

# Staining

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## **Types of staining methods:**

**Simple staining** : is composed of one type of stain e.g, crystal violate, safranine. methylene blue.

**Differential staining** : It gives different color to different bacteria e.g.

Gram stain, and Ziehl neelsen stain.

**Special staining** : like Capsule stain , flagella , spores



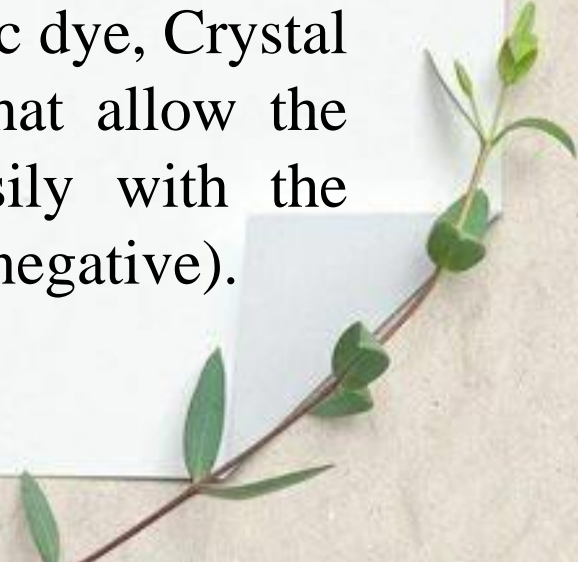


# Differential staining

## A:Gram Stain

### Introduction

First devised by Hans Christian Gram during the late nineteenth century, the gram stain can be used to divide most bacterial species effectively into two large groups: those that take up the basic dye, Crystal violet (gram – positive), and those that allow the crystal violet dye to wash out easily with the decolorizer alcohol or acetone (gram – negative).





# Reagents Used In Gram Stain

1. Gram **Crystal Violet**
2. Gram **Iodine**
3. Gram **Decolorizer**
  - a. Methanol
  - b. Acetone
4. Gram **Safranin**



## Cont...

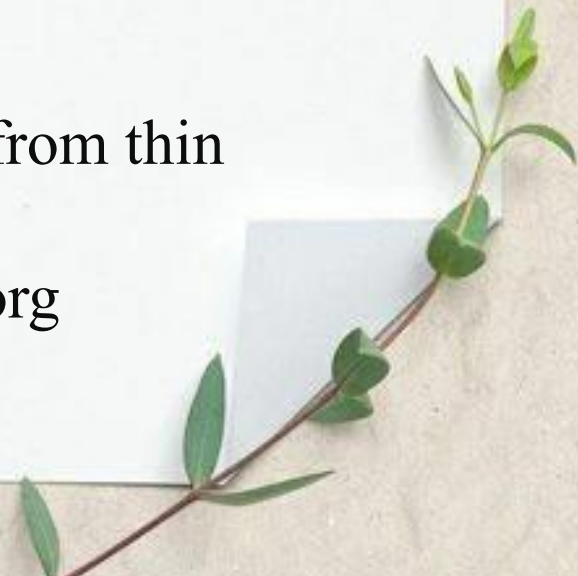
### 1. CRYSTAL VIOLET

- **Primary** stain
- **Violet** colored, stains all micro-org

### 2. GRAM **IODINE**

- **Mordant**
- Forms Crystal **violet iodine** complexes

### 3. DECOLORIZER

- Acetone + Methanol
  - Removes Crystal violet iodine complex from thin peptidoglycan layers
  - Dissolves outer layer of Gram negative org
- 



## 4. GRAM **SAFRANINE**

- **Counter** stain
- **Red** colored
- Stains thin walled Gram negative  
org



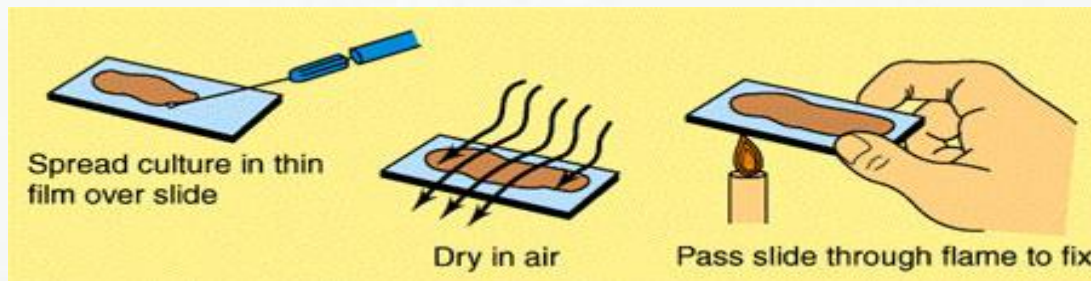
**Gram Staining kit**

# The Gram Stain Procedure

## Step 1-Prepare a Smear

### “Bacteria”

Suspend some of the material to be stained in a drop of water on a microscope slide ,Then spread the drop . Fix by Heat or Allow to air dry.



