-xylonic nitrile	9.604	343	343.090332		60,73,112,133,238,281	antioxidant and anti-inflammatory
25	4,6-Heptadien-3-one,1,7-diphenyl-	9.856	262	262.135765		51,77,91,128,157,262	anti-oxidative of compounds

26	Pentanoic acid, 2,2,4-trimethyl-3-carboxyisopropyl, isobutyl ester	9.948	286	286.214409		55,71,97,1 59,243	Unknown
27	D-Fructose, diethyl mercaptal, pentaacetate	11.693	496	496.14369		60,97,129, 154,273,31 6,375,436	anti- bacterial activity
28	Phytol	15.074	296	296.307917		57,71,81,9 5,111,123, 137,196,22 1,249,278	Antinociceptive and Antioxidant Activities
29	Hexadecanamide	15.961	255	255.256215		59,72,86,1 28,170,212 ,255	analgesic and anti-inflammatory
30	Tributyl acetylcitrate	16.413	402	402.225368		57,112,129 ,157,185,2 13,231,259 ,273,329	anti-bacterial
31	Cholestan-3-one, cyclic 1,2-ethanediyl aetal, (5β)-	18.925	430	430.38108		55,69,99,1 25,149,194 ,232,282,3 40,384,430	analgesic, anti-ulcer, anticancer
32	Dasycarpidan-1-methanol, acetate (ester)-	19.990	326	326.199429		69,97,180, 222,256,32 6	inflammatory, anti-bacterial, anti-fungal, anti-diabetic, anti-cancer

33	9-Desoxo-9-x-acetoxy-3,8,12-tri-O-acetylgingol	22.296	536	536.262146		55,69,122, 207,236,29 7,357,417, 477,536	anti-bacterial activity
34	(+)- $\gamma$ -Tocopherol, O-methyl-	22.828	430	430.38108		57,69,137, 165,205,27 4,316,358, 430	anti-oxidant activity
35	Campesterol	23.531	400	400.370516		55,81,145, 213,255,28 9,315,382, 400	anti- inflammatory effects
36	Stigmasterol	23.737	412	412.370516		55,83,133, 213,255,30 0,351,369, 412	anti- microbial

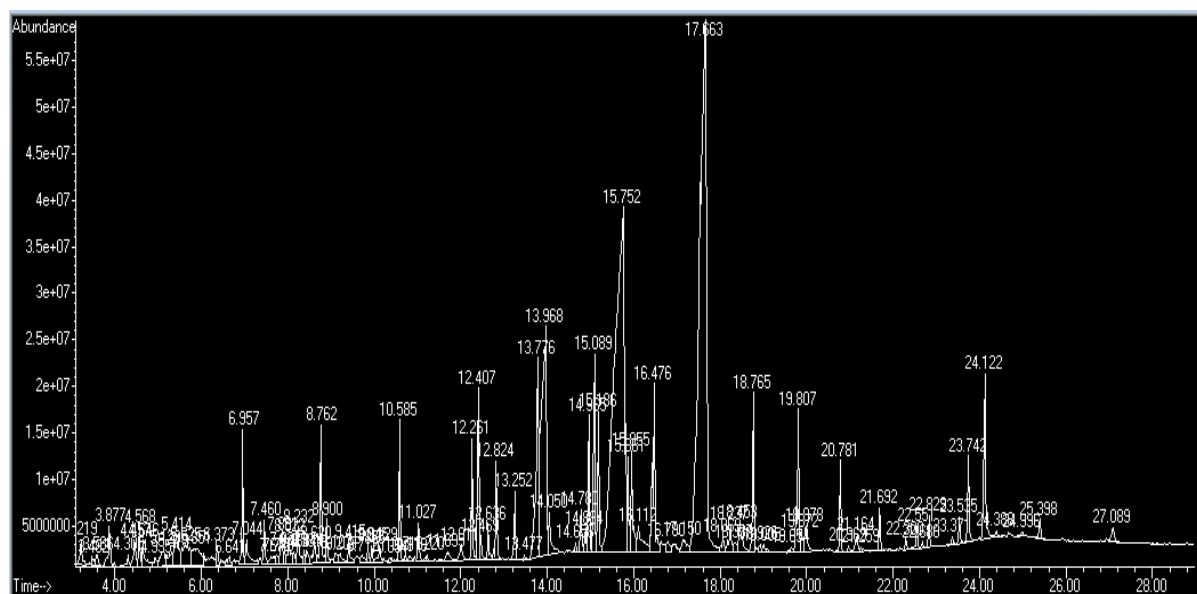


Figure 1: GC-MS chromatogram of methanolic extract of *Malva sylvestris*.

rotatory shaker. The filtrates were used for further phytochemical analysis<sup>21-30</sup>. It was again filtered through sodium sulphate in order to remove the traces of moisture.

#### Gas chromatography – mass spectrum analysis

GC-MS is a powerful technique used for many applications which has very high sensitivity and specificity. One  $\mu\text{L}$  of the methanol extract of *Malva sylvestris* was injected into the GC-MS using a micro syringe and the scanning was done for 45 minutes. The time from when the injection was made (Initial time) to

when elution occurred referred to as the Retention time (RT). While the instrument was run, the computer generated a graph from the signal called Chromatogram. Each of the peaks in the chromatogram represented the signal created when a compound eluted from the gas chromatography column into the detector. As individual compounds eluted from the gas chromatographic column, they entered the electron ionization (mass spectroscopy) detector, where they were bombarded with a stream of electrons causing them to break apart into fragments. The fragments obtained were actually charged ions with a

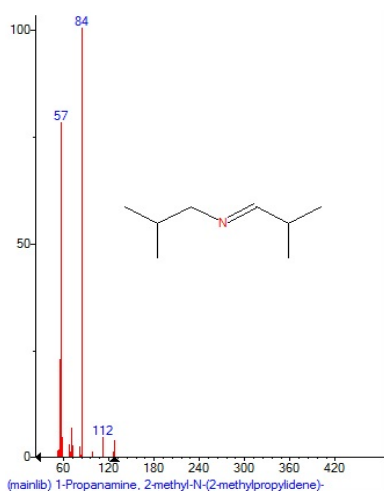


Figure 2: Mass spectrum of 1-Propanamine, 2-methyl-N-(2-methylpropylidene)- with Retention Time (RT)= 3.218.