## Step 2-Apply the Primary Stain

Flood the Smear with Crystal Violet

Allow to stand for 1 min

Rinse with water to remove excess stain





### **Step 3-Apply the Mordant**


Flood the Smear with Iodine solution  
Allow to stand 2 min


### **Step 4 -Rinse**

Rinse with water to remove excess Iodine

### **Step 5 -Decolorize**

Decolorizer by alcohol  
across the slide about 5 sec  
The slide should appear clear





**Step 6 - Rinse**

Rinse with water to remove excess alcohol

**Step 7 - Counter stain**

Flood the slide with Safranin solution

Let stand for 2 minutes

**Step 8 - Rinse , Dry and Observe**

**Gram-Positive Gram-Negative**

Rinse with water to remove excess stain

Blot dry

Observe under Oil Immersion





**1** Application of crystal violet (purple dye)



**2** Application of iodine (mordant)



**3** Alcohol wash (decolorization)



**4** Application of safranin (counterstain)

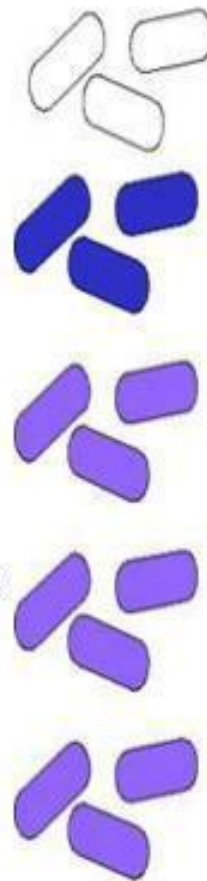
KEY	
	Crystal violet
	Iodine
	Alcohol
	Safranin

# Gram Stain

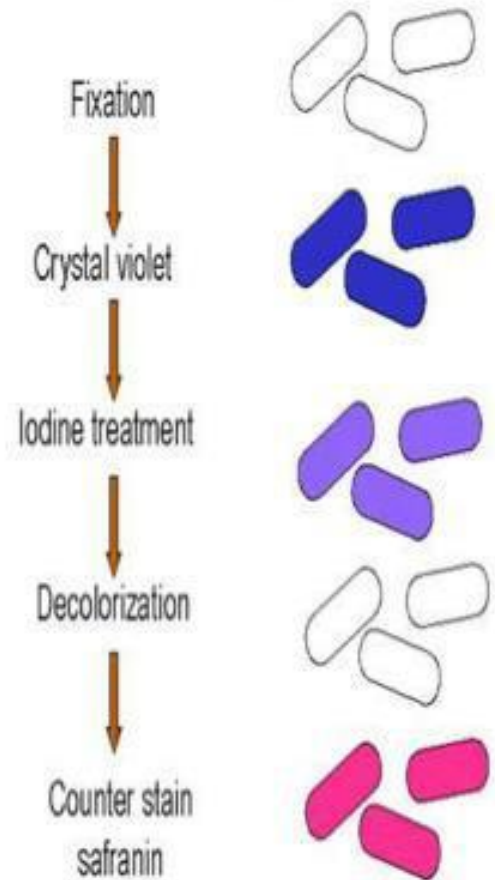
## Principle of staining technique:

1. Primary stain:- Crystal Violet
2. Mordant(fixes the dye):- Iodine
3. Decolorizing agent:- Alcohol/Acetone
4. Counter stain;- Safranin

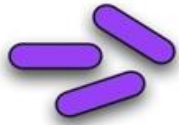
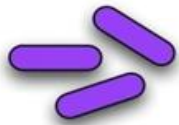
Gram Positive



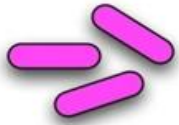
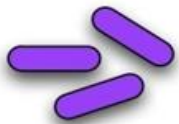
Gram Negative



GRAM-POSITIVE



GRAM-NEGATIVE



Fixation



Crystal Violet



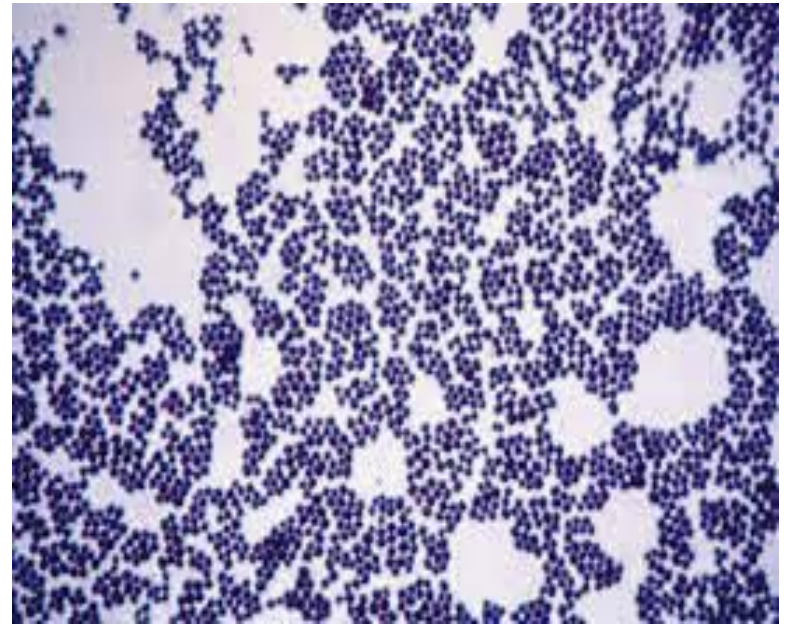
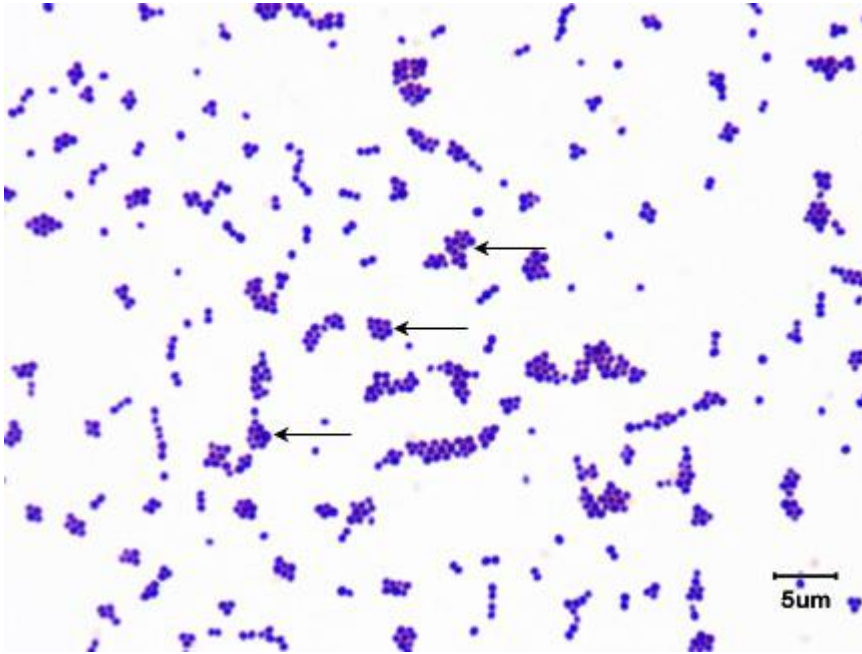
Iodine Treatment