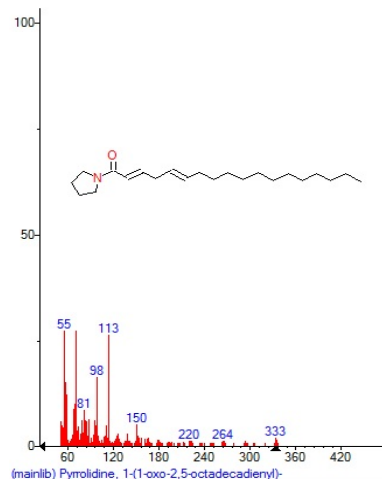


Figure 3: Mass spectrum of Pyrrolidine, 1-(1-oxo-2,5-octadecadienyl)- with Retention Time (RT)= 3.396.

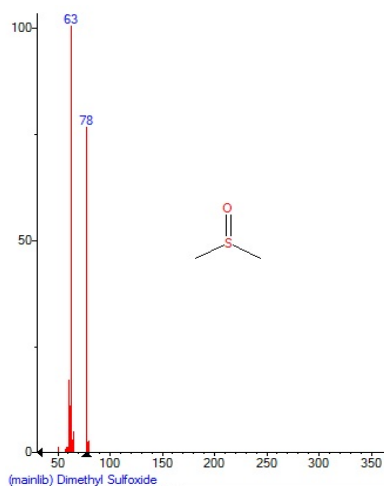


Figure 4: Mass spectrum of Dimethyl sulfoxide with Retention Time (RT)= 3.476.

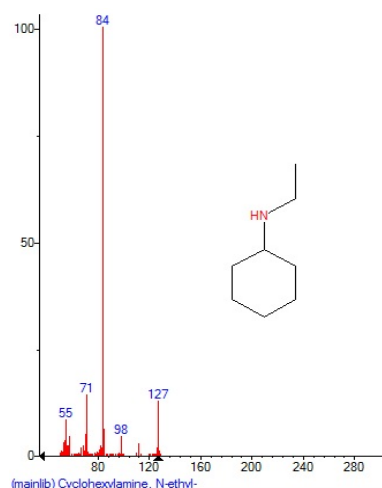


Figure 5: Mass spectrum of Cyclohexylamine, N-ethyl- with Retention Time (RT)= 3.590.

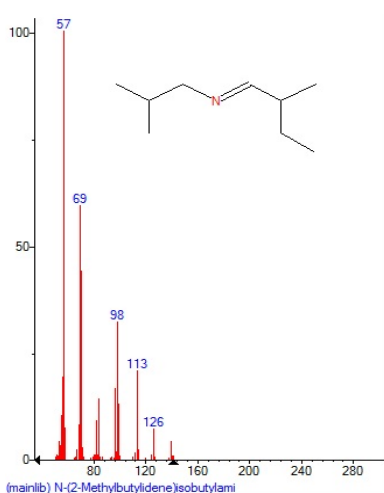


Figure 6: Mass spectrum of N-(2-Methylbutylidene)isobutylamine with Retention Time (RT)= 3.877.

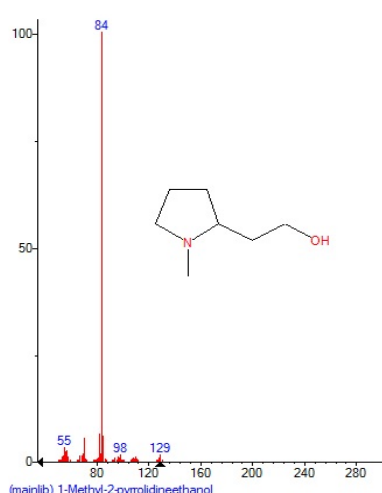


Figure 7: Mass spectrum of 1-Methyl-2-pyrrolidineethanol with Retention Time (RT)= 4.306.

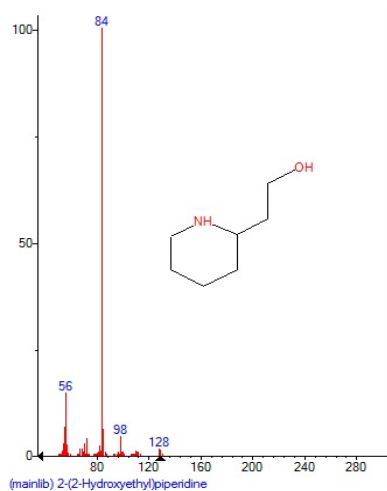


Figure 8: Mass spectrum of 2-(2-Hydroxyethyl)piperidine with Retention Time (RT)= 4.449.

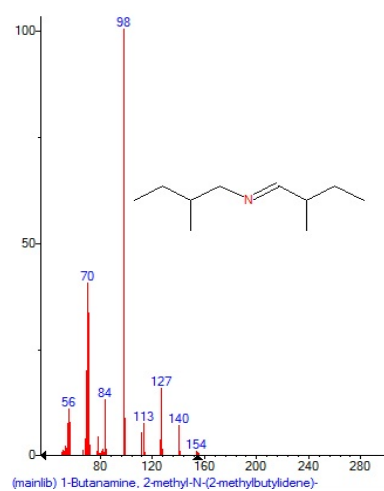


Figure 9: Mass spectrum of 1-Butanamine, 2-methyl-N-(2-methylbutylidene)-with Retention Time (RT)= 4.563.

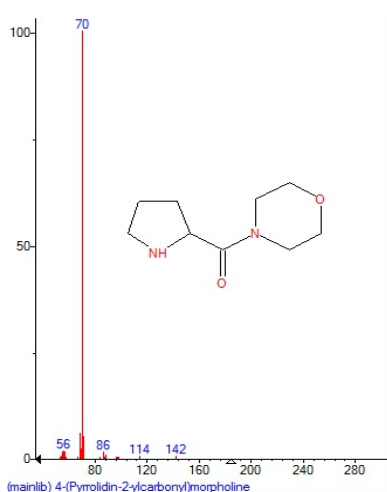


Figure 10: Mass spectrum of 4-(Pyrrolidin-2-ylcarbonyl)morpholine with Retention Time (RT)= 4.649.

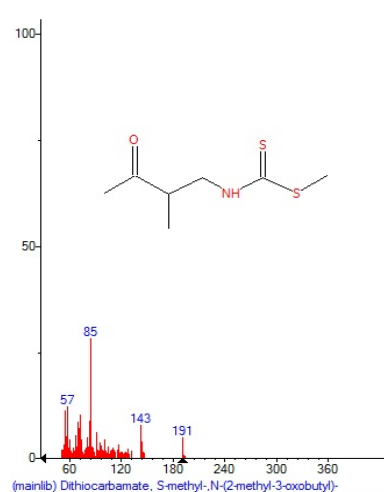


Figure 11: Mass spectrum of Dithiocarbamate, S-methyl-, N-(2-methyl-3-oxobutyl)- with Retention Time (RT)= 5.215.