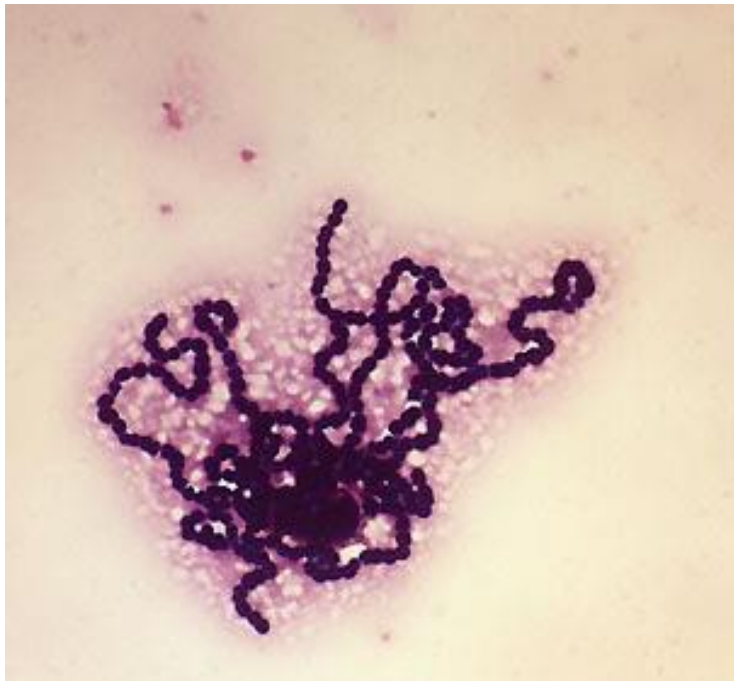
Decolorisation



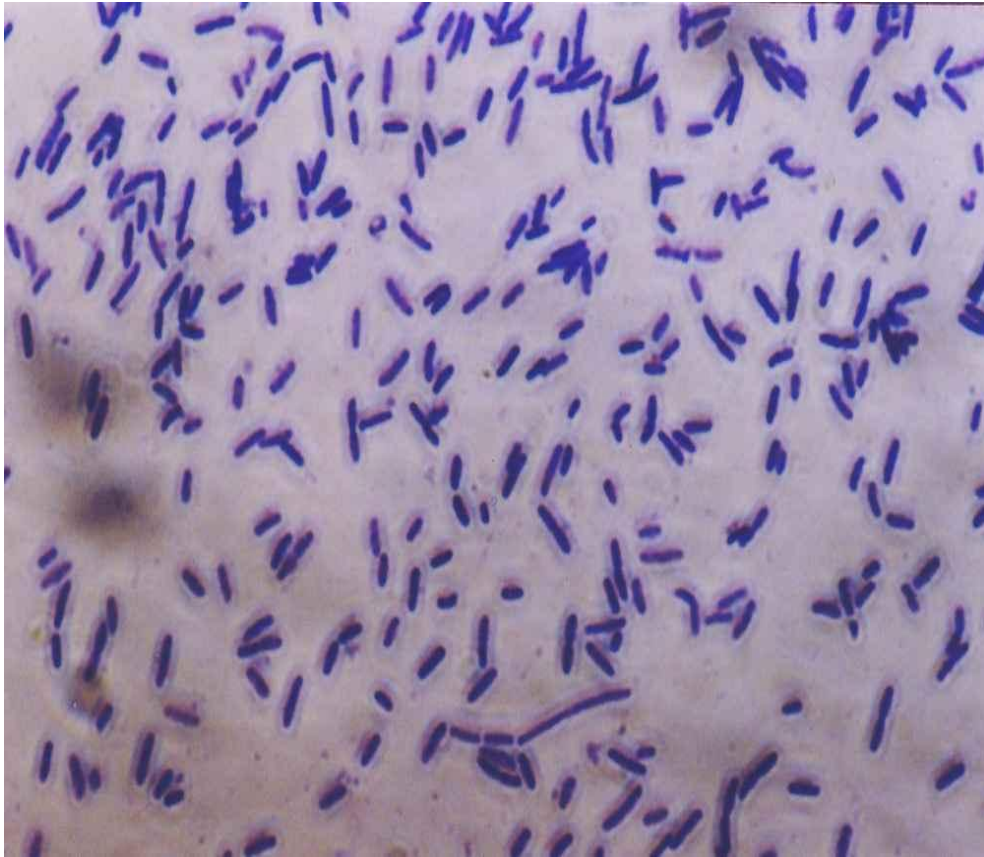
Counter stain with Safranin



**Gram Positive  
Staphylococci**

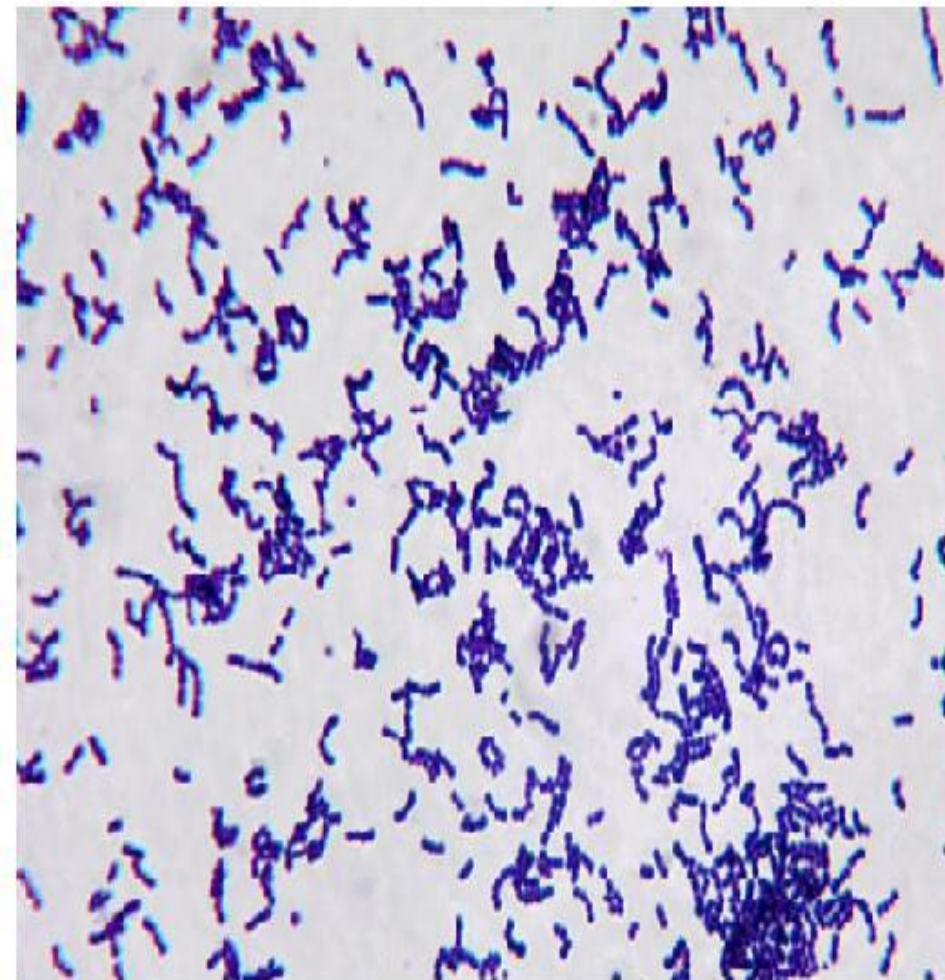


**Gram Positive**  
*(Streptococci)*

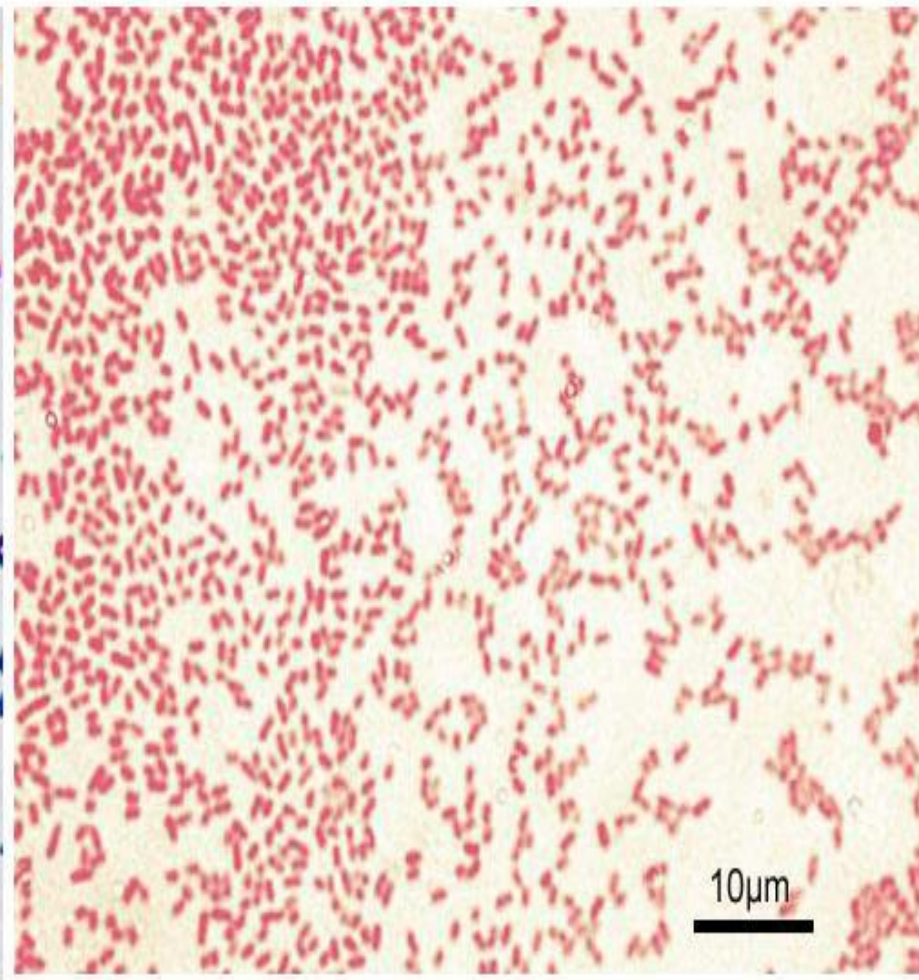


**Gram Positive  
Bacilli**





Gram Positive Bacteria



Gram Negative Bacteria

# SHAPES OF BACTERIA

## COCCI

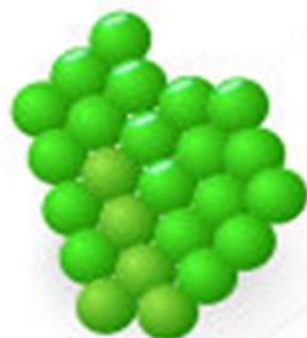


**Diplococci**  
(*Streptococcus pneumoniae*)



**Streptococci**  
(*Streptococcus pyogenes*)

### **Tetrad**



**Staphylococci**  
(*Staphylococcus aureus*)



**Sarcina**  
(*Sarcina ventriculi*)

## BACILLI



**Chain of bacilli**  
(*Bacillus anthracis*)



**Flagellate rods**  
(*Salmonella typhi*)



**Spore-former**  
(*Clostridium botulinum*)

## OTHERS



**Vibrios**  
(*Vibrio cholerae*)



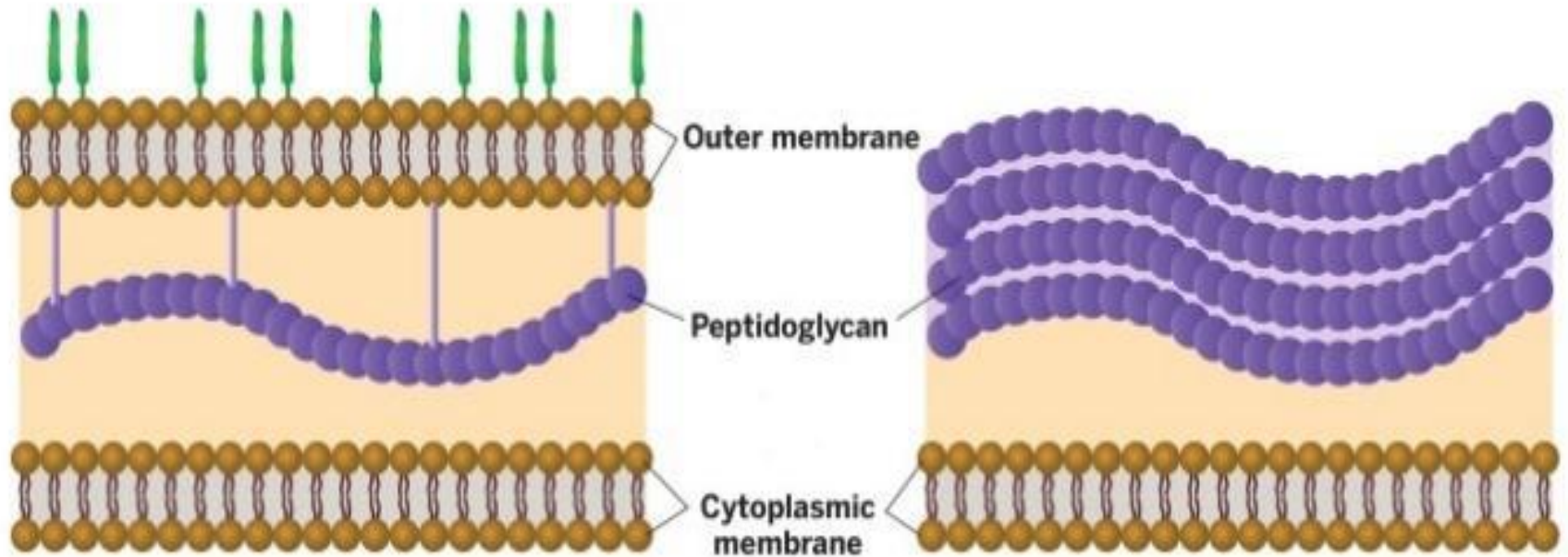
**Spirilla**  
(*Helicobacter pylori*)



**Spirochaetes**  
(*Treponema pallidum*)

# GRAM-NEGATIVE

# GRAM-POSITIVE



## **CELL WALL IN GRAM +VE AND GRAM -VE BACTERIA**

<b>Cell Wall Structures</b>	<b>Gram Positive organisms</b>	<b>Gram Negative organisms</b>
Inner cytoplasmic membrane	Present	Present
Peptidoglycan layer	Thick	Thin
Teichoic Acid	Present	Absent
Outer membrane layer	Absent	Present
Lipid A, LPS , Lipo-protien components	Absent	Present
Peri-plasmic space	Absent	Present