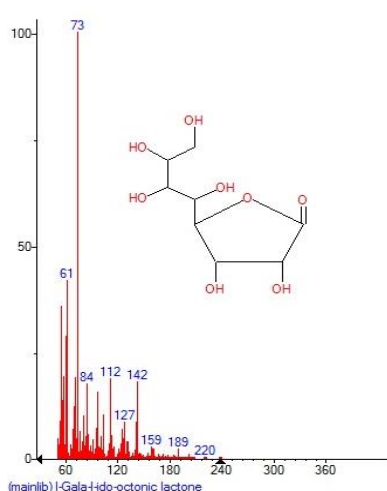


Figure 12: Mass spectrum of 1-Gala-1-ido-octonic lactone with Retention Time (RT)= 6.057.

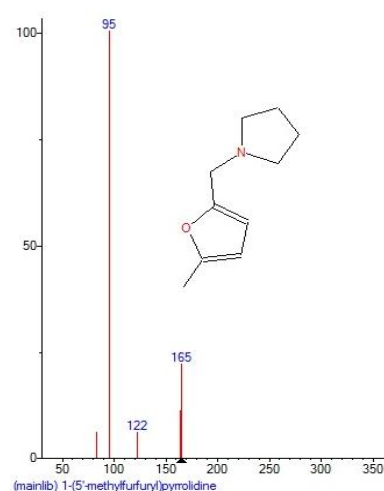


Figure 13: Mass spectrum of 1-(5'-methylfurfuryl)pyrrolidine with Retention Time (RT)= 6.371.



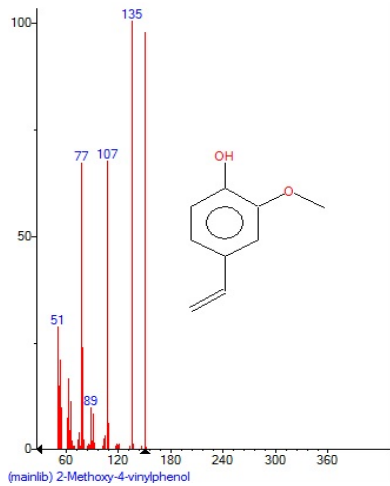


Figure 14: Mass spectrum of 2-Methoxy-4-vinylphenol with Retention Time (RT)= 7.041.

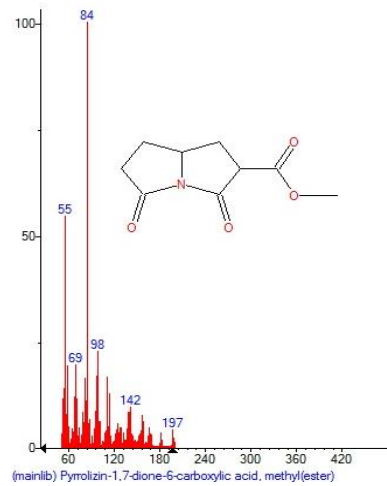


Figure 15: Mass spectrum of Pyrrolizin-1,7-dione-6-carboxylic acid methyl(ester) with Retention Time (RT)= 7.361.

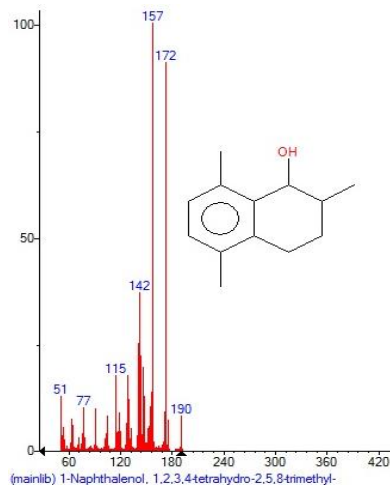


Figure 16: Mass spectrum of 1-Naphthalenol, 1,2,3,4-tetrahydro-2,5,8-trimethyl- with Retention Time (RT)= 7.853.

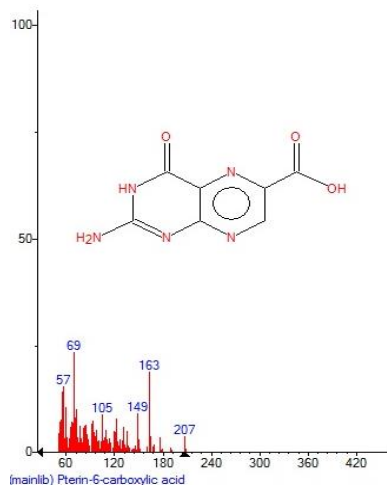


Figure 17: Mass spectrum of Pterin-6-carboxylic acid with Retention Time (RT)= 7.916.

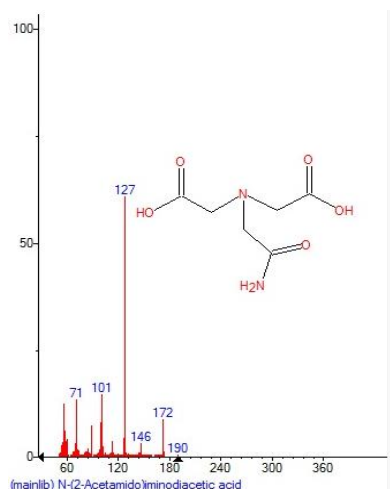


Figure 18: Mass spectrum of N-(2-Acetamido)iminodiacetic acid with Retention Time (RT)= 8.025.

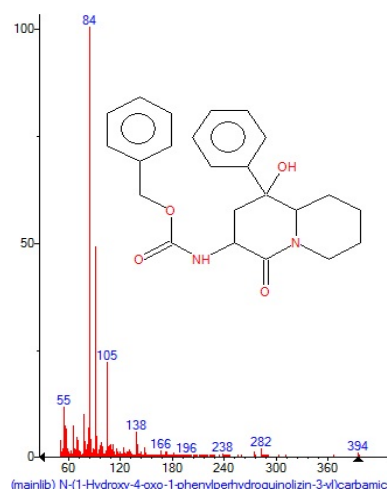


Figure 19: Mass spectrum of N-(1-Hydroxy-4-oxo-1-phenylperhydroquinolizin-3-yl)carbamic with Retention Time (RT)= 8.248.

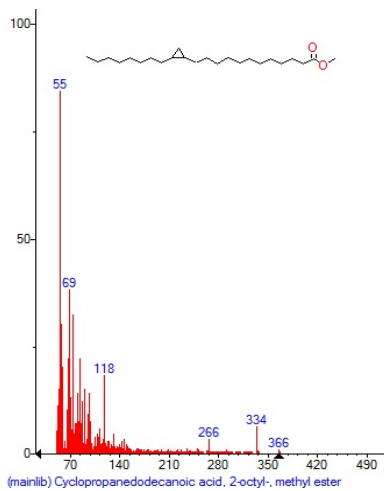


Figure 20: Mass spectrum of Cyclopropanedodecanoic acid , 2-octyl-,methyl ester with Retention Time (RT)= 8.563.