## **Errors in Gram stain technique:-**

- 1. Over decolorization and conc. of alcohol**
- 2. Old culture**
- 3. Thick smear**
- 4. More exposure to heat (during the fixation of smear)**



**B- Acid fast stain(Ziehl- Neelsen stain ) :-**

Most bacteria in the genus *Mycobacterium* contain **very high percentage lipid in their cell wall.**

This stain is used in the identification of the tuberculosis bacillus , ***Mycobacterium tuberculosis*** , and the leprosy organism , ***Mycobacterium leprae*** .





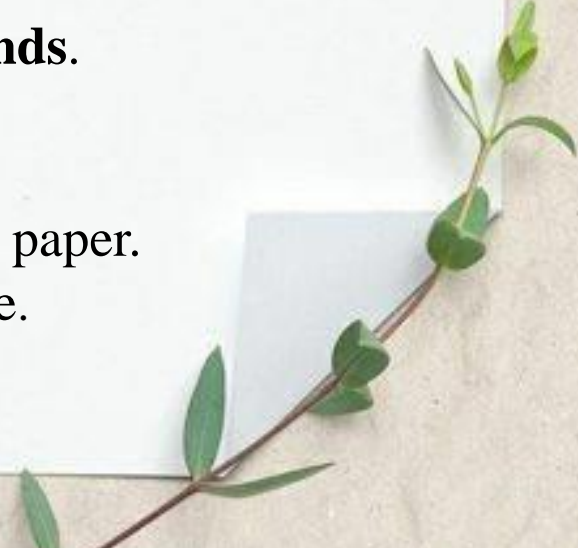
## The Ziehl-Neelsen stain composed of

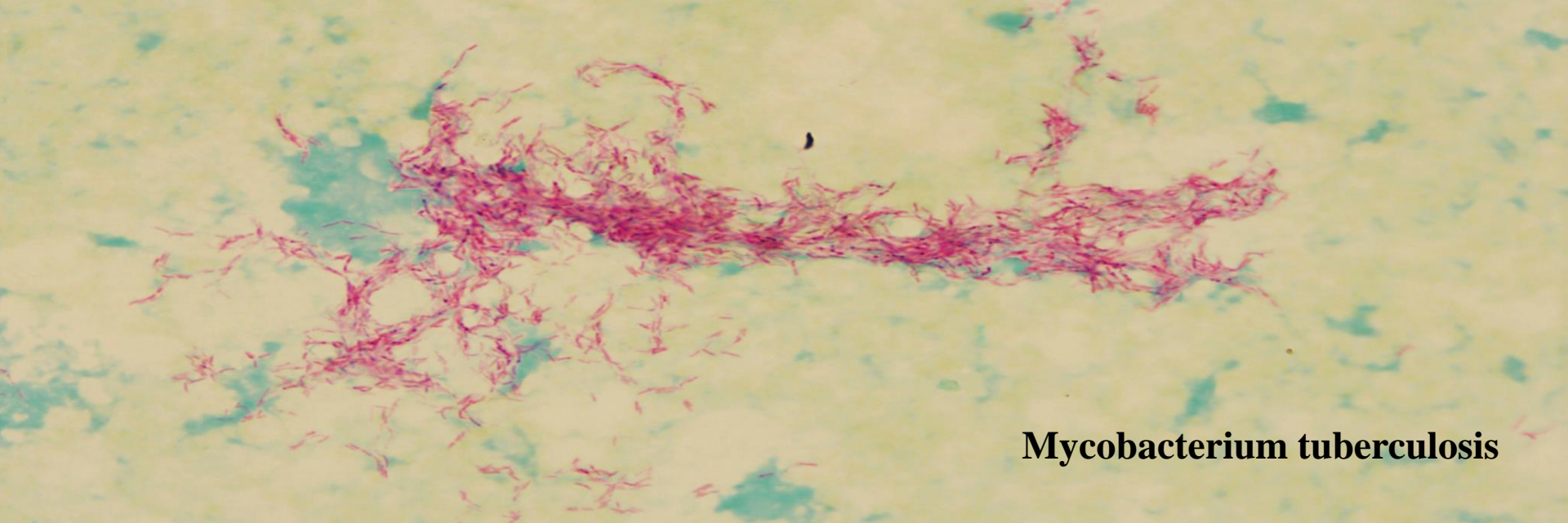
Carbol fuchsin solution (**red color**).