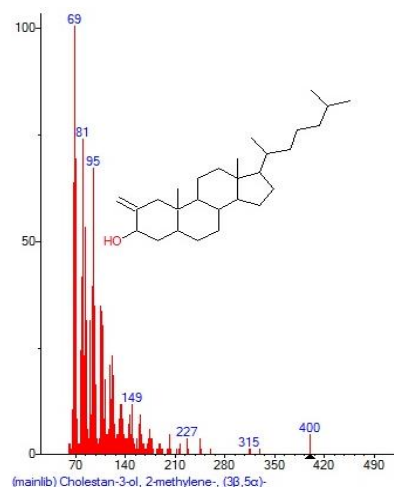


Figure 21: Mass spectrum of Cholestan-3-ol,2-methylene-, (3β,5α)-with Retention Time (RT)= 8.626.

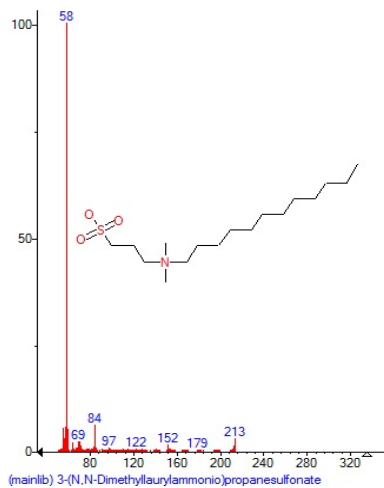


Figure 22: Mass spectrum of 3-(N,N-Dimethyl-laurylammonio)propanesulfonate with Retention Time (RT)= 8.906.

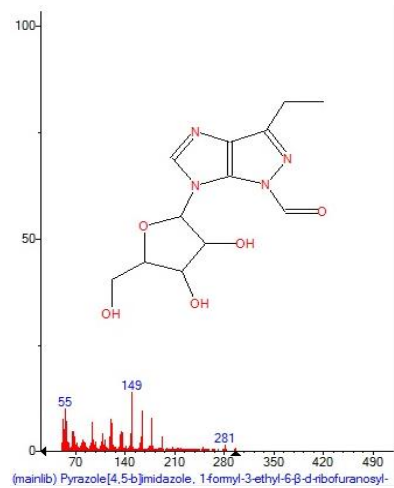


Figure 23: Mass spectrum of Pyrazole[4,5-b]imidazole , 1-formyl-3-ethyl-6-β-d-ribofuranosyl- with Retention Time (RT)= 9.101.

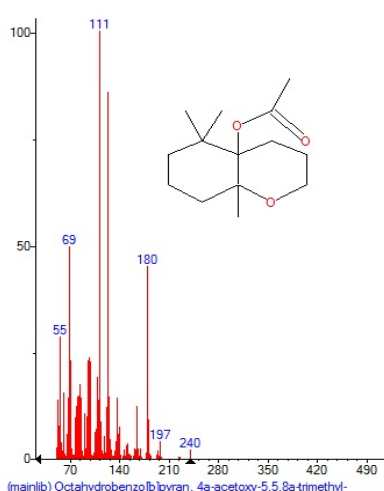


Figure 24: Mass spectrum of Octahydrobenzo[b]pyran , 4a-acetoxy-5,5,8a-trimethyl- with Retention Time (RT)= 9.427.

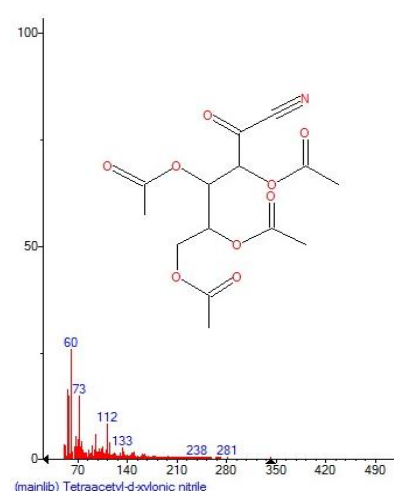


Figure 25: Mass spectrum of Tetraacetyl-d-xylonic nitrile with Retention Time (RT)= 9.604.

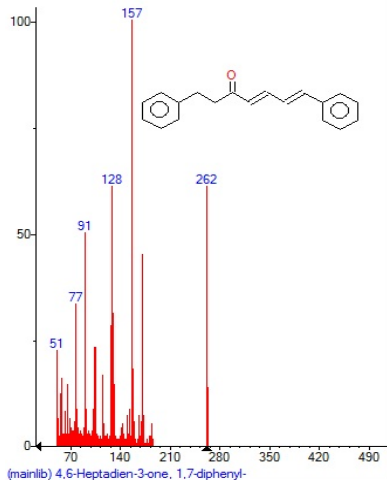


Figure 26: Mass spectrum of 4,6-Heptadien-3-one,1,7-diphenyl- with Retention Time (RT)= 9.856.

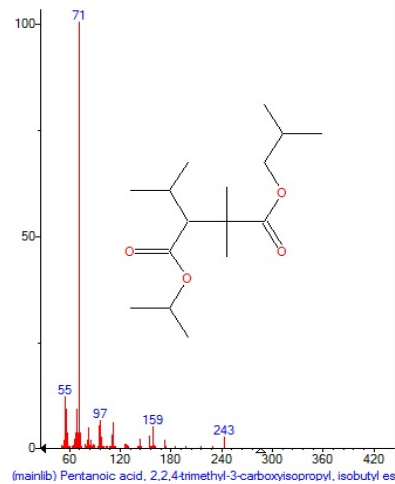


Figure 27: Mass spectrum of Pentanoic acid ,2,2,4-trimethyl-3-carboxyisopropyl, isobutyl ester with Retention Time (RT)= 9.948.

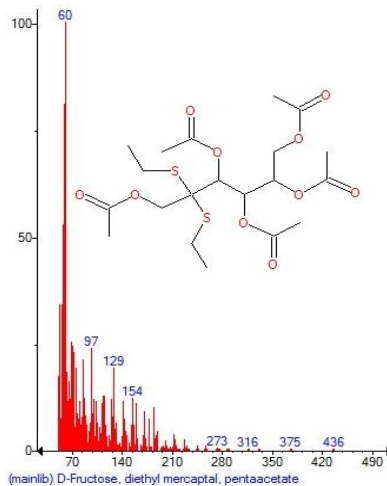


Figure 28: Mass spectrum of D-Fructose, diethyl mercaptal, pentaacetate with Retention Time (RT)= 11.693.

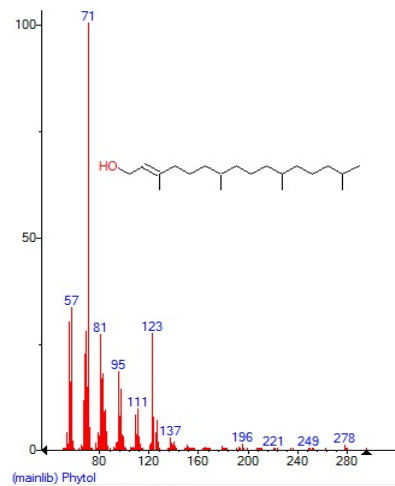


Figure 29: Mass spectrum of Phytol with Retention Time (RT)= 15.074.

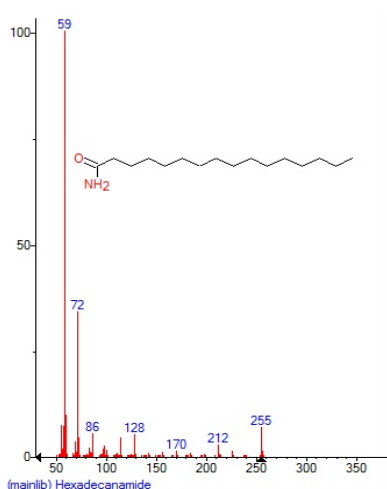


Figure 30: Mass spectrum of Hexadecanamide with Retention Time (RT)= 15.961.

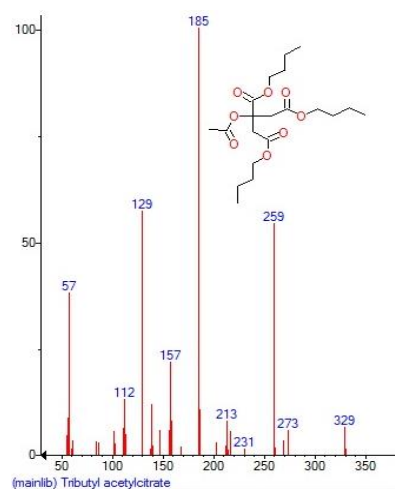


Figure 31: Mass spectrum of Tributyl acetylacrylate with Retention Time (RT)= 16.413.

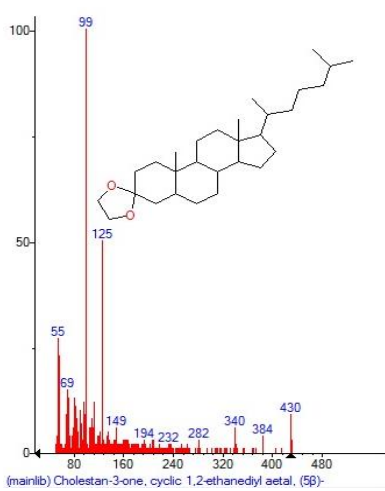


Figure 32: Mass spectrum of Cholestan-3-one, cyclic 1,2-ethanediyl aetal, (5 $\beta$ )- with Retention Time (RT)= 18.925.

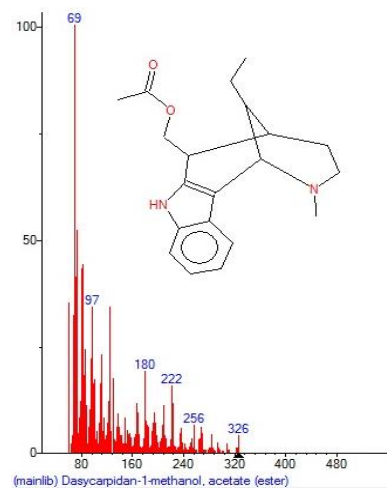


Figure 33: Mass spectrum of Dasycarpidan-1-methanol, acetate (ester)- with Retention Time (RT)= 19.990.

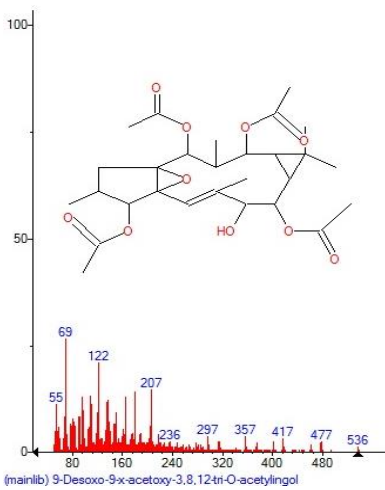


Figure 34: Mass spectrum of 9-Desoxo-9-x-acetoxy-3,8,12-tri-O-acetylgingol with Retention Time (RT)= 22.296.

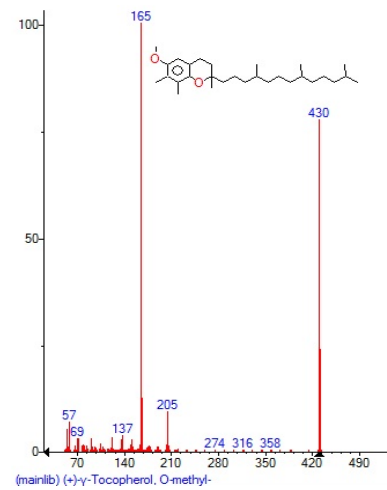


Figure 35: Mass spectrum of (+)- $\gamma$ -Tocopherol, O-methyl- with Retention Time (RT)= 22.828.

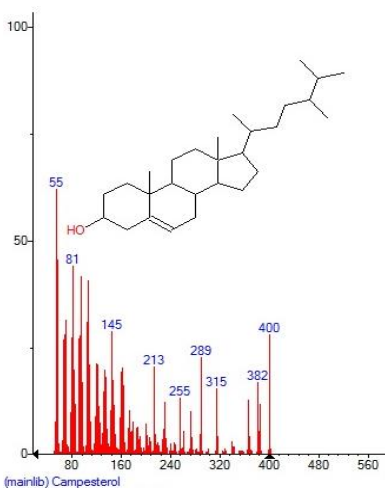


Figure 36: Mass spectrum of Campesterol with Retention Time (RT)= 23.531.