Ethanol 95% acidified with 20% HCL or H<sub>2</sub>SO<sub>4</sub>.

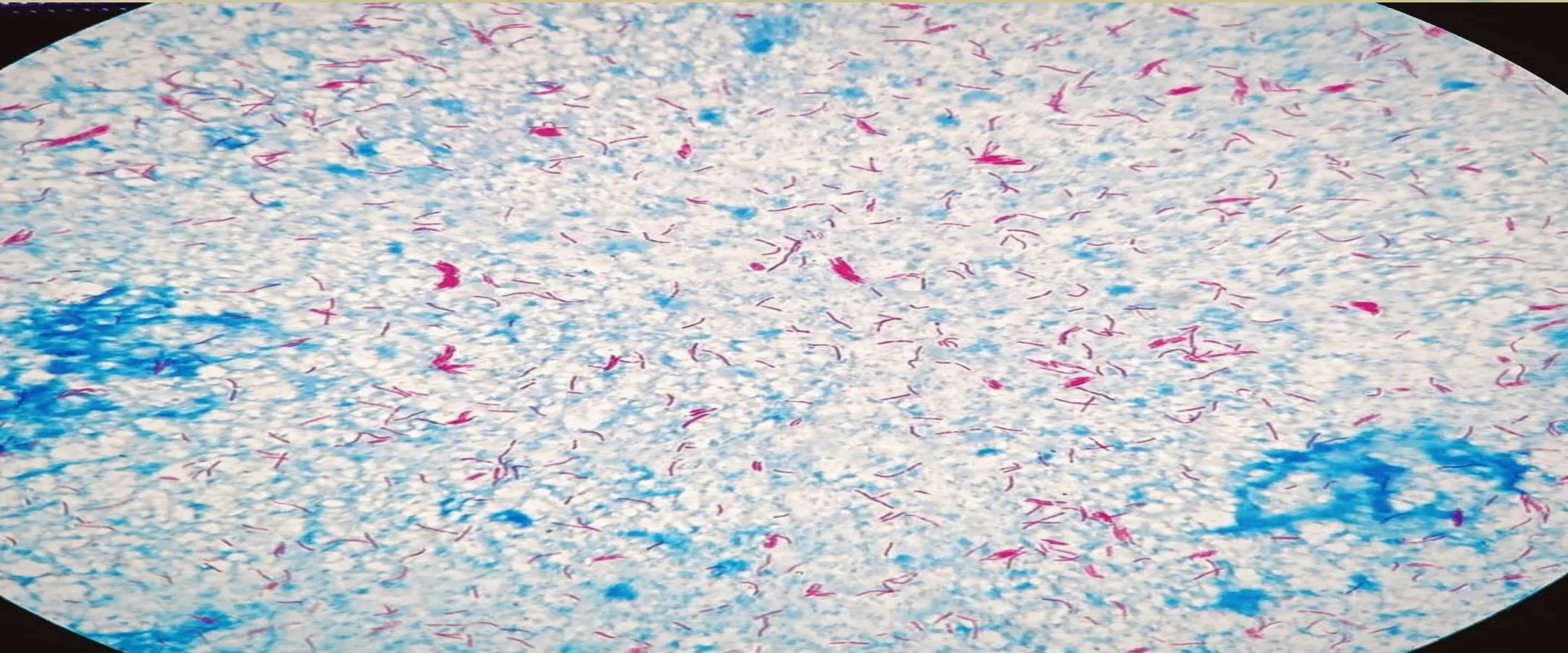
**Methylene blue** solution

### Procedure:-

- 1-Prepare microbial **smear (the sputum ) specimen** & fixed it.
  - 2-Flood the slide with **carbol fuchsin** stain then heat it on the Benson burner for 5minutes.
  - 3-Wash the slide with water.
  - 4-Decolorize with the **acid- alcohol for 10-20seconds**.
  - 5-Wash the slide with water.
  - 6-Add **methylene blue** for 30 seconds .
  - 7-Wash the slide with water & dry it with bibulous paper.
  - 8-Examine the prepared slide under the microscope.
- 



**Mycobacterium tuberculosis**







Thank  
you!

The image features a central hexagonal frame with a black outer border and a gold inner border. The frame is surrounded by a lush arrangement of green leaves and branches, including some fern-like leaves on the right side. The text 'Thank you!' is written in a black, elegant cursive font in the center of the frame. A faint 'dreamstime.' watermark is visible behind the text.