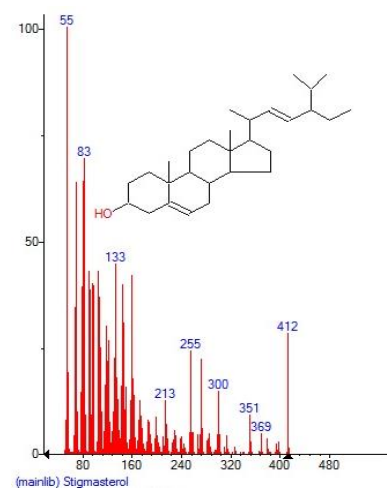


Figure 37: Mass spectrum of Stigmasterol with Retention Time (RT)= 23.737.

certain mass. Helium gas was used as a carrier as well as an eluent<sup>31-51</sup>. The flow rate of helium was set to 1ml per

minute. The electron gun of mass detector liberated electrons having energy of about 70eV. The identity of the components in the extracts was assigned by the

comparison of their retention indices and mass spectra fragmentation patterns with those stored on the computer library and also with published literatures. Compounds were identified by comparing their spectra to those of the Wiley and NIST/EPA/NIH mass spectral libraries<sup>52-61</sup>.

## RESULTS AND DISCUSSION

Gas chromatography and mass spectroscopy analysis of compounds was carried out in methanolic leaves extract of *Malva sylvestris*, shown in Table 1. The GC-MS chromatogram of the 31 peaks of the compounds detected was shown in Figure 1. Chromatogram GC-MS analysis of the methanol extract of *Malva sylvestris* showed the presence of thirty one major peaks and the components corresponding to the peaks were determined as follows. The First set up peak were determined to be Propanamine, 2-methyl-N-(2-methylpropylidene)-, Pyrrolidine,1-(1-oxo-2,5-octadecadienyl)-, Dimethyl sulfoxide, Cyclohexylamine, N-ethyl-, N-(2-Methylbutylidene)isobutylamine, 1-Methyl-2-pyrrolidideethanol, 2-(2-Hydroxyethyl)piperidine, 1-Butanamine, 2-methyl-N-(2-methylbutylidene)-, 4-(Pyrrolidin-2-ylcarbonyl)morpholine, Dithiocarbamate, S-methyl-,N-(2-methyl-3-oxobutyl)-, 1-Gala-1-ido-octonic lactone, 1-(5'-methylfurfuryl)pyrrolidine, 2-Methoxy-4-vinylphenol, Pyrrolizin-1,7-dione-6-carboxylic acid, methyl(ester), 1-Naphthaienol, 1,2,3,4-tetrahydro-2,5,8-trimethyl-, Pterin-6-carboxylic acid, N-(2-Acetamido)iminodiacetic acid, N-(1-Hydroxy-4-oxo-1-phenylperhydroquinolizin-3-yl)carbamic, Cyclopropanedodecanoic acid, 2-octyl-,methyl ester, Cholestan-3-ol,2-methylene-,(3 $\beta$ ,5 $\alpha$ )-, 3-(N,N-Dimethyl-laurylammonio)propanesulfonate, Pyrazole[4,5-b]imidazole, 1-formyl-3-ethyl-6- $\beta$ -d-ribofuranosyl-, Octahydrobenzo[b]pyran, 4a-acetoxy-5,5,8a-trimethyl-, Tetraacetyl-d-xylonic nitrile, 4,6-Heptadien-3-one,1,7-diphenyl-, Pentanoic acid, 2,2,4-trimethyl-3-carboxyisopropyl, isobutyl ester, D-Fructose, diethyl mercaptal, pentaacetate, Phytol, Hexadecanamide, Tributyl acetyl-citrate, Cholestan-3-one,cyclic 1,2-ethanediy l aetal, (5 $\beta$ )-, Dasycarpidan-1-methanol, acetate (ester)-, 9-Desoxo-9-x-acetoxy-3,8,12-tri-O-acetylingol, (+)-y-Tocopherol, O-methyl-, Campesterol and Stigmasterol (Figure 2-37). From ancient times, plants have been used for many purposes, including food, medicine, flavoring agents, cosmetics and other uses<sup>62-68</sup>. Antioxidant and antiprotozoic activities; quercetin is the major flavonoid in the human diet and has been reported to exhibit antioxidative. *M. sylvestris* extracts are reported for their radical scavenging effect. *Malva sylvestris* extracts are reported for their radical scavenging effect as well as *E. camaldulensis* and *C. sativa*; the later demonstrated also antineoplastic activity in B16 cells<sup>69-71</sup>.

## CONCLUSION

The results clearly shows that the plants which have the highest organic and aqueous extraction yields have the highest antioxidant activities. Wild natural *Malva sylvestris* plants leaves had slightly higher antioxidant activity than their cultivated species. Based on the above

presented results the rareness of natural antioxidant use which is usually due to the cost and unavailability of the wild plant could be overcome by using cultivated species. The cultivated *Malva sylvestris* plants leaves, could be used as a possible new source of natural antioxidants in the food, nutraceuticals, pharmaceuticals and cosmetic industry.

## REFERENCES

1. Ali-Shtayeh MS, Jamous RM, Al-Shafie JH, Elgharabah WA, Kherfan FA, Qarariah KH, Isra'S K, Soos IM, Musleh AA, Isa BA. Traditional knowledge of wild edible plants used in Palestine (Northern West Bank): a comparative study. Journal of Ethnobiology and Ethnomedicine. 2008; 4: 13-22.
2. Ali-Shtayeh MS, Yaniv Z, Mahajna J. Ethnobotanical survey in the Palestinian area: a classification of the healing potential of medicinal plants. Journal of Ethnopharmacology. 2000;73: 221-232.
3. Aruna C, Chaithra D, Alekhya C, Yasodamma N. Pharmacognostic studies of *Aeschynomene indica* L. International Journal of Pharmacy and Pharmaceutical Sciences. 2012; 4:76-77.
4. Barros L, Carvalho AM, Ferreira IC. Leaves, flowers, immature fruits and leafy flowered stems of *Malva sylvestris*: A comparative study of the nutraceutical potential and composition. Food and Chemical Toxicology. 2010; 48:1466-1472.
5. Camejo-Rodrigues J, Ascensão L, Bonet M, Valles J. An ethnobotanical study of medicinal and aromatic plants in the Natural Park of "Serra de São Mamede"(Portugal). Journal of Ethnopharmacology. 2003 ;89:199-209.
6. Cragg GM, Newman DJ, Weiss RB. Coral reefs, forests, and thermal vents: the worldwide exploration of nature for novel antitumor agents, Seminars in oncology. 1997;156-163.
7. Eberhardt MV, Lee CY, Liu RH. Nutrition: Antioxidant activity of fresh apples. Nature. 2000; 405: 903-904.
8. Erdemoglu N, Kupeli E, Yesilada E. Anti-inflammatory and antinociceptive activity assessment of plants used as remedy in Turkish folk medicine. Journal of Ethnopharmacology. 2003; 89:123-129.
9. Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. Nature. 2000; 408: 239-247.
10. Gasparetto JC, Martins CF, Hayashi SS, Otuky MF, Pontarolo R. Ethnobotanical and scientific aspects of *Malva sylvestris* L.: a millennial herbal medicine. Journal of Pharmacy and Pharmacology. 2012; 64: 172-189.
11. Gey K. The antioxidant hypothesis of cardiovascular disease: epidemiology and mechanisms. Biochemical Society Transactions. 1990;18:1041-1045.
12. Husain SZ, Malik RN, Javaid M, Bibi S. Ethnobotanical properties and uses of medicinal plants of Morgah biodiversity park, Rawalpindi. Pak J Bot. 2008; 40:1897-1911.
13. Ji T, Liu C, Wang A, Yang J, Su Y, Yuan L, Feng X. Studies on the chemical constituents of *Urtica dioica*

- L. grown in Tibet Autonomous Region. *Zhongyaocai Journal of Chinese medicinal materials*. 2007; 30: 662-664.
14. Kavtaradze NS, Alaniya M. Anthocyan Glycosides from *Urtica dioica*. *Chemistry of natural compounds*. 2003; 39: 315-315.
15. Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE, Hilpert KF, Griel AE, Etherton, TD. Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *The American journal of medicine*. 2002; 113: 71-88.
16. Madhavi D, Deshpande S, Salunkhe D. 1996. Food antioxidants: technological, toxicological, health perspective. Marcel Dekker, New York. Mermelstein NH. Determining antioxidant activity. *Food technology*. 2008; 11: 63-66.
17. Oliver-Bever B. Vegetable drugs for cancer therapy. *Pharmaceutical Biology*. 1971; 11:1665-1683.
18. Otles S, Yalcin B. Phenolic compounds analysis of root, stalk, and leaves of nettle. *The Scientific World Journal*. 2012;2012:12-24.
19. Pardo-De-Santayana M, Tardío J, Morales R. The gathering and consumption of wild edible plants in the Campoo (Cantabria, Spain). *International Journal of Food Sciences and Nutrition*. 2005; 56: 529-542.
20. Quave CL, Plano LR, Pantuso T, Bennett BC. Effects of extracts from Italian medicinal plants on planktonic growth, biofilm formation and adherence of methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology*. 2008; 118: 418-428.
21. Kadhim MJ, Sosa AA, Hameed IH. Evaluation of anti-bacterial activity and bioactive chemical analysis of *Ocimum basilicum* using Fourier transform infrared (FT-IR) and gas chromatography-mass spectrometry (GC-MS) techniques. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 127-146.
22. Mohammed GJ, Kadhim MJ, Hussein HM. Characterization of bioactive chemical compounds from *Aspergillus terreus* and evaluation of antibacterial and antifungal activity. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 889-905.
23. Hameed IH, Altameme HJ, Idan SA. *Artemisia annua*: Biochemical products analysis of methanolic aerial parts extract and anti-microbial capacity. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2016; 7(2): 1843- 1868.
24. Hussein AO, Mohammed GJ, Hadi MY, Hameed IH. Phytochemical screening of methanolic dried galls extract of *Quercus infectoria* using gas chromatography-mass spectrometry (GC-MS) and Fourier transform-infrared (FT-IR). *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 49-59.
25. Sosa AA, Bagi SH, Hameed IH. Analysis of bioactive chemical compounds of *Euphorbia lathyris* using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(5): 109-126.
26. Altameme HJ, Hadi MY, Hameed IH. Phytochemical analysis of *Urtica dioica* leaves by fourier-transform infrared spectroscopy and gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(10): 238-252.
27. Mohammed GJ, Omran AM, Hussein HM. Antibacterial and Phytochemical Analysis of *Piper nigrum* using Gas Chromatography-Mass Spectrum and Fourier-Transform Infrared Spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 977-996.
28. Hamza LF, Kamal SA, Hameed IH. Determination of metabolites products by *Penicillium expansum* and evaluating antimicrobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(9): 194-220.
29. Jasin H, Hussein AO, Hameed IH, Kareem MA. Characterization of alkaloid constitution and evaluation of antimicrobial activity of *Solanum nigrum* using gas chromatography mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(4): 56-72.
30. Hadi MY, Mohammed GJ, Hameed IH. Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 8-24.
31. Hameed IH, Ibraheam IA, Kadhim HJ. Gas chromatography mass spectrum and fourier-transform infrared spectroscopy analysis of methanolic extract of *Rosmarinus officinalis* leaves. *Journal of Pharmacognosy and Phytotherapy*. 2015; 7 (6): 90-106.
32. Shareef HK, Muhammed HJ, Hussein HM, Hameed IH. Antibacterial effect of ginger (*Zingiber officinale*) roscoe and bioactive chemical analysis using gas chromatography mass spectrum. *Oriental Journal of Chemistry*. 2016; 32(2): 20-40.
33. Al-Jassaci MJ, Mohammed GJ, Hameed IH. Secondary Metabolites Analysis of *Saccharomyces cerevisiae* and Evaluation of Antibacterial Activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(5): 304-315.
34. Mohammed GJ, Al-Jassani MJ, Hameed IH. Antibacterial, Antifungal Activity and Chemical analysis of *Punica grantanum* (Pomegranate peel) using GC-MS and FTIR spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(3): 480-494.
35. Al-Marzoqi AH, Hadi MY, Hameed IH. Determination of metabolites products by *Cassia angustifolia* and evaluate antimicrobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 25-48.
36. Altameme HJ, Hameed IH, Abu-Serag NA. Analysis of bioactive phytochemical compounds of two medicinal plants, *Equisetum arvense* and *Alchemilla vulgaris* seed using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Malays. Appl. Biol*. 2015; 44(4): 47-58.

37. Hameed IH, Hamza LF, Kamal SA. Analysis of bioactive chemical compounds of *Aspergillus niger* by using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Journal of Pharmacognosy and Phytotherapy*. 2015;7(8): 132-163.
38. Hameed IH, Hussein HJ, Kareem MA, Hamad NS. Identification of five newly described bioactive chemical compounds in methanolic extract of *Mentha viridis* by using gas chromatography-mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7 (7): 107-125.
39. Hussein HM, Hameed IH, Ibraheem OA. Antimicrobial Activity and spectral chemical analysis of methanolic leaves extract of *Adiantum Capillus-Veneris* using GC-MS and FT-IR spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(3): 369-385.
40. Hussein HJ, Hadi MY, Hameed IH. Study of chemical composition of *Foeniculum vulgare* using Fourier transform infrared spectrophotometer and gas chromatography - mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 60-89.
41. Kadhim MJ, Mohammed GJ, Hameed IH. In vitro antibacterial, antifungal and phytochemical analysis of methanolic fruit extract of *Cassia fistula*. *Oriental Journal of Chemistry*. 2016; 32(2): 10-30.
42. Altameme HJ, Hameed IH, Idan SA, Hadi MY. Biochemical analysis of *Origanum vulgare* seeds by fourier-transform infrared (FT-IR) spectroscopy and gas chromatography-mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(9): 221-237.
43. Hussein HM. Determination of phytochemical composition and ten elements content (CD, CA, CR, CO, FE, PB, MG, MN, NI AND ZN) of *CARDARIA DRABA* by GC-MS, FT-IR and AAS technique. *Int. J Pharm Bio Sci*. 2016; 7(3): (B) 1009 –1017.
44. Hussein HM. Analysis of trace heavy metals and volatile chemical compounds of *Lepidium sativum* using atomic absorption spectroscopy, gas chromatography-mass spectrometric and fourier-transform infrared spectroscopy. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2016; 7(4): 2529 – 2555.
45. Hameed IH. A new polymorphic positions discovered in mitochondrial DNA hypervariable region HVIII from central and north-central of Iraq. *Mitochondrial DNA*. 2016; 27(5): 3250-4.
46. Jaddoa HH, Hameed IH, Mohammed GJ. Analysis of volatile metabolites released by *Staphylococcus aureus* using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Orient J Chem*. 2016; 32(4).
47. Hameed IH, Salman HD, Mohammed GJ. Evaluation of antifungal and antibacterial activity and analysis of bioactive phytochemical compounds of *Cinnamomum zeylanicum* (Cinnamon bark) using gas chromatography-mass spectrometry. *Orient J Chem*. 2016; 32(4).
48. Hameed IH, Jebor MA, Ommer AJ, Abdulzahra AI. Haplotype data of mitochondrial DNA coding region encompassing nucleotide positions 11,719–12,184 and evaluate the importance of these positions for forensic genetic purposes in Iraq. *Mitochondrial DNA*. 2016; 27(2): 1324-1327.
49. Kadhim MJ, Mohammed GJ, Hussein HM. Analysis of bioactive metabolites from *Candida albicans* using (GC-MS) and evaluation of antibacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
50. Mohammad A, Imad H. Autosomal STR: From locus information to next generation sequencing technology. *Research Journal of Biotechnology*. 2013.
51. Hameed, I.H., Abdulzahra, A.I., Jebor, M.A., Kqueen, C.Y., Ommer, A.J. Haplotypes and variable position detection in the mitochondrial DNA coding region encompassing nucleotide positions 10,716-11,184. *Mitochondrial DNA*. 2015.
52. Ubaid JM, Hussein HM, Hameed IH. Analysis of bioactive compounds of *Tribolium castaneum* and evaluation of anti-bacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
53. Altaee N, Kadhim MJ, Hameed IH. Detection of volatile compounds produced by *Pseudomonas aeruginosa* isolated from UTI patients by gas chromatography-mass spectrometry. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
54. Altaee N, Kadhim MJ, Hameed IH. Characterization of metabolites produced by *E. coli* and analysis of its chemical compounds using GC-MS. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
55. Hussein JH, Ubaid JM, Hameed IH. Gas chromatography – mass spectrum analysis of volatile components of methanolic leaves extract of *Cordia myxa*. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
56. Kadhim MJ, Kaizal AF, Hameed IH. Medicinal plants used for treatment of rheumatoid arthritis: A review. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
57. Hameed, I.H., Al-Rubaye A.F. and Kadhim, M.J. Antimicrobial Activity of Medicinal Plants and Urinary Tract Infections. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
58. Kadhim WA, Kadhim, M.J., Hameed, I.H. Antibacterial Activity of Several Plant Extracts Against *Proteus Species*. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
59. Kadhim MJ. In Vitro antifungal potential of *Acinetobacter baumannii* and determination of its chemical composition by gas chromatography-mass spectrometry. *Der Pharma Chemica*, 2016; 8(19): 657-665.
60. Al-Yaseri A, Kadhim WA, Hameed IH. Detection of volatile compounds emitted by *Proteus mirabilis* isolated from UTI patients and its anti-fungal

- potential. *Der Pharma Chemica*, 2016; 8(19): 671-678.
61. Ubaid JM, Kadhim MJ, Hameed IH. Study of bioactive methanolic extract of *Camponotus fellah* using Gas chromatography – mass spectrum. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
62. Esteves PF, Sato A, Esquibel M.A, Buzzi F C, Meira AV, Filho, VC. Antinociceptive Activity of *Malva sylvestris* L. *Lat. Am. J. Pharm.* 2009; 28 (3): 454-6.
63. Guarrera PM. Traditional phytotherapy in central Italy (Marche, Abruzzo, and Latium). *Fitoterapia*. 2005; 76: 1-25.
64. Calzada F, Meckes M, Cedillo-Rivera R. Antiamoebic and anti-giardial activity of plant flavonoids. *Planta medica*. 1999; 65: 78-80.
65. Saleem A, Husheem M, Harkonen P, Pihlaja K. Inhibition of cancer cell growth by crude extract and the phenolics of *Terminalia*. *Fruit. J. of Ethnopharmacology* .2002; 81, 327–36.
66. Chen CC, Liu LK, Hsu JD, Huang HP, Yang MY, Wang CJ. Mulberry extract inhibits the development of atherosclerosis in cholesterol fed rabbits. *Food Chemistry* .2005. 91, 601–7.
67. Calliste CA, Trouillas P, Allais DP, Simon A, Duroux, JL. Free radical scavenging activities measured by electron spin resonance spectroscopy and B16 cell antiproliferative behaviors of seven plants. *Journal of Agricultural and Food Chemistry*. 2001; 49: 3321-27.
68. Shirwaikar A.A., Prabu L.S., Mahalaxmi R., Rajendran K., Studies of disintegrant properties of seed mucilage of *Ocimum gratissimum*, *Ind. J. of Pharma. Sci.*, Dec. 2007, 753-758.
69. Shetty A.J., Alwar M.C., Shyamjith., Acute toxicity studies and determination of median lethal dose, *Current science*, 2007, 93, 917-920.
70. Jerald E., Dosi S., Raj A., Application of hibiscus leaves mucilage as suspending agent, *Ind. J. Pharm. Edu. Res.*, 2007, 41(4).
71. Mann A.S., Jain N.K., Khrya M.D., Evaluation of suspending properties of *Cassia tora* mucilage on sulphadimide suspension, *Asian J. Exp. Sci.*, 2007, 21(1), 63-67